

Supplementary material

Phosphorylation of the mitochondrial autophagy receptor Nix enhances its interaction with LC3 proteins

Vladimir V. Rogov^{1,*}, Hironori Suzuki^{2,3,*}, Mija Marinković⁴, Verena Lang⁵, Ryuichi Kato³, Masato Kawasaki³, Maja Buljubašić⁴, Matilda Šprung⁶, Natalia Rogova¹, Soichi Wakatsuki⁷, Anne Hamacher-Brady^{5,8}, Volker Dötsch^{1,¶}, Ivan Dikić^{4,9}, Nathan R. Brady^{8,10} and Ivana Novak^{4,¶}

Supplementary material figure legends

Figure S1. Characterization of the Nix phosphorylation effect on LC3B interaction by ITC and NMR. (a) K_D values calculated from LC3B CSP upon titration with P0 based on previous data (left plot) and current data (right plot). (b) Overlaid sections of [¹⁵N, ¹H]-TROSY HSQC spectra indicating exchange modes of selected LC3B resonances upon titration with different LIR peptides. Rainbow color-code represents molar ratio LC3B:peptide from 0 (red) to 8 (violet).

Figure S2. Detailed structure of Nix LIR and LC3B interaction. (a) The electron density map of Nix LIR of molecule A. Atoms are shown and colored similarly to Figure 3. Electron density maps of 2Fo-Fc at 1 σ are shown in gray wire frame representation. The N-terminus of LIR (PPAGL) could not be assigned due to weak electron density. (b) The interaction of Glu34 in the Nix LIR (magenta) with the Arg10, Arg11, Lys51 of LC3B (green). The distances (Å) between atoms are shown in dotted black lines. Glu34 forms salt bridges with Lys51 and Arg10. However, the distance between the carboxyl Glu34 and Arg11 is longer, requiring a rotamer change in Arg11 to facilitate an interaction.

Figure S3. Lysine residue 49 in LC3B increases interaction and colocalization with Bnip3. (a) Co-immunoprecipitation of EGFP-LC3B wild type and K49A together with pmCherry-Bnip3-S17,24E, pmCherry-Nix-S34,35E or tRFP-p62 was performed 24 hrs post transfection using α -GFP. GFP, RFP and tRFP were detected via Western blotting and representative blot is shown. Statistical significance was tested vs LC3B wt using Students one-tailed paired t-test for four independent experiments; *, $p \leq 0.05$, **, $p \leq 0.01$. (b) Colocalization of EGFP-LC3B wild type and Bnip3. Scale bar 10 μ m, scale bar for zoom images 5 μ m.

Figure S4. Mitochondrial colocalization of Nix wild type and mutants with LC3B. (a) Colocalization of Nix mutants with mitochondrial marker Tom20. EGFP-Nix wild

type, Δ LIR, S34,35A, S34,34D and S34,35E (green) show colocalization with endogenous Tom20 (red). (b) Colocalization of Nix mutants with LC3B upon treatment with CCCP. EGFP-Nix wild type, Δ LIR, S34,35A, S34,34D and S34,35E (green) and LC3B (red).

Figure S5. (a) LC3B Arginine 10 and Arginine 11 are necessary for interaction with Nix. Immunoprecipitation of EGFP-LC3B wild type and R10A or R11A together with pmCherry-Nix wild type, S34,35A and S34,35E mutant or tRFP-p62. Immunoprecipitation was performed 24 hrs post transfection using α -GFP. GFP, RFP and tRFP were detected via Western blotting and representative blot is shown. LC3B R10A and R11A do not coimmunoprecipitate with Nix wild type and mutants; similarly, LC3B R10A and R11A weakly coimmunoprecipitates with p62. (b) Colocalization profiles for Figure 4a, 4d and 4e. The area of plot profiles is shown in zoom images (Figure 4a, 4d and 4e) represented by a white line.

Figure S6. Analysis of mitochondrial removal determined by flow cytometry using GFP-Nix, Δ LIR, S34,35A and S34,35E mutants. (a) Example of flow cytometry parameters used for normalization of the experiments (singlets analysis (left), exclusion of dead cells determined by propidium iodide (middle) and fluorescence intensity (right)). (b) Analysis of fold change between CCCP and CCCP/Bafilomycin A treated cells using GFP-Nix, Δ LIR, S34,35A and S34,35E mutants in flow cytometry. (c) Western blot analysis of cell lysates used in flow cytometry experiment. α -GFP, α -TOM20, α -LC3 and α -actin were used on the representative blot shown.

