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**Abstract.** The South African endemic bee genus *Redivivoides* Michener, 1981 is revised and redefined. The genus comprises seven species, six of which are described here as new: *Redivivoides capensis* sp. nov. ♀♂, *R. eardleyi* sp. nov. ♀, *R. kamieskroonensis* sp. nov. ♀, *R. karooensis* sp. nov. ♀♂, *R. namaquaensis* sp. nov. ♀♂ and *R. variabilis* sp. nov. ♀♂. A key to species is provided.

**Key words.** *Redivivoides*, bees, South Africa, new species, taxonomy.


**Introduction**

The bee genus *Redivivoides* was described by Michener (1981) based on a single species, *R. simulans* Michener, 1981, collected in the winter rainfall region of western South Africa. Two other species were only known from a few female specimens and remained undescribed. Since then much more material of this genus has become available representing now a total of seven species, six of which are described here as new. The morphological diversity of the newly described species required a re-evaluation of the characters defining *Redivivoides* as a genus.

*Redivivoides* belongs to the subfamily Melittinae Schenck, 1860 and the tribe Melittini Schenck, 1860 that also includes the genera *Melitta* Kirby, 1802 and *Rediviva* Friese, 1911 (Michez *et al*. 2009). Michener (1981) suggested that *Redivivoides* is the sister-group of *Rediviva* and the close relationship of both genera was later confirmed by phylogenetic studies (Danforth *et al*. 2006; Michez *et al*. 2009). However, unlike *Redivivoides*, the females of *Rediviva* collect floral oil that is mixed with pollen for nest provisioning from a range of oil-producing flowers, with *Diascia* (Scrophulariaceae) as their principal floral host (Whitehead & Steiner 2001; Pauw 2006; Whitehead *et al*. 2008). In several *Rediviva* species the forelegs are elongate, sometimes longer than the entire body, and the lengths of floral spurs and bee legs in some cases show co-variation at the population level, suggesting co-evolution (Steiner & Whitehead 1990, 1991). Because Melittidae are the most basal bees (Danforth *et al*. 2006), *Redivivoides* and *Rediviva* are a key group for understanding the evolution of oil-collecting in bees (Michez *et al*. 2009) and they might also help to understand the origin of the unusual bee diversity in the Greater Cape Floristic Region (Kuhlmann 2009).
The goal of this publication is to describe the six new species and to provide a key for species identification to facilitate further research on this fascinating group of bees. Based on the descriptions and the analysis of the morphology of the new species, *Redivivoides* is redefined as a genus and its relationship to its putative sister group, the oil-collecting *Rediviva* bees, is briefly discussed.

**Material and methods**

Terminology for the description of species is based on Michener (2007) for general morphology. Puncture density is expressed as the relationship between puncture diameter (d) and the space between them (i), such as $i = 1.5 \ d$ or $i < d$. The following abbreviations were used for morphological structures:

$T =$ metasomal tergum

$S =$ metasomal sternum

Body length was measured from the vertex to the apex of the body.

Acronyms for collections (after Arnett *et al*. 1993) from which specimens were borrowed or deposited, are as follows:

- **AMGS** = Albany Museum, Grahamstown, South Africa
- **CUIC** = Cornell University Collection, Ithaca, USA
- **EMUS** = Entomological Museum, Utah State University, USA
- **LPCT** = Laurence Packer collection, Toronto, Canada
- **SAMC** = South African Museum, Cape Town, South Africa
- **SANC** = South African National Collection of Insects, Pretoria, South Africa
- **RCMK** = research collection of Michael Kuhlmann, London, UK

Nomenclature of plant names follows Germishuizen & Meyer (2003). If not given on the labels, coordinates of collecting sites are given in square brackets and places were identified using Microsoft Encarta® World Atlas (version 10, 2001). Geographical coordinates are given in the following format: degree.minutes.seconds. Distribution maps were generated using DMAP V7.2 ([www.dmap.co.uk](http://www.dmap.co.uk)).

Scanning electron microscopy (SEM) was carried out using a Leo 1455VP. The backscattered electron images were taken under low vacuum (variable pressure) of uncoated specimens. In this mode electron-poor organic material appears darker than electron-rich material (e.g. soil dust particles) consisting of heavier chemical elements.

**Results**

Classis *Hexapoda* Blainville, 1816
Ordo *Hymenoptera* Linnaeus, 1758
Superfamilia *Apoidea* Latreille, 1802
Epifamilia *Anthophila* Latreille, 1804
Familia *Melittidae* Schenck, 1860
Subfamilia *Melittinae* Schenck, 1860
Tribus *Melittini* Schenck, 1860
Genus *Redivivoides* Michener, 1981
Redivivoidea Michener, 1981

Redivivoidea Michener, 1981: 42-44, figs 80, 82-84 (p. 43), 97 (p. 48).

Type species: Redivivoidea simulans Michener, 1981, by original designation.

The diagnosis of Redivivoidea given by Michener (1981) was solely based on R. simulans, which was the only known species of the genus at that time. However, in some respects R. simulans is an exception within the genus so a redefinition of Redivivoidea is required in the light of the six new species described in this paper (see discussion).

A comprehensive phylogenetic analysis of Redivivoidea and related genera is in preparation to investigate the intra-generic relationships, so for convenience the species are here listed in alphabetical order.

Redivivoidea capensis sp. nov.
Figs 1E–F, 2, 3, 4

Diagnosis

Males and females of R. capensis sp. nov. can be separated from other Redivivoidea species by their extensively and brightly red metasomal terga (Figs 2D, 3C) in combination with an impunctate and densely shagreened scutum (Figs 2C, 3B).

Etymology
Named after the Cape of Good Hope ("the Cape").

Type material (18 specimens)

Holotype

Paratypes
SOUTH AFRICA: 4 ♀♀ same date as Holotype (RCMