

Supplementary information

A Chemical Toolbox for the Study of Bromodomains and Epigenetic Signaling

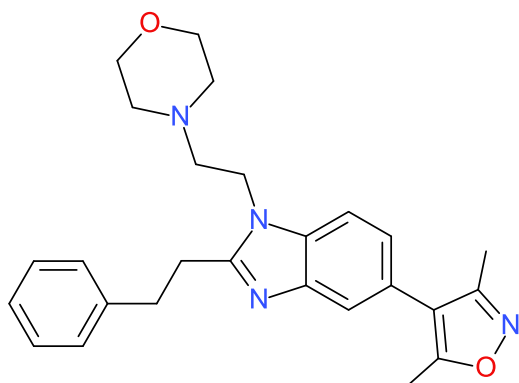
Knapp and Arrowsmith et al.

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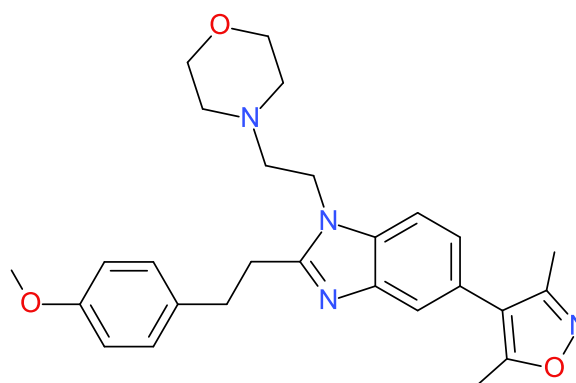
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Supplementary figure 1

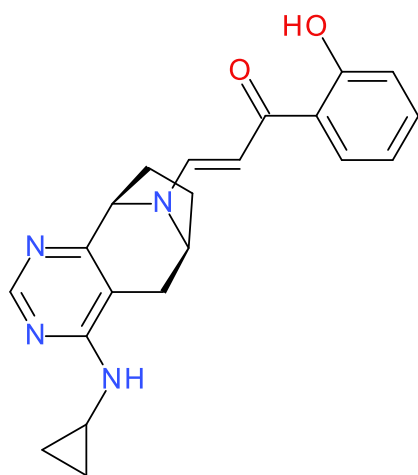
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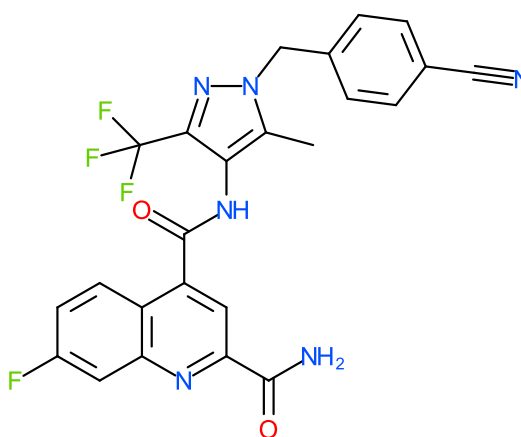
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PFI-3 D1

