

Expanded View Figures

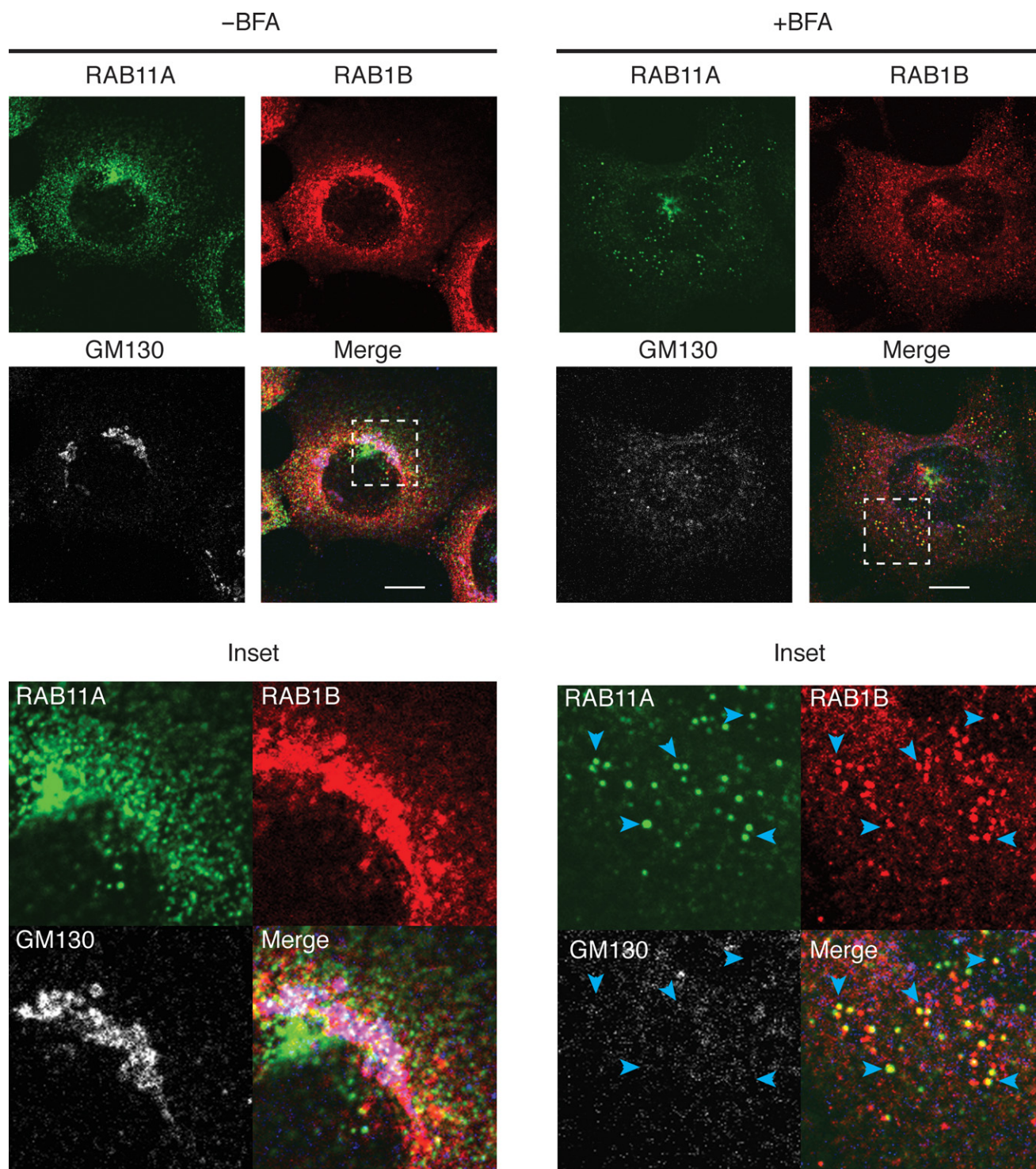


Figure EV1. Co-localisation of endogenous RAB1B and RAB11A.

HEK293A cells were treated with 10 $\mu\text{g/ml}$ BFA for 30 min (+ BFA) or not (-BFA), fixed and stained for RAB11A (green), RAB1B (red) and GM130 (white, blue in merge). Scale bars, 10 μm . Blue arrowheads indicate RAB1B- and RAB11A-positive structures—note the absence of the early Golgi marker GM130 on these puncta.

Figure EV2. Identification of an endogenous TRAPPIII-like complex in mammalian cells.

- A Scatterplot of \log_{10} -transformed iBAQ values for myc-BioID versus myc-BioID-TBC1D14 non-denaturing pulldowns analysed by mass spectrometry.
- B Scatterplot of \log_{10} -transformed iBAQ values for myc-BioID-TBC1D14 denaturing versus myc-BioID-TBC1D14 non-denaturing pulldowns analysed by mass spectrometry.
- C HEK293A cells (1×10^6 cells per condition) were lysed in 400 μ l TNTE and subjected to immunoprecipitation with mouse anti-TRAPPC12/TTC15 or mouse anti-GFP bound to protein G-Sepharose. The bound proteins were immunoblotted for TRAPPC8 and TRAPPC12.
- D TRAPPC8, TRAPPC12 and the core subunit TRAPPC4 co-elute in a complex of approximately 600 kDa. Blots are representative of three independent experiments. Approximate elution profiles of molecular weight standards are indicated above the fractions.
- E TBC1D14 overexpression does not promote GTP binding to RAB1B. Bar chart is quantification of the percentage of RAB1B precipitated (normalised to GFP-Golgin-84) upon myc-TBC1D14 overexpression compared to empty vector expression from three independent experiments. n.s. = not significant, unpaired *t*-test.
- F Lysates of HEK293A cells expressing GFP-Golgin-84 and transfected with RISC-free control siRNA or siRNA targeting TBC1D14, TRAPPC8 or RAB1B were subjected to GFP-Trap IP as in (E) and immunoblotted for RAB1B, TBC1D14, TRAPPC8, TRAPPC4 and GFP. Bar chart is quantification of the percentage of RAB1B precipitated (normalised to GFP-Golgin-84) upon myc-TBC1D14 overexpression compared to RISC-free siRNA from three independent experiments. ***P* < 0.01, ****P* < 0.001, one-way ANOVA followed by Sidak's multiple comparison test.