Figure S7. LC3B Lysine 51 is necessary to recognize phosphorylated Bnip3 LIR. Immunoprecipitation of EGFP-LC3B wild type and K51A together with pmCherry-Bnip3 wild type, 2SA (S17,24A) and -2SE (S17,24E) mutant. Immunoprecipitation was performed 24 hrs post transfection using α -GFP. GFP, RFP and tRFP were detected via Western blotting and representative blot is shown. LC3B K51A does not coimmunoprecipitate with Bnip3 wild type and mutants.

Figure S8. Extended ITC study of the interactions between LC3B variants and different LIR-containing peptides. (a) ITC profiles for the titration of wild type LC3B and LC3B mutants (R11A, K49A and K51A) with phosphorylated Nix LIR (P2) peptide. (b) ITC profiles for the titration of the same LC3B variants with p62 LIR peptide. (c) ITC profiles for the titration of wild type LC3B and LC3B K49A with Nix LIR (P0) peptide (left plot) and with B2L13 LIR peptide (right plot). In each plot the top diagrams display the raw measurements and the bottom diagrams show the integrated heat per titration step.

Figure S1.

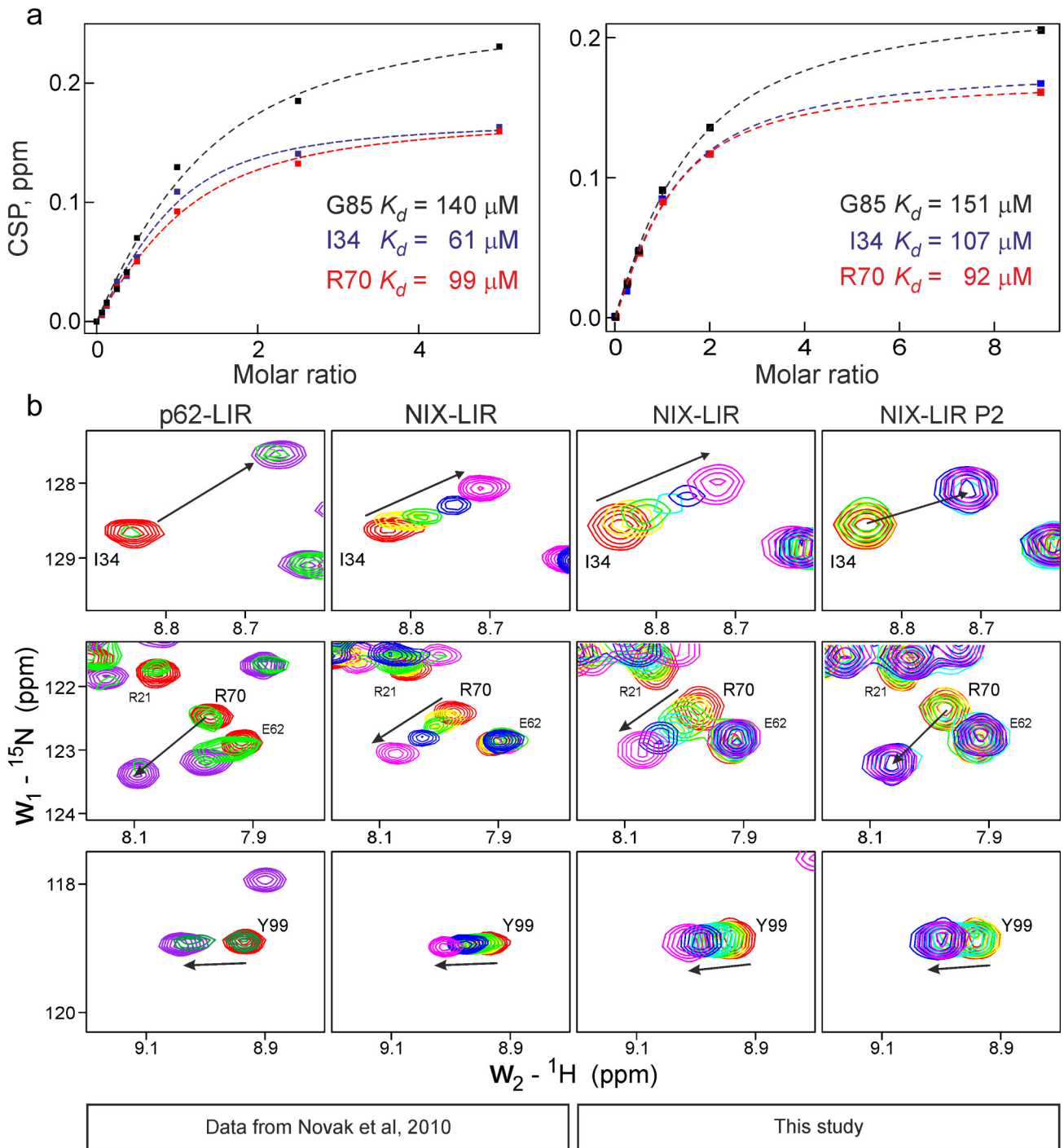
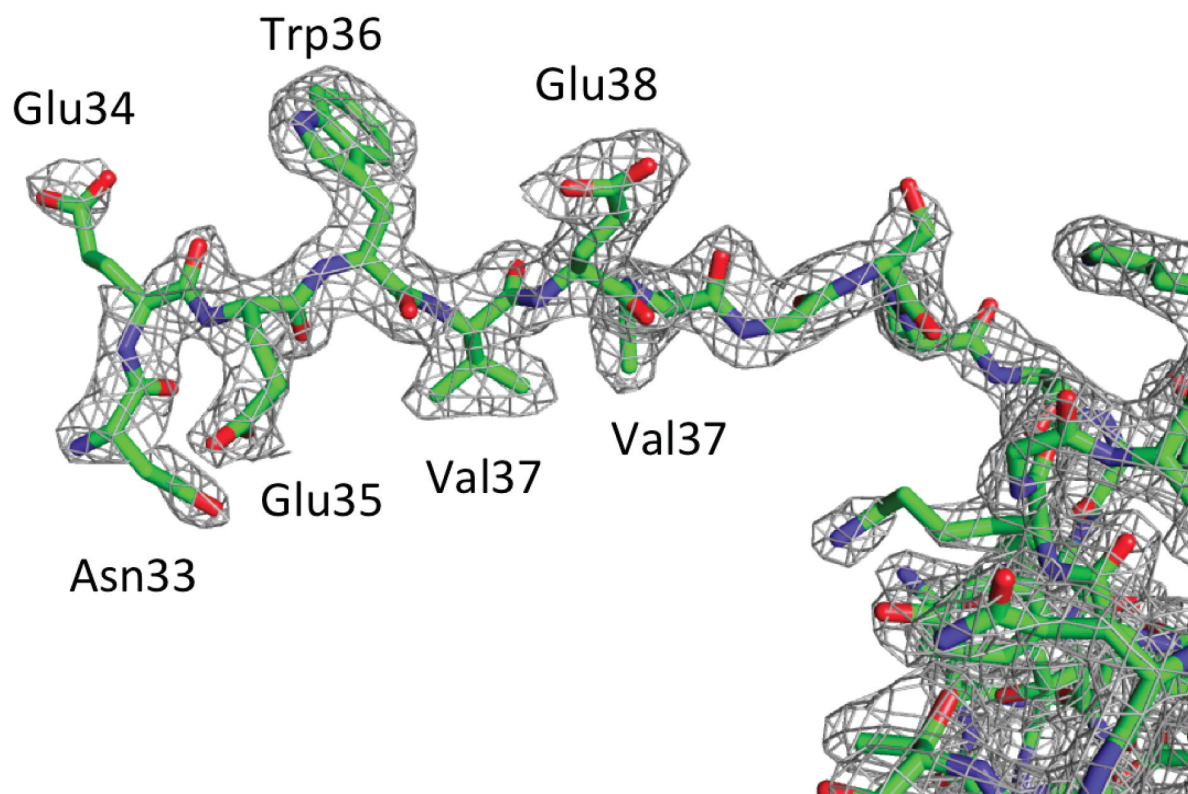


Figure S2.