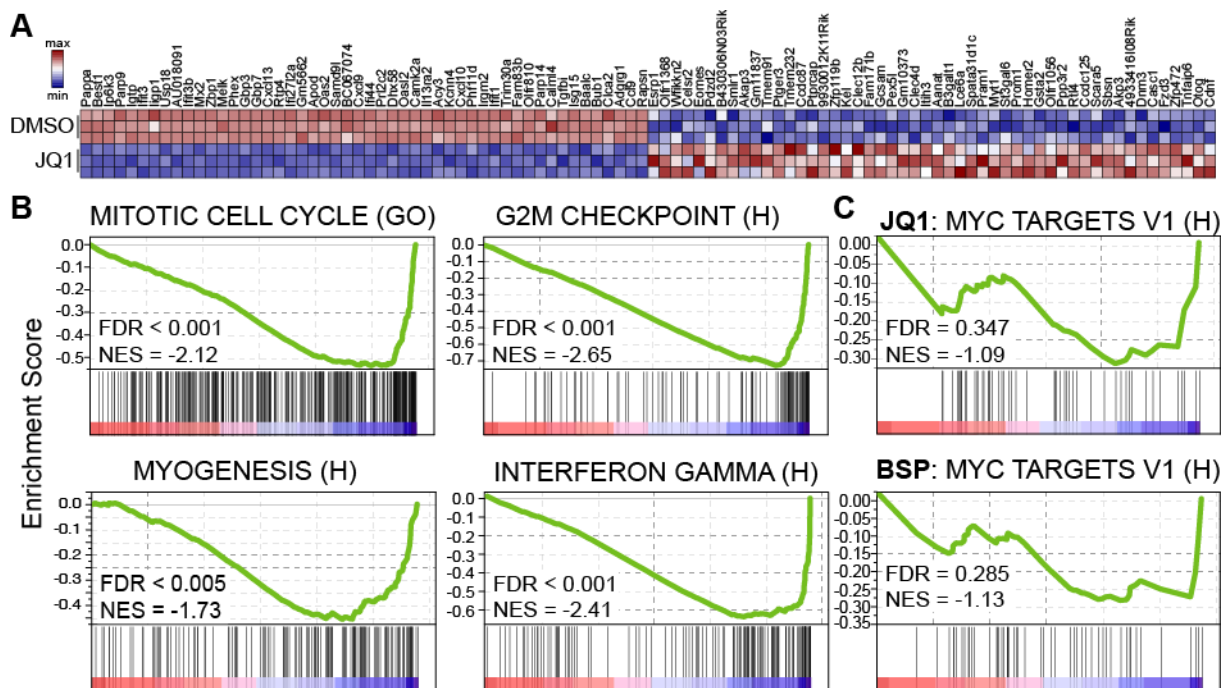


BAY-876



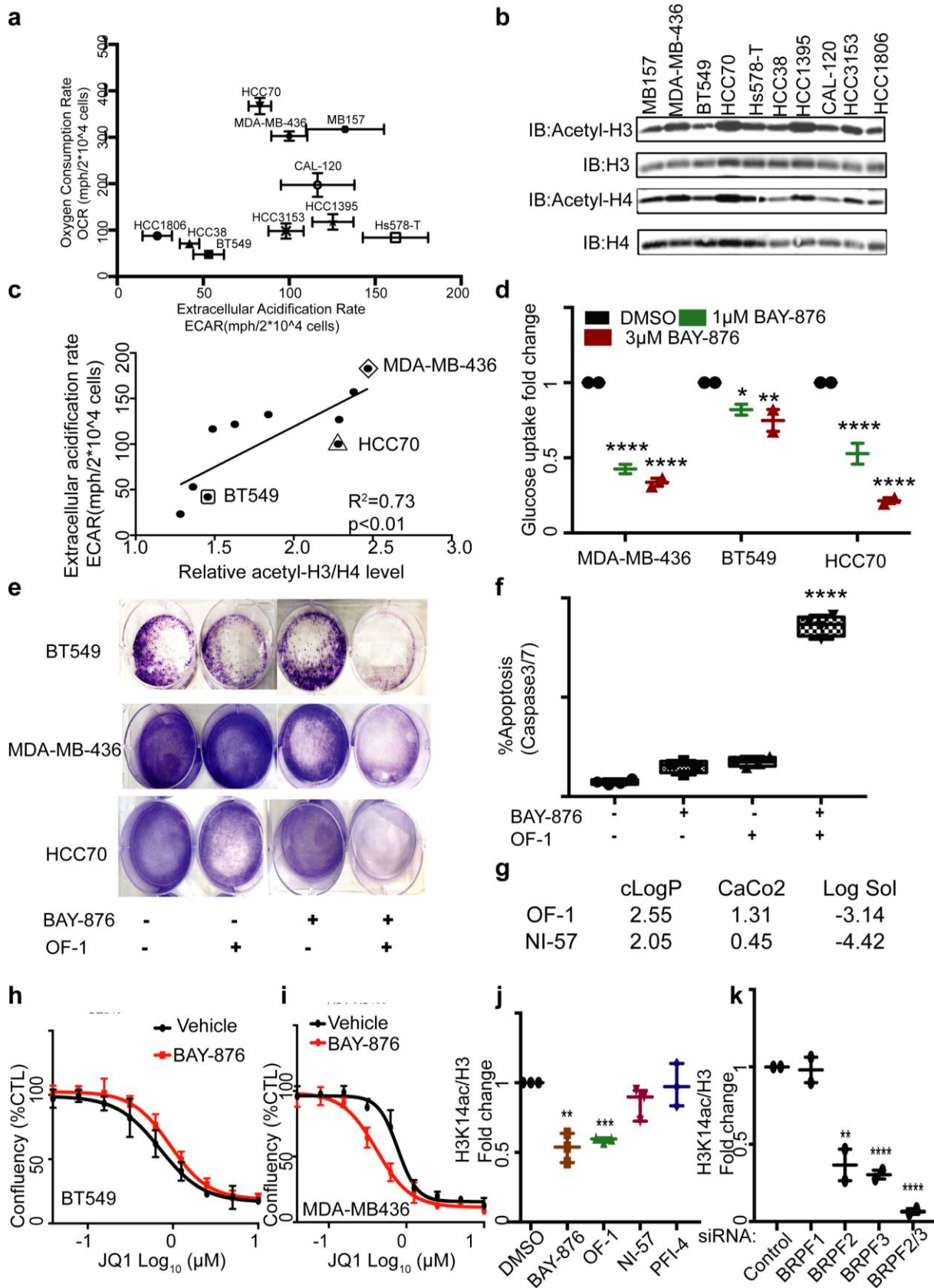
Supplementary figure 1: Chemical Structures of probe related compounds and BAY-876.

Supplementary Figure 2



Supplementary Figure 2: Transcriptional response to BSP and JQ1 in C2C12 myoblasts. **A)** Heatmap of the top 50 up/down regulated genes in C2C12 myoblasts following 12 h JQ1 treatment based on 2-sided signal to noise ratio (SNR) score and $P < 0.05$. Dark blue indicates lowest expression; dark red indicates highest expression, with intermediate values represented by lighter shades, as indicated in the inset. Data are column-normalized. **B)** GSEA demonstrating strong association with mitotic cell cycle (from the Gene Ontology MSigDB set, top left), G2M checkpoint (from hallmark MSigDB signatures, top-right), myogenesis (from hallmark MSigDB signatures, bottom-left) and interferon gamma signalling, following 12 h treatment of C2C12 cells with JQ1. **C)** GSEA demonstrating lack of association with MYC response following 12 h treatment of C2C12 cells with JQ1 (top panel) and BSP (lower panel).