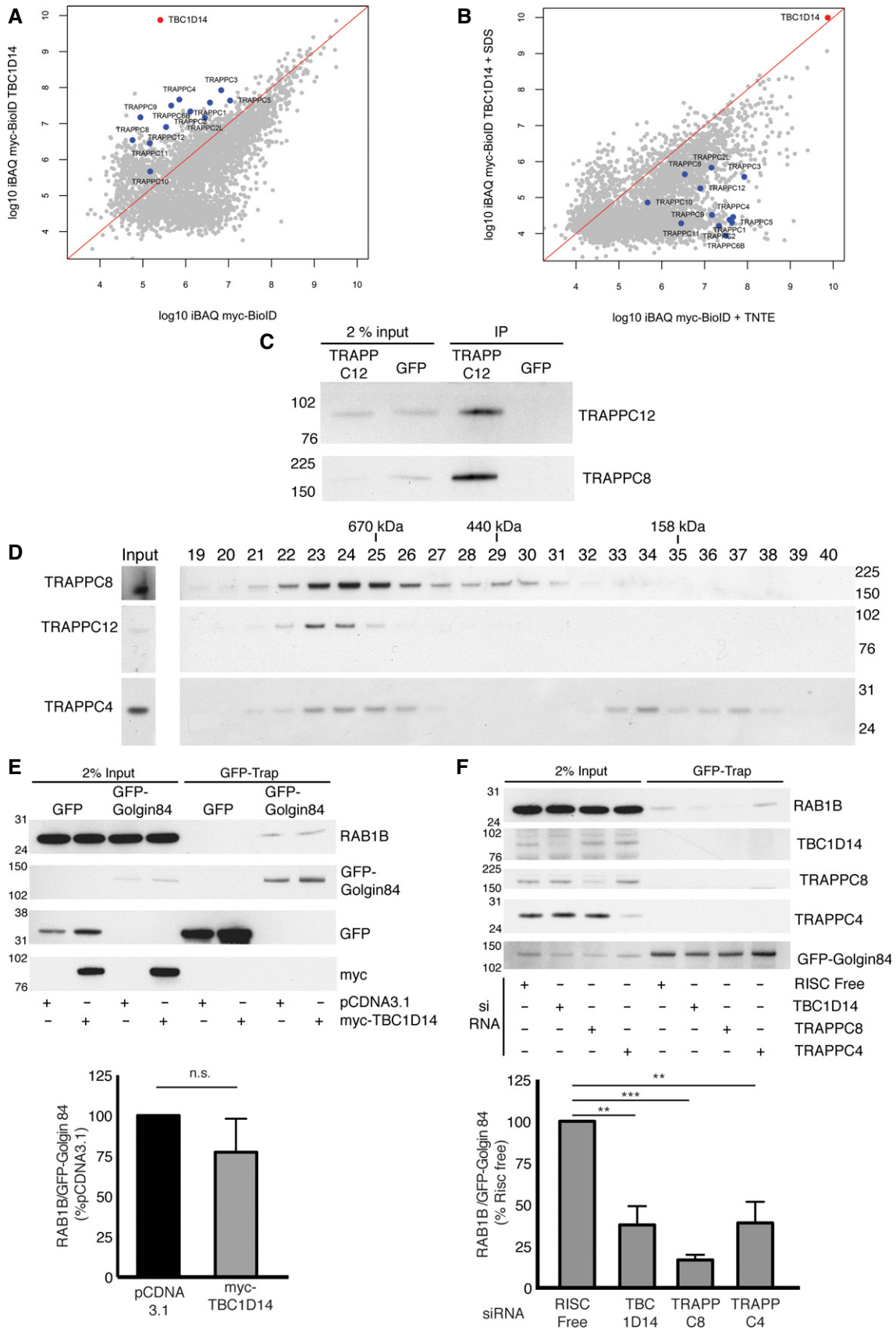