a



b

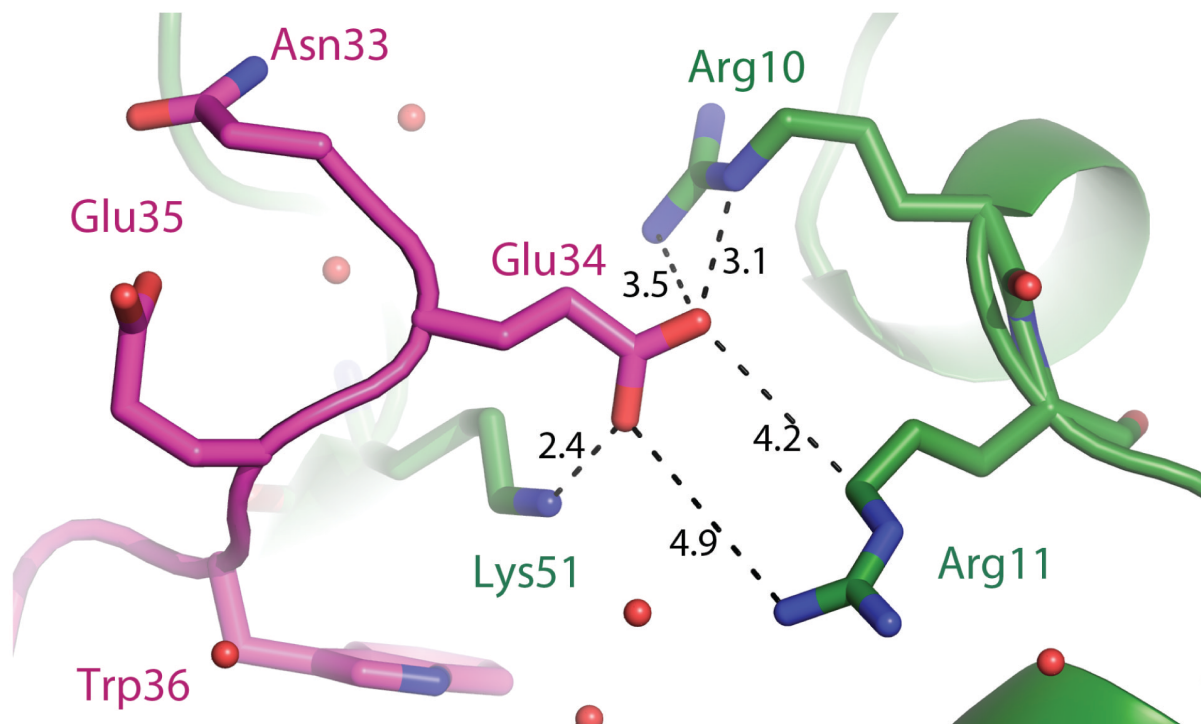
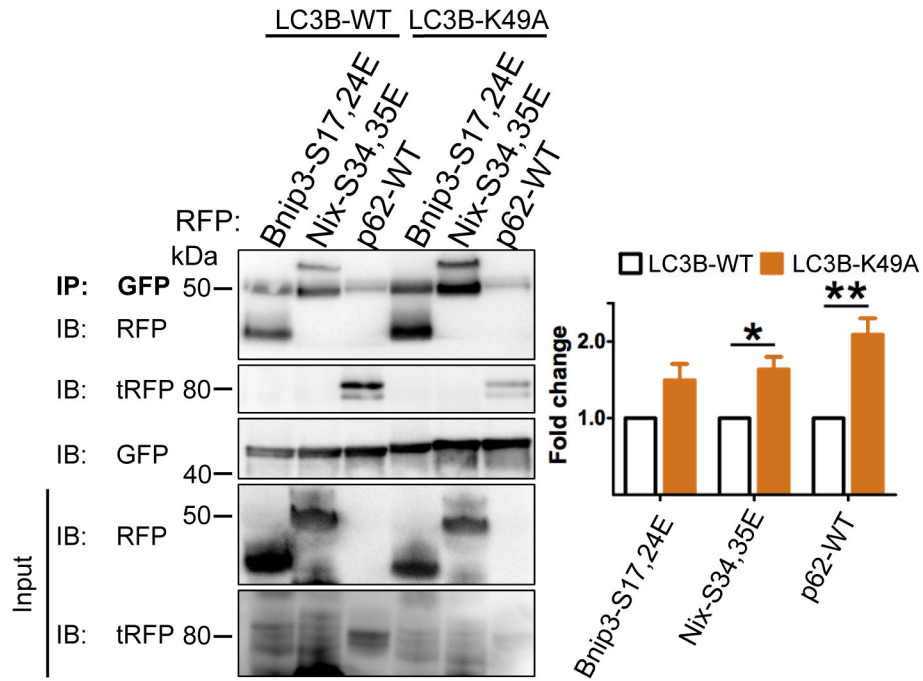


Figure S3.

a.



b.