Supplementary Figure 3

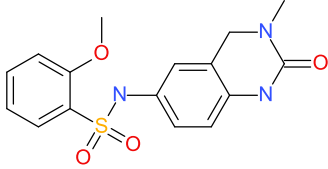
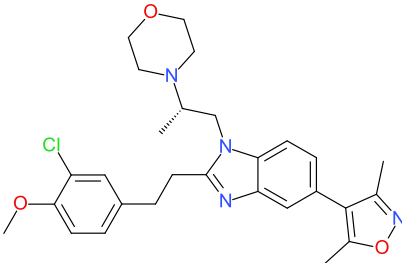
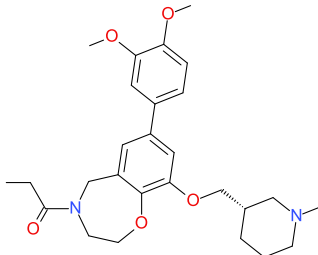
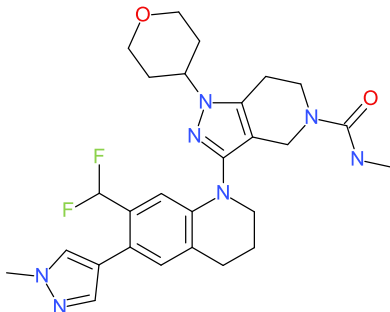
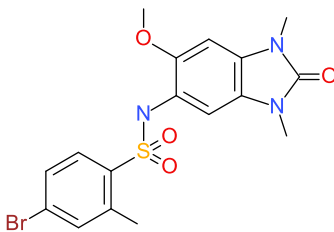


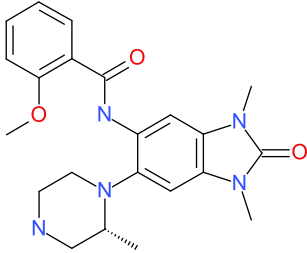
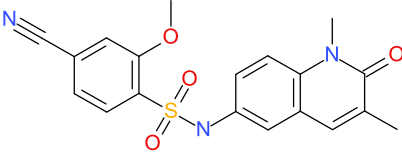
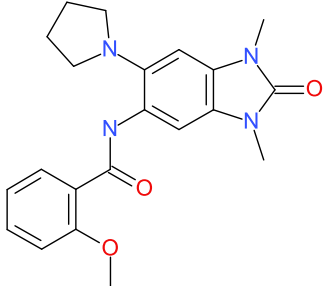
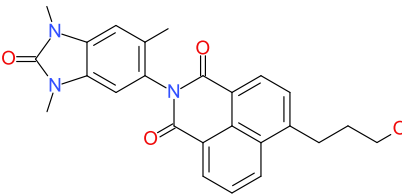
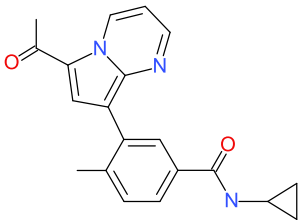
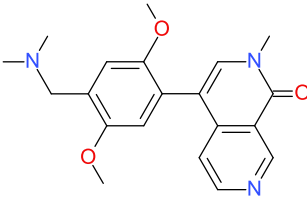
Supplementary Figure 3: Metabolic profiles and synergy with of BRPF bromodomain inhibition with BAY-876 a) Metabolic profiles of ten TNBC lines based on Seahorse XF96 measurements. Depicted are the ECAR (X axis) and the OCR (Y axis) measurements. Graph