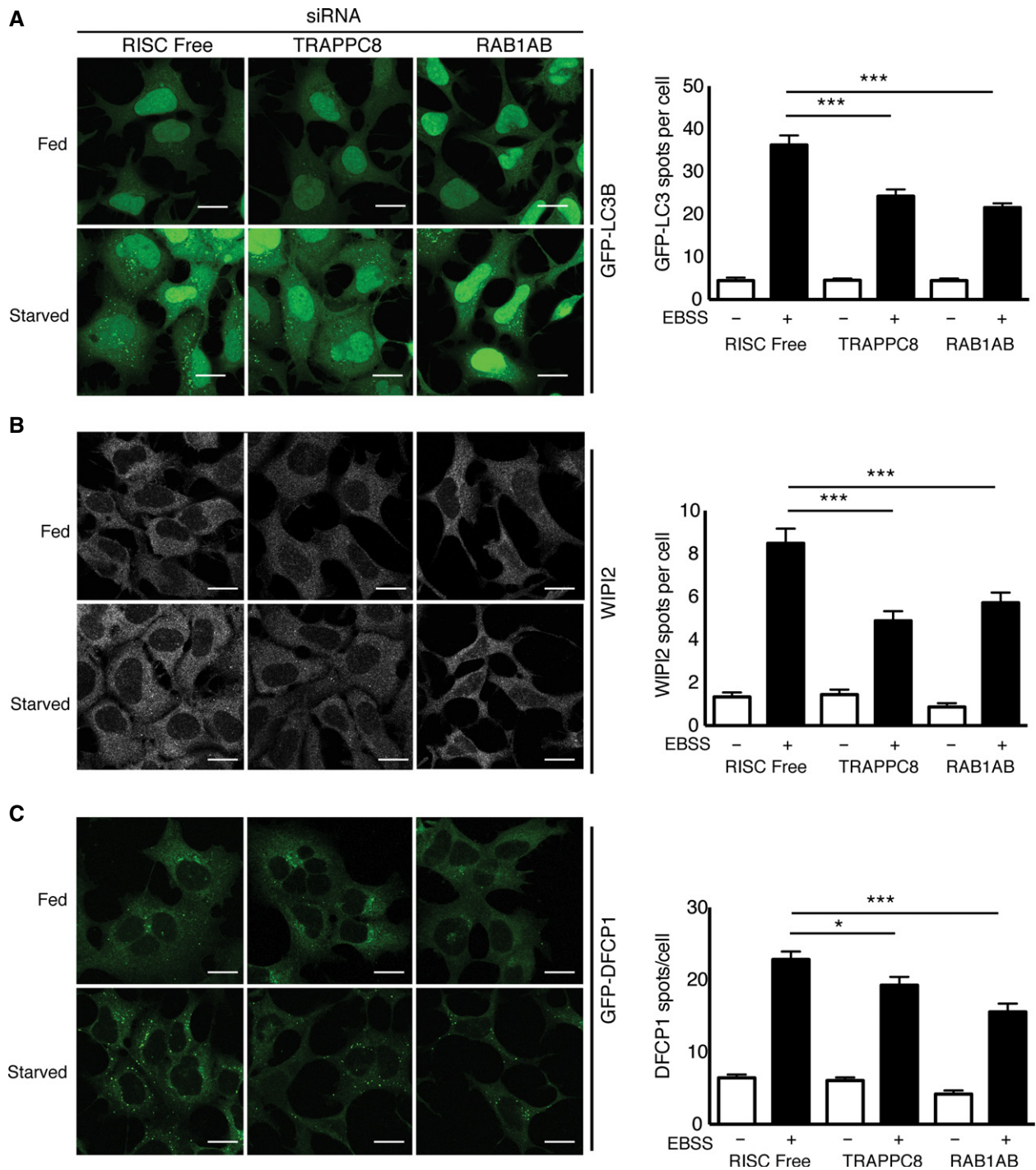