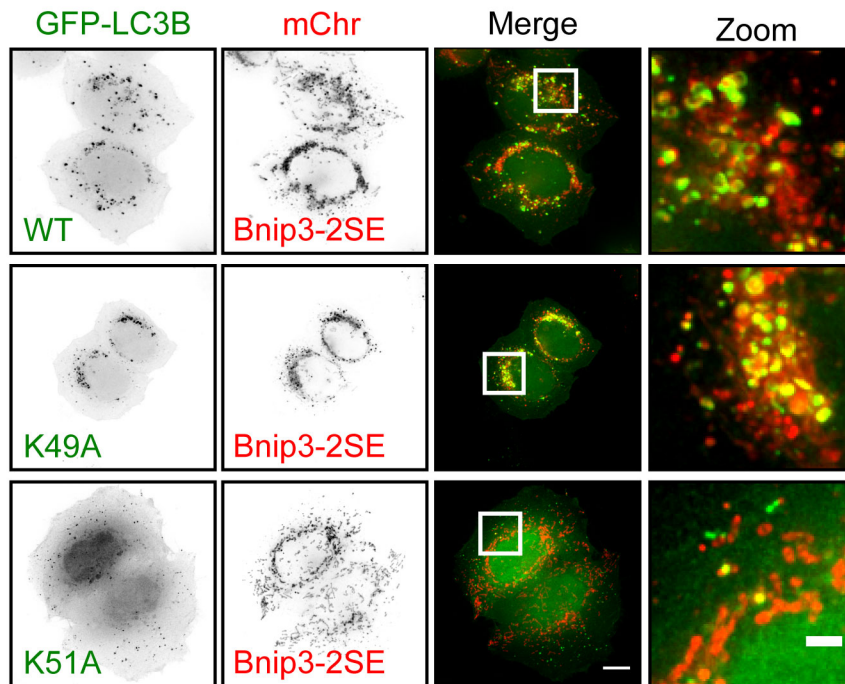


Figure S4.