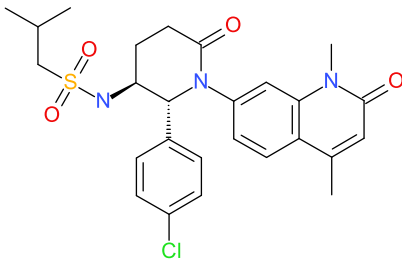
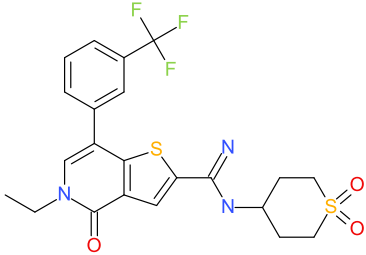
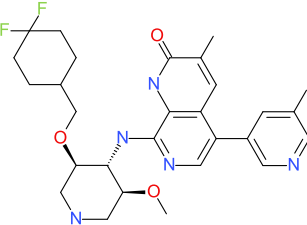
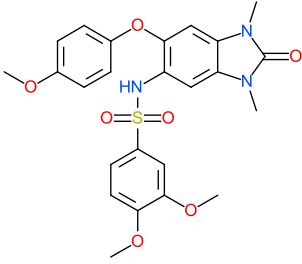
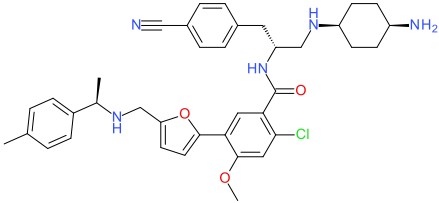
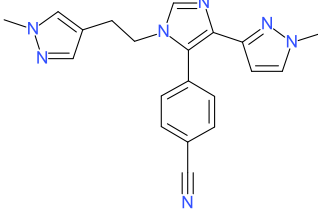
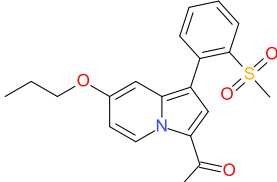
indicate mean and error bars denote standard deviation from eight wells (from two independent assays). **b)** A representative immunoblot of histone acetylation level across ten TNBC lines. **c)** Plot of ECAR and histone acetylation across ten TNBC lines. Labeled dots denote the representative TNBC lines, having extreme or medial glycolytic rates and histone acetylation levels, used for the following assays. **d)** Glucose uptake in three representative cell lines in respond to BAY-876 treatment relative to vehicle. Experiment repeated two times. Cells were treated with DMSO or indicated concentration of BAY-876 for 5 days. Graph indicate mean and error bars denote standard deviation from three independent assays. p value computed using a one-way ANOVA test; **** p<0.001. **e)** The colony formation of indicated cells with 3µM indicated chemical probes treatment for 14 days. **f)** MDA-MB436 cells were treated with 3µM indicated chemical probes for 5 days. Cell apoptotic activity was quantified by the fluorescent caspase 3/7 active objects. Graph indicate mean and error bars denote standard deviation from four independent assays. p value computed using one-way ANOVA t test; **** p<0.001. **g)** Pharmacokinetic properties of NI-57 and OF-1 predicted by pkCSM (<http://biosig.unimelb.edu.au/pkcsm/prediction>). **h)** Dose dependent curves for cell lines treated with indicated concentrations of JQ-1 with or without 3µM BAY876 for 7 days in BT549 cell line. Graph indicate mean and error bars denote standard deviation from four independent assays. p value computed using a two- stage step-up Benjamini t test **i)** Dose dependent curves for cell lines treated with indicated concentrations of JQ-1 with or without 3µM BAY876 for 7 days in MDA-MB-436 cell line. Graph indicate mean and error bars denote standard deviation from four independent assays. p value computed using a two- stage step-up Benjamini t test **j)** Immunoblot based quantification of H3K14ac abundance using total H3 as a control in MDA-MB-436 cells following control or 3 µM of the indicated chemical probes during a 5-day treatment. Graph indicate mean and error bars denote standard deviation from three independent assays. p value computed using a two-stage step-up Benjamini t test; **p<0.01; *** p<0.001. **k)** Immunoblot based quantification of H3K14ac abundance using total H3 as a control in MDA-MB-436 BRPF knockdown cells. Graph indicate mean and error bars denote standard deviation from two independent assays. p value computed using a two-stage step-up Benjamini t test; **p<0.01; *** p<0.001.

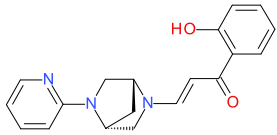
Supplementary Table 1: Probe set for the human bromodomain family

Target	Inhibitor Negative control Supplier [†]	SMILE	K _D (nM)	Ref.
Family I: PCAF/GCN5	L-Moses Negative control: D-Moses Probe: T Control: SGC		ITC data: 126 nM PCAF 600 nM GCN5	1
Family I: PCAF/GCN5	GSK4027 Negative control: GSK4028 SGC		BROMOscan 1.4nM PCAF 1.4nM GCN5	2
Family I: CECR2	NVS-CECR2-1 C, M, T		ITC data: 80nM CECR2	3
Family I: CECR2	GNE-886 MedKoo		EC ₅₀ 370nM CECR2	4
Family 2 BET (BRD4, 3, 2,T)	(+)-JQ1 Negative control: (-)-JQ1 C, M, T		ITC data: 49nM BRD4(1) 90nM BRD4(2) 59nM BRD3(1) 82nM BRD3(2) 128nM BRD2(1) 190nM BRDT(1)	5
Family 2 BET (BRD4, 3, 2,T)	I-BET151 C, T		Cell free assay IC ₅₀ of 500 nM, 25 nM, and 790nM	6

<p>Family 2 BET (BRD4, 3, 2,T)</p>	<p>PFI-1 C, M, T</p>		<p>ITC data: 47nM BRD4(1) 195nM BRD4(2) 80nM BRD3(1) 76nM BRD3(2) 108nM BRD2(1) 144nM BRD2(2)</p> <p>85nM BRDT(1) 221nM BRDT(2)</p>	<p>7</p>
<p>Family III CBP/p300</p>	<p>SGC-CBP30 Negative control: BDOIA513 Probe: C, M, T Control: SGC</p>		<p>ITC data: 21nM CBP 38nM p300</p>	<p>8</p>
<p>Family III CBP/p300</p>	<p>I-CBP112 C, M, T</p>		<p>ITC data: 151nM CBP 625nM p300</p>	<p>9</p>
<p>Family III CBP/p300</p>	<p>GNE-781 MedKoo</p>		<p>TR-FRET: 0.94 nM BRET: 6.2 nM</p>	<p>10</p>
<p>Family IV pan-BRPF</p>	<p>OF-1 C, M, T</p>		<p>ITC data: 101nM BRPF1B 505nM BRPF2 240nM BRPF3</p> <p>Off target BRD4(1) 3900 nM</p>	<p>11</p>