Figure EV2.



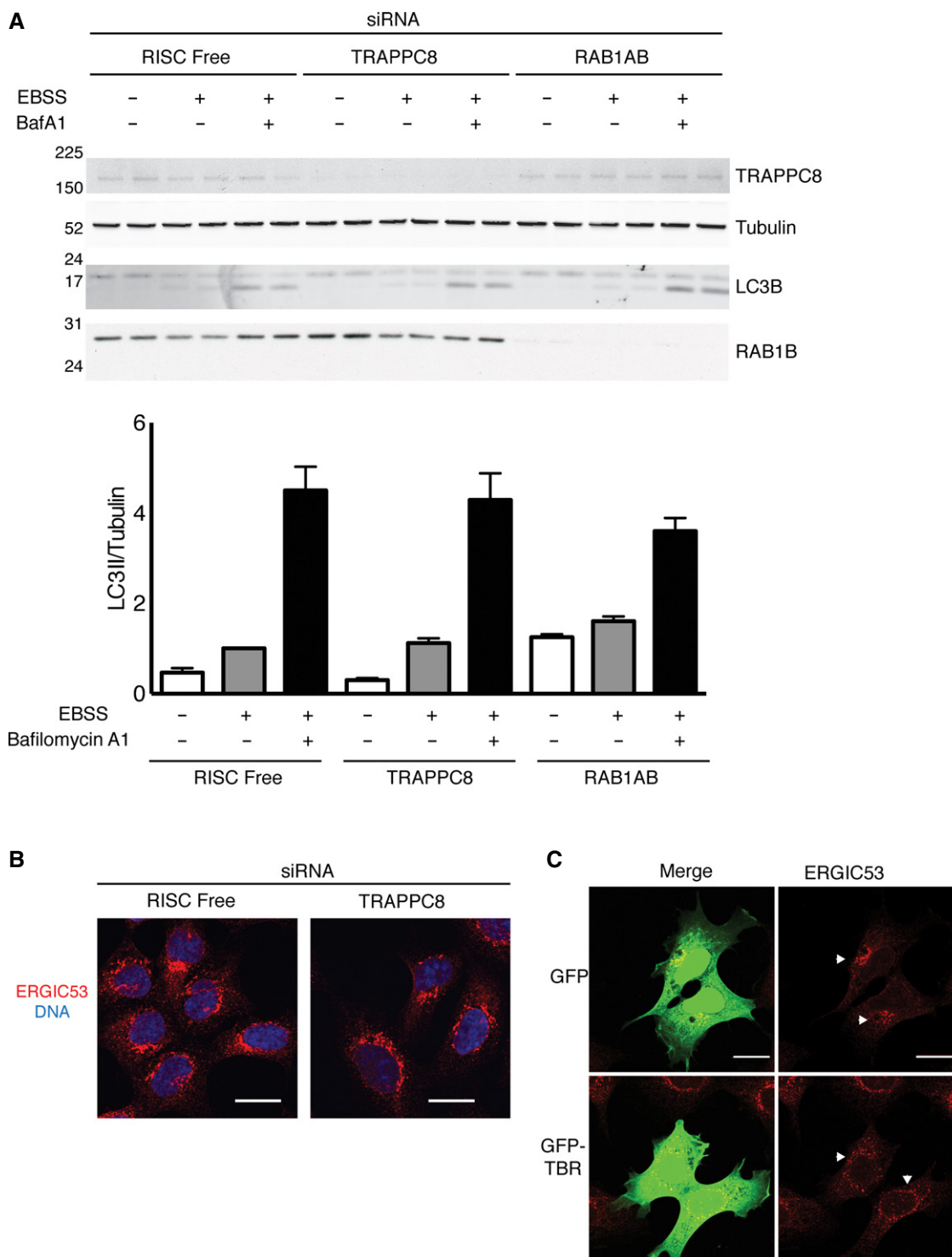


Figure EV4. Depletion of TRAPPC8 does not affect LC3 lipidation, and RAB1 depletion has a mild effect.

A HEK293A cells transiently transfected with control RISC-free control siRNA or siRNA duplexes directed against TRAPPC8 or RAB1AB were treated with EBSS, EBSS + 100 nM bafilomycin A1 or not for 2 h, lysed and subjected to immunoblotting for LC3B, tubulin, TRAPPC8 and RAB1B. The amount of LC3B/tubulin for each condition from three independent experiments is shown in the bar graph, \pm s.e.m.

B Cells depleted for TRAPPC8 or not (RISC free) were immunostained for ERGIC53 (red) and Hoechst (DNA; blue). Scale bars, 20 μ m.

C Cells expressing GFP or GFP-TBR (green) were immunostained for ERGIC53 (red; arrowheads indicate transfected cells) and analysed by confocal microscopy. Scale bars, 20 μ m.

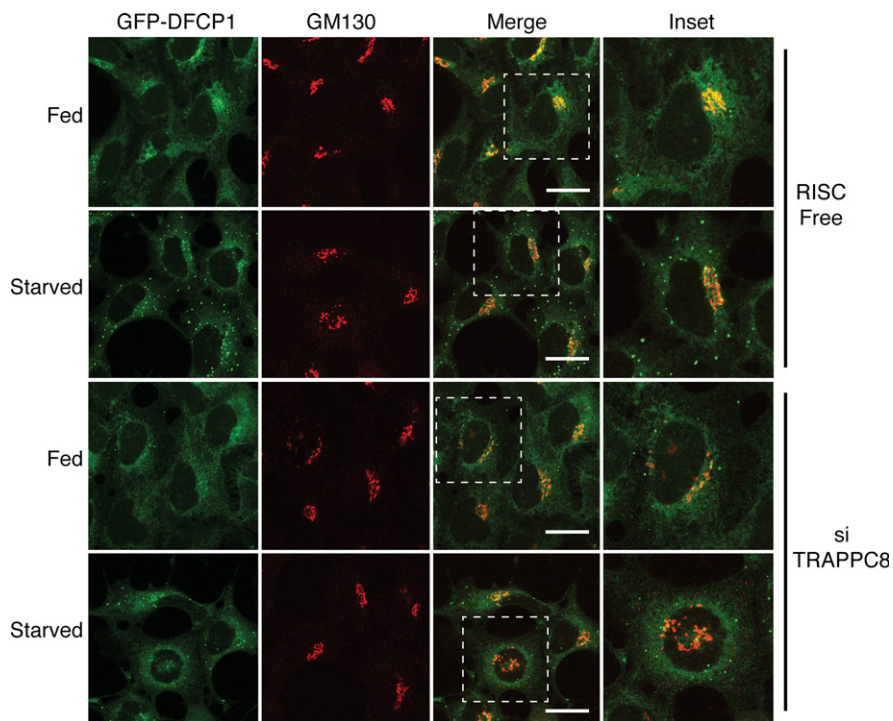


Figure EV5. Depletion of TRAPPC8 results in mis-localisation of GFP-DFCP1.

HEK293A cells stably expressing GFP-DFCP1 (green) were transiently transfected with control siRNA or siRNA targeting TRAPPC8, starved in EBSS for 2 h (starved) or not (fed), fixed, stained for GM130 (red) and analysed by confocal microscopy. Scale bars, 20 μ m.