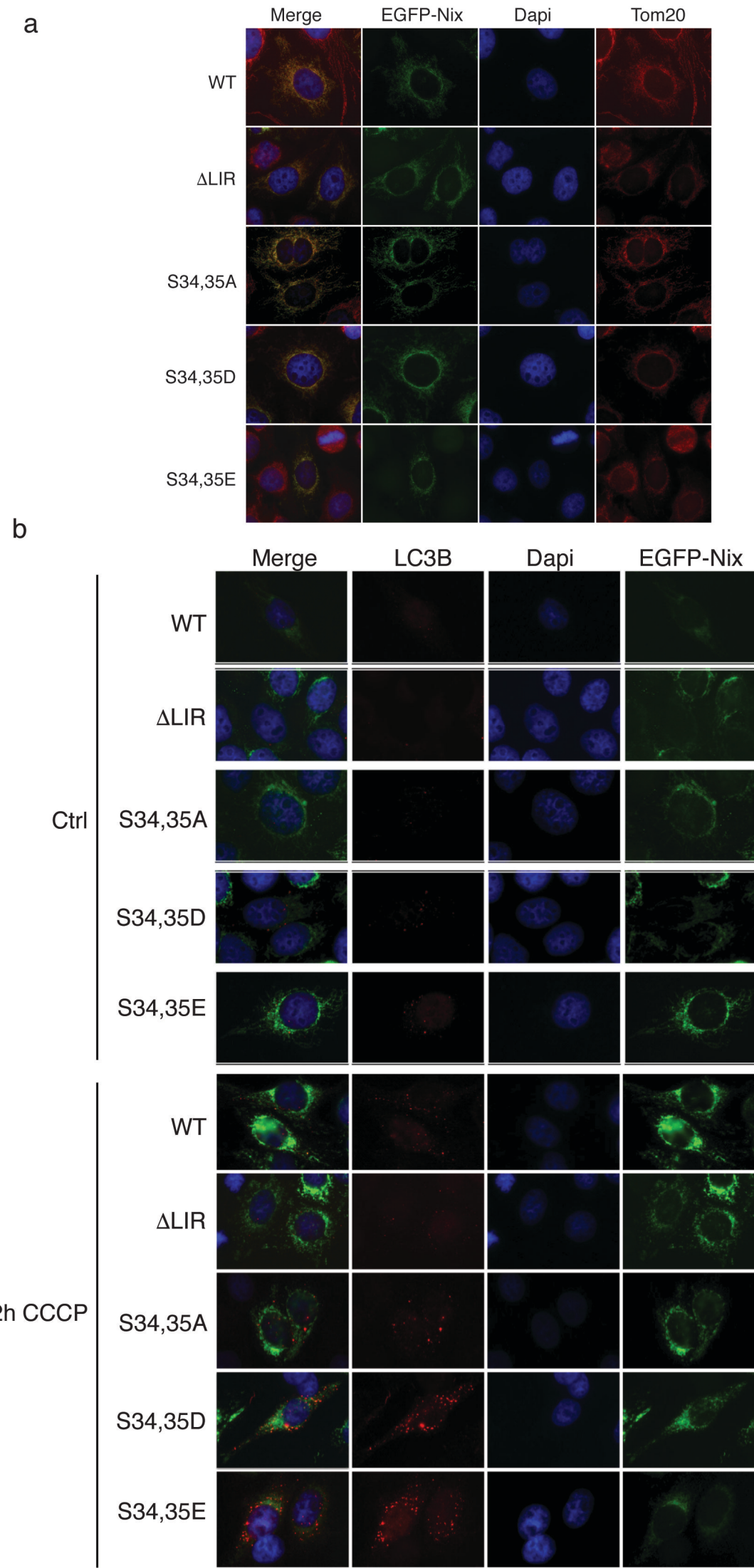


Figure S5.

a

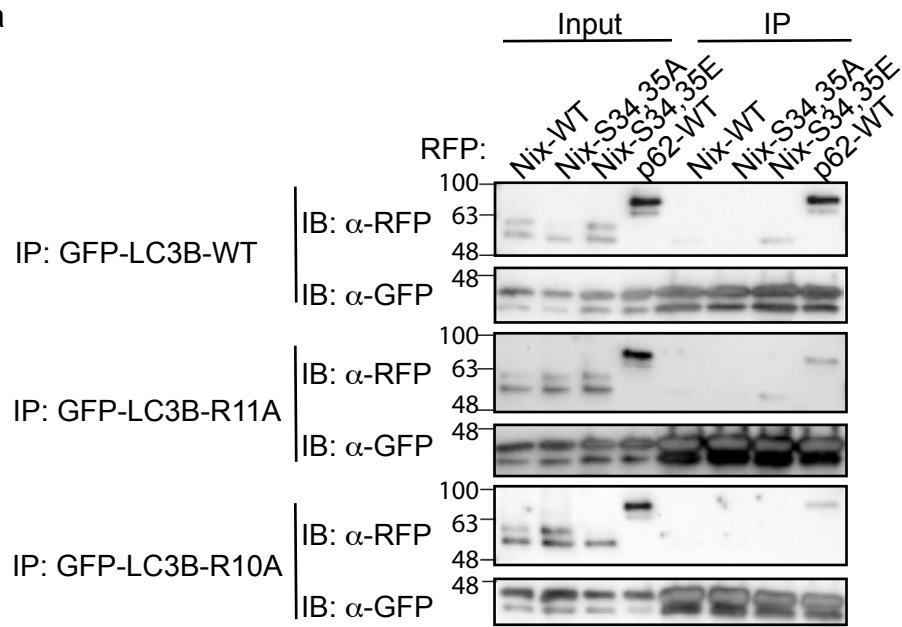


Figure S5.