<p>Family IV pan-BRPF</p>	<p>GSK6853</p> <p>Negative control: GSK9311</p> <p>Probe: C, M Control: M</p>		<p>TR-FRET 81nM BRPF1B</p>	<p>12</p>
<p>Family IV pan-BRPF</p>	<p>NI-57</p> <p>C, M, T</p>		<p>ITC data: 31nM BRPF1B 108nM BRPF2 409nM BRPF3 Off target BRD9 998nM</p>	<p>11</p>
<p>Family IV BRPF1B</p>	<p>PFI-4</p> <p>C, M, T</p>		<p>ITC data: 13nM BRPF1B 775nM BRPF2 Off target CECR2 2350nM</p>	<p>11</p>
<p>Family IV BRPF2</p> <p>Family VII TAF1(2) Taf1L(2)</p>	<p>BAY-299</p> <p>Negative control: BAY-364</p> <p>C, M, T</p>		<p>ITC data: 44nM BRPF2 16nM TAF1(2) 234nM TAF1L(2)</p> <p>Off target 1390nM CBP</p>	<p>13</p>
<p>Family IV BRD7/BRD9</p>	<p>TP-472</p> <p>Negative control: TP-472N</p> <p>C, M, T</p>		<p>ITC data: 33nM BRD9 340nM BRD7</p>	<p>14</p>
<p>Family IV BRD7/BRD9</p>	<p>BI-9564</p> <p>C, M, T</p>		<p>ITC data: 15nM BRD9 BROMOscan 0.75nM BRD9 0.3 nM BRD7 Off target ITC CECR2: 187nM</p>	<p>15</p>

<p>Family IV BRD7/BRD9</p>	<p>LP-99 Negative control: (2S,3R)-LP99 Probe: C, M, T Control: SGC</p>		<p>ITC data: 99nM BRD9 909nM BRD7</p>	<p>16</p>
<p>Family IV BRD7/BRD9</p>	<p>I-BRD9 C, M, T</p>		<p>BROMOscan: 1.9 nM BRD9 380nM BRD7</p> <p>Off target CECR2 140nM CBP 740nM p300 770nM BRPF1 2100nM</p>	<p>17</p>
<p>(BRPF1B) (ATAD2 8Family IV ATAD2B</p>	<p>GSK8814 Negative control: GSK8815 SGC</p>		<p>ITC 10 nM ATAD2B</p>	<p>18</p>
<p>Family IV and VI TRIM24 BRPF1</p>	<p>Benzimidazolone 8 SGC</p>		<p>ITC 220nM TRIM24 140nM BRPF1B 1240nM BRPF2</p>	<p>19</p>
<p>Family IV ATAD2A</p>	<p>BAY-850 Negative control: BAY-460 SGC</p>		<p>BROMOscan 120nM ATAD2A</p>	<p>20</p>
<p>Family VI BAZ2A/B</p>	<p>BAZ2-ICR C, M, T</p>		<p>ITC: 109nM BAZ2A 170nM BAZ2B</p> <p>Off-target 1550nM CECR2</p>	<p>21</p>
<p>Family VI BAZ2A/B</p>	<p>GSK2801 Negative control: GSK8573 Probe: C, M, T</p>		<p>ITC: 257nM BAZ2A 136nM BAZ2B</p>	<p>22</p>

Family VIII SMARCA2/4 PB1(5)	PFI-3 Negative control: BDF25488524 Probe: C, M, T Control: SGC		ITC: 89nM SMARCA4 48nM PB1 (5)	23
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‡Cayman Chemical (C), Millipore-Sigma (M), Tocris (T)

References:

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Supplementary Table 2

Compound	TTK	CK2A1	CAMK1G	AAK1	ABL1	AURKB	BRAF	CAMKK2	STK10
L-Moses	-0.5	-0.1	-0.3	-1.6	0.0	0.4	-0.1	-0.1	-0.4
I-CBP112	0.0	-1.9	-0.2	-0.7	0.0	0.3	-0.1	-0.4	-0.4
PFI-4	-0.6	-0.5	-0.6	-0.9	0.0	0.4	0.6	0.4	-0.4
LP-99	-0.1	-1.4	-0.2	-0.8	-0.1	0.1	-0.2	0.0	-0.3
PFI-3	-0.7	0.3	0.0	-0.3	-0.1	0.2	-0.1	-0.1	-0.2
NVS-CECR2-1	0.4	-1.0	-0.4	-0.7	0.0	0.3	-0.1	-0.3	-0.1
OF-1	0.1	-2.4	-0.7	-0.9	-0.1	0.3	0.0	-0.3	-0.2
BAY-299	-0.1	-1.4	-0.6	-0.4	0.0	0.2	0.1	-0.4	-0.2
I-BRD9	0.2	-1.1	-0.3	-0.4	0.0	0.2	0.0	-0.2	-0.2
JQ1	-0.5	-1.4	-0.9	-2.8	-0.2	-0.1	-0.4	-0.6	-0.6
GSK6853	0.1	-0.7	-0.5	-0.8	0.1	1.0	0.1	0.5	0.1
TP-472	0.2	-1.1	-0.2	-0.3	0.1	0.2	0.8	0.0	-0.2
BAZ2-ICR	0.3	-1.3	-0.1	0.8	0.0	0.0	0.1	0.0	0.0
SGC-CBP30	-0.4	-1.8	-0.6	-0.6	-0.2	0.0	-0.5	-0.6	0.0
NI-57	0.0	-1.7	-0.6	0.2	-0.1	0.0	0.0	-0.4	0.2
BI-9564	0.1	-1.2	-0.6	0.1	0.0	0.1	-0.2	-0.2	-0.2
GSK2801	0.4	-1.0	-0.3	0.5	0.0	-0.1	-0.3	0.1	-0.5
	EPHA2	FES	GPRK5	GSG2	MERTK	PAK4	MAPK1	MAPK8	STK3
L-Moses	1.1	-0.2	-0.3	0.1	0.0	-0.3	0.1	0.6	0.0
I-CBP112	1.1	-0.1	-0.4	0.1	-0.2	-0.2	0.2	0.3	0.0
PFI-4	1.8	0.0	-0.3	0.3	-0.1	-0.2	0.2	-0.2	0.0
LP-99	0.9	0.0	-0.3	-0.2	-0.2	-0.4	0.2	-0.1	-0.2
PFI-3	-0.2	-0.2	-0.2	0.0	-0.1	-0.2	-0.1	0.1	-0.1
NVS-CECR2-1	1.5	-0.5	-0.1	0.2	0.0	-0.1	0.3	-0.3	-0.1
OF-1	1.2	-0.6	-0.4	0.2	-0.1	-0.2	0.3	-0.1	0.0
BAY-299	2.0	0.0	-0.3	0.1	-0.2	-0.1	0.0	-0.4	0.1
I-BRD9	1.2	0.1	-0.1	0.1	0.0	-0.1	0.1	-0.1	0.1
JQ1	0.9	-0.8	0.1	0.1	-0.3	-0.4	0.2	-0.8	0.1
GSK6853	1.5	-0.2	-0.3	0.6	0.3	0.1	0.2	0.1	0.3
TP-472	2.1	0.0	-0.1	0.1	-0.1	0.0	0.2	0.2	0.1
BAZ2-ICR	1.1	0.0	0.0	0.0	0.0	0.0	0.1	0.2	0.0
SGC-CBP30	1.2	-0.6	0.1	0.0	-0.1	0.0	0.2	0.8	0.0
NI-57	1.5	-0.4	-0.3	0.0	0.0	0.1	0.1	0.1	0.2
BI-9564	2.1	-0.1	-0.4	0.0	0.2	0.0	0.1	0.0	0.0
GSK2801	1.2	-0.2	-0.5	0.2	0.0	-0.2	0.2	0.1	0.1
	CASK	BMX	CSNK1D	DAPK3	DYRK1A	PHGK2	PIM1	RPS6KA1	
L-Moses	-0.1	-0.1	0.6	0.2	0.2	-0.1	2.3	0.2	
I-CBP112	0.0	-0.1	0.5	0.0	-0.4	0.1	-0.1	0.1	
PFI-4	-0.2	0.6	1.3	0.4	-0.5	0.1	0.5	0.1	
LP-99	-0.3	0.2	0.2	-0.1	-0.3	-0.1	-0.2	-0.1	
PFI-3	-0.2	0.3	0.3	0.0	-0.1	0.1	0.1	-0.2	
NVS-CECR2-1	0.1	0.0	0.4	0.7	-0.3	-0.2	0.4	0.2	
OF-1	0.0	0.5	0.6	-0.1	-0.8	0.1	-0.2	0.0	
BAY-299	-0.1	0.3	1.2	-0.2	-0.5	0.0	-0.2	0.1	
I-BRD9	-0.2	0.3	0.0	-0.2	-0.6	0.0	0.1	0.1	
JQ1	0.1	-0.3	0.4	-0.1	0.2	-0.3	-0.3	-0.1	

GSK6853	0.1	0.3	1.3	1.1	-0.2	0.2	3.7	0.2
TP-472	-0.1	0.5	1.1	0.0	-0.3	0.1	0.4	0.1
BAZ2-ICR	0.2	0.5	0.4	-0.1	-0.1	-0.1	0.2	0.1
SGC-CBP30	0.5	-0.2	0.3	0.2	-0.2	-0.3	-0.2	0.0
NI-57	0.3	0.0	0.6	0.0	0.1	0.1	0.1	0.2
BI-9564	0.0	0.1	1.1	0.1	0.1	-0.1	0.0	0.1
GSK2801	0.5	0.2	0.2	-0.2	0.4	-0.2	-0.3	-0.1

Supplementary table 2: Selectivity screening data (protein kinases): Temperature shift data measured on protein kinases. Shown are average data in Kelvin of three measurements. No significant T_m shifts have been observed.