b

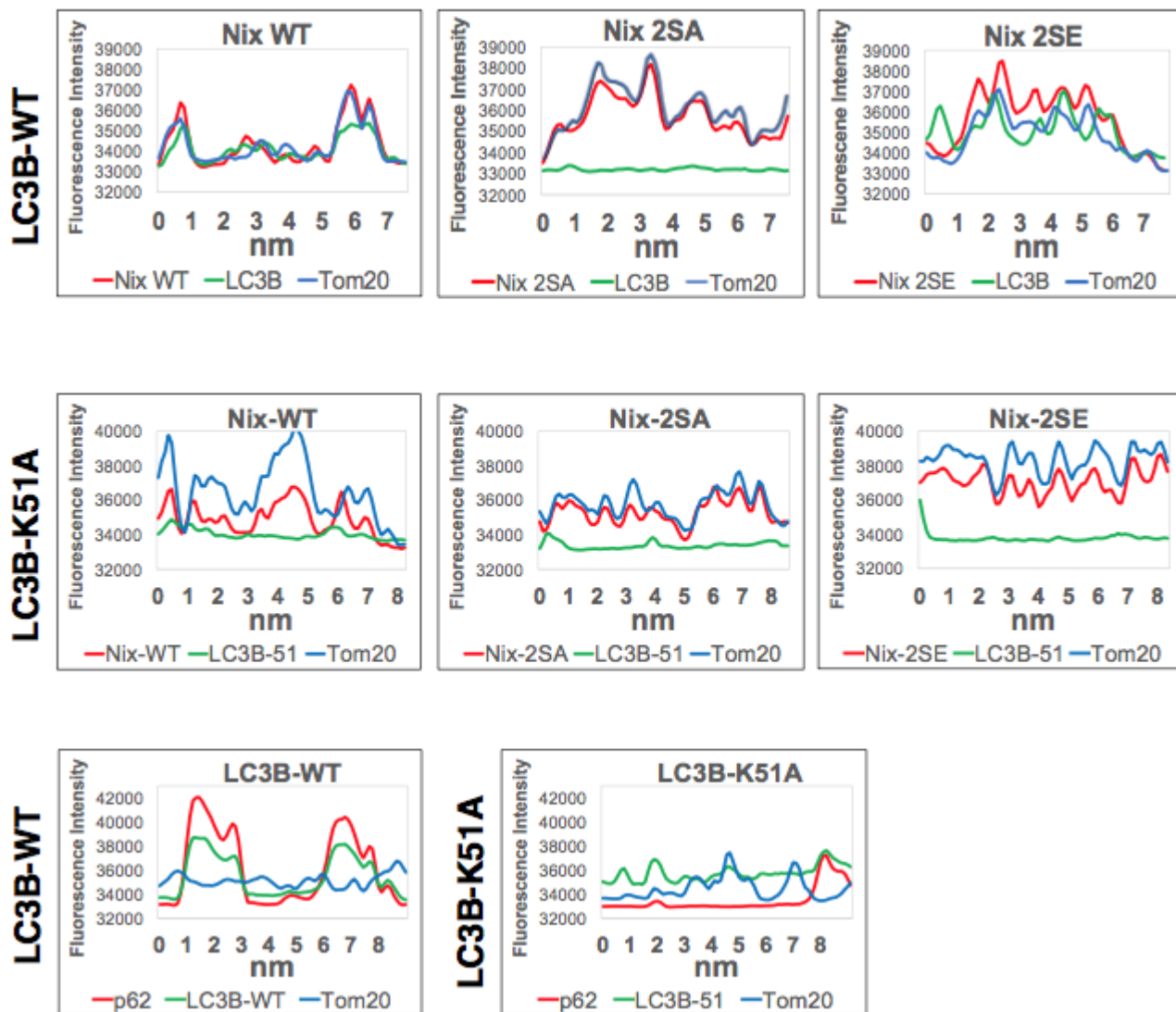
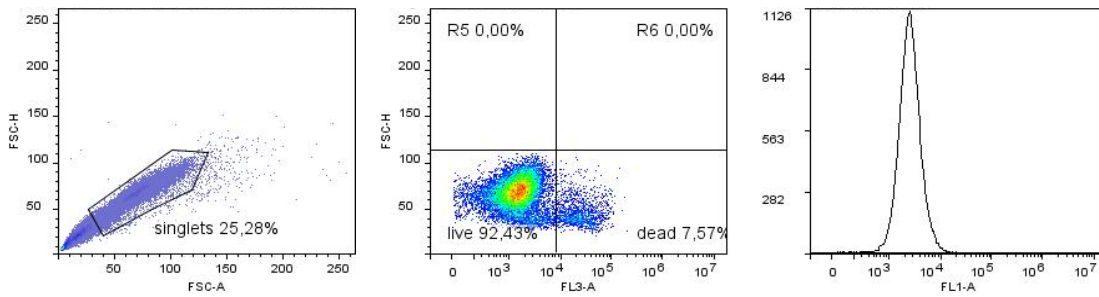
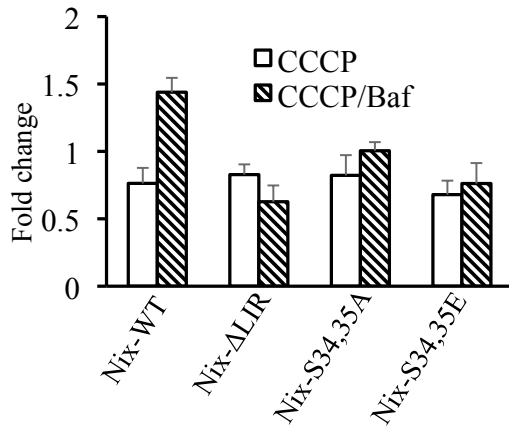


Figure S6.