Supplementary Table 3

Probes	ΔT_m ($^{\circ}\text{C}$) 10 μM									
	UHRF1	RBBP1	53BP1	BRPF1	BRPF3	WDR5	EED	TDRD3	FXR1	SND1
BAY-299	NA	-1.0	0.3	-2.5	1.5	0.2	NA	-1.4	0.8	NA
BAY-588	-0.4	-0.1	-2.3	-0.2	NA	-7.3	-0.5	-5.3	-1.2	-1.0
BAY-850	-0.3	-1.0	-0.7	-2.2	-0.4	0.2	NA	-1.8	-1.7	-1.6
BAY-876	-0.2	-0.3	0.0	-0.3	-0.3	0.4	-0.7	-1.0	0.8	-0.9
BI-9564	-0.2	0.0	-0.3	-0.3	-0.6	-0.2	-0.7	-0.6	0.3	-0.9
GSK4027	-0.2	-0.7	-0.1	-0.8	-0.3	0.2	-0.7	-0.6	0.3	-1.0
GSK6853	-0.2	-0.7	-0.1	-0.9	-0.4	0.0	-0.9	-0.7	-1.1	-0.6
GSK8814	-0.2	-0.1	-0.1	-0.2	-0.2	0.2	-0.4	-0.6	0.4	-0.5
I-BRD9	-0.1	0.0	0.1	0.2	0.1	0.1	-0.7	-0.4	0.1	-0.3
I-CBP112	0.0	-0.1	0.3	-0.2	0.0	-0.2	-0.5	-0.6	0.9	-0.7
JQ1	-0.2	-0.7	-0.4	-0.6	-0.7	-0.3	-0.9	-1.4	-0.5	-0.6
L-Moses	NA	-0.4	NA	-2.7	1.0	0.5	-0.3	-0.9	0.3	-1.0
LP99	NA	-0.7	0.3	-2.0	0.0	0.4	-0.2	-0.5	0.1	-0.4
NI57	NA	-0.1	-0.1	NA	-0.4	0.3	-0.4	-0.7	-0.5	-0.4
NVS-CECR2-1	-0.3	-1.0	-0.6	-0.2	-0.3	-0.2	-1.0	-3.3	-5.1	-1.7
OF-1	0.2	-0.7	-0.1	-0.6	0.0	-0.2	-0.7	-1.0	0.7	-0.7
PFI-1	0.2	-0.6	-0.2	-0.3	-0.2	0.0	-0.9	-1.7	-0.1	-0.7
PFI-3	0.5	-0.7	0.1	-0.5	0.4	0.2	-0.9	-0.6	0.4	-0.6
PFI-4	-0.5	-1.2	-0.7	-0.7	0.0	-0.5	-2.2	-1.5	-1.2	-1.1
SGC-CBP30	0.0	-1.2	-0.2	0.6	0.2	0.2	-0.4	-1.1	-1.2	-0.4
TP-238	0.0	-0.4	0.4	-0.3	0.2	0.2	-0.3	-0.7	0.4	-0.5
TP-472	-0.1	-0.5	0.1	0.2	0.3	0.0	-0.5	-0.1	0.8	-0.5

Supplementary Table 3: Selectivity screening data (non-BRD reader domains): Temperature shift assay data of BRD probes screened against a diverse panel of methyl-lysine reader domains. Compounds were screened a concentration of 10 μM . Shown are temperature shifts in Kelvin. No significant T_m shifts have been observed.

Supplementary Table 4

Probes	HATs Activity (%) at 1 and 10 μ M									
	EP300		HAT1		MYST1		MYST3		GCN5L2	
	1 μ M	10 μ M	1 μ M	10 μ M	1 μ M	10 μ M	1 μ M	10 μ M	1 μ M	10 μ M
BAY-299	95	93	100	93	99	99	98	85	99	103
BAY-588	103	99	99	97	100	93	99	96	94	95
BAY-850	98	87	101	101	93	94	96	97	99	102
BAY-876	99	95	97	89	96	91	99	101	100	97
BI-9564	99	94	99	96	106	101	99	100	102	100
GSK4027	93	90	102	89	102	98	101	101	99	102
GSK6853	98	104	100	94	99	101	107	104	107	107
GSK8814	96	102	100	95	99	103	96	102	102	100
I-BRD9	96	99	103	101	104	104	101	99	104	102
I-CBP112	97	99	100	94	98	103	98	99	105	102
JQ1	99	102	98	103	99	95	108	100	104	98
L-Moses	94	90	100	95	97	96	92	90	99	95
LP99	99	98	104	95	90	88	96	100	92	89
NI57	94	93	104	104	96	91	98	99	100	96
NVS-CECR2-1	90	84	99	88	96	94	98	103	100	100
OF-1	100	103	92	93	102	94	100	102	98	93
PFI-1	99	100	102	93	101	95	105	86	96	93
PFI-3	98	97	96	94	99	99	95	96	99	101
PFI-4	99	101	98	96	98	101	97	102	98	96
SGC-CBP30	98	92	96	97	103	97	103	103	101	101
TP-238	99	95	93	100	99	97	97	99	96	92
TP-472	95	96	98	97	100	103	99	98	94	92

Supplementary Table 4: Selectivity screening data (HATs): Effects of BRD probe compounds on the activity of histone acetyltransferases (HATs). Shown are inhibition data (% inhibition) at a compound concentration of 1 μ M and 10 μ M.