a



b



c

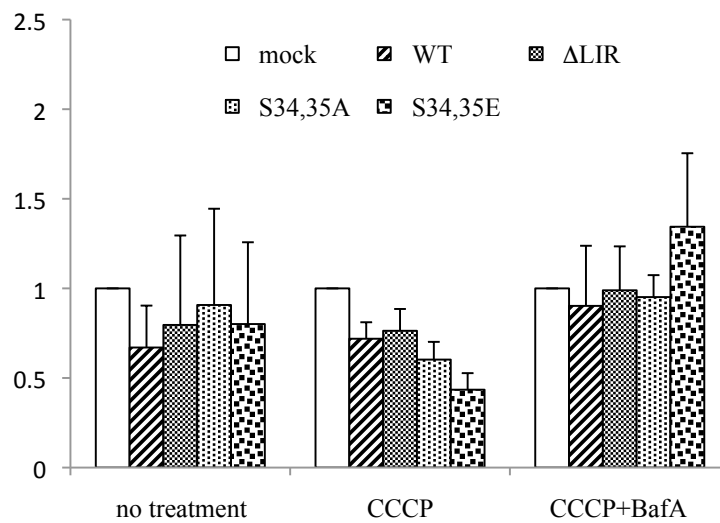
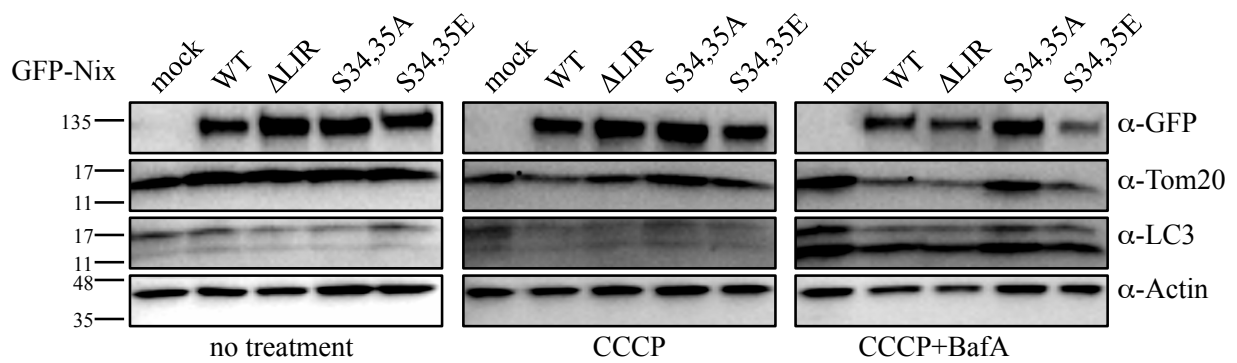


Figure S7.

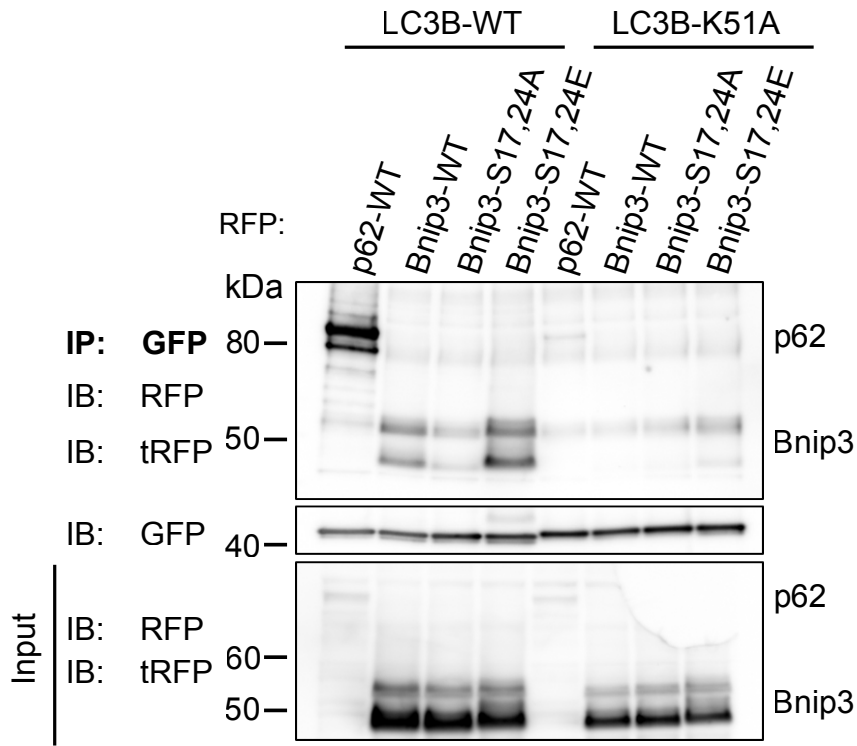


Figure S8.