Supplementary Table 5

MTases	MTases Inhibition (%) at 10 μ M							
	BAY-588	BAY-850	BAY-876	GSK4027	GSK6853	GSK8814	L-Moses	TP-238
ASH1L	-6	3	4	13	4	2	-3	-4
G9A	3	7	-3	1	-2	-2	-3	7
GLP	2	2	0	2	3	2	2	1
MLL1 (Trimeric)	7	-1	11	8	7	6	20	17
MLL3 (pentameric)	7	0	0	2	2	0	1	-3
NSD2	2	-1	0	15	13	24	2	3
NSD1	11	2	2	4	1	19	-6	1
NSD3	-2	14	3	10	7	13	5	15
PRC2-EZH1	3	19	-4	-1	1	1	5	6
PRC2-EZH2	3	3	5	-5	0	2	-4	3
PRDM9	3	10	0	3	-2	0	4	-1
SETD2	0	7	-4	-1	1	7	11	19
SETD7	10	11	10	12	7	7	7	-1
SETD8	-3	2	0	10	-6	-7	-8	-6
SETDB1	5	8	-4	-1	1	1	1	-1
SMYD2	8	19	-10	2	-3	-2	-10	-2
SMYD3	7	6	3	1	9	8	10	7
SUV39H1	12	5	16	-3	-3	-2	-2	-1
SUV39H2	12	-2	-2	5	1	3	5	7
SUV420H1	-5	5	-1	8	5	-3	4	3
SUV420H2	-3	0	-1	-1	-2	0	6	0
DOT1L	3	3	-6	-2	10	5	1	6
PRMT1	0	15	2	4	-7	3	-2	0
PRMT3	5	3	1	2	1	3	0	5
PRMT4	9	25	1	-2	2	1	6	3
PRMT5	4	4	-1	8	3	-2	-3	-12
PRMT6	4	13	-3	6	5	0	5	6
PRMT7	13	3	12	15	11	11	8	9
PRMT8	3	4	9	15	3	5	5	3
PRMT9	3	15	12	8	12	3	13	7
BCDIN3D	16	28	17	14	5	7	9	4
DNMT1	9	4	4	3	-8	-3	3	3
DNMT3A/3L	-1	21	6	3	13	8	2	8
DNMT3B/3L	-6	28	1	1	8	18	-1	0

Supplementary table 5: Selectivity screening data measured against a panel of lysine methyl transferases (MTases). Inhibition (%) was assessed at inhibitor concentration of 10 μ M. Shown are average values of two measurements.

Supplementary Figure 4

Figure 6F-full gel

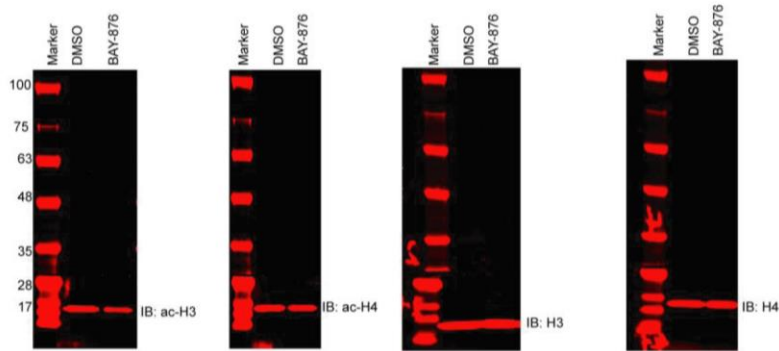


Figure 7D-full gel

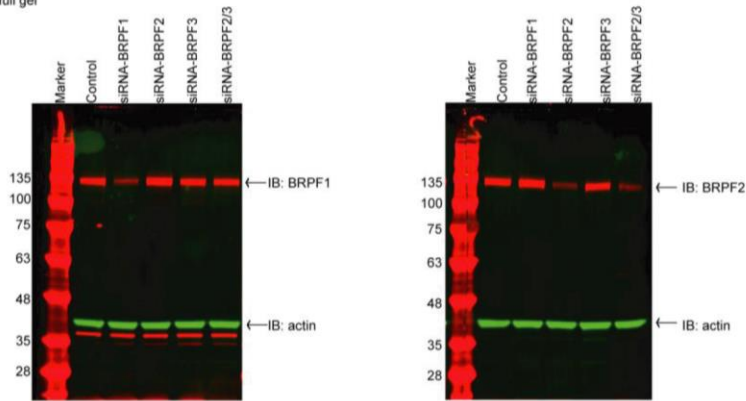


Figure 7G-full gel

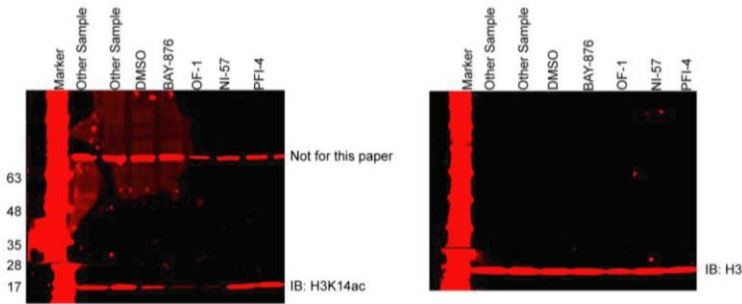
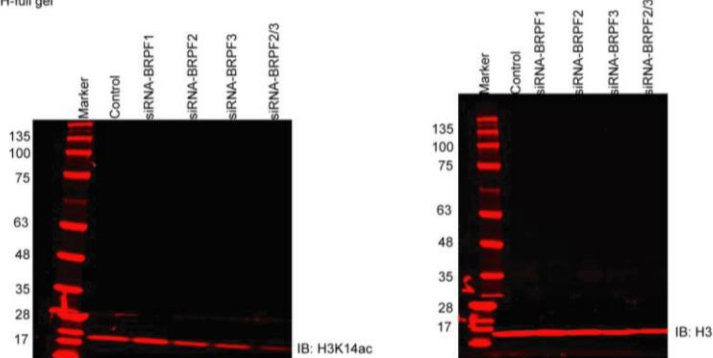


Figure 7H-full gel



Supplementary Figure 4: Raw data blots used in figure 6 and 7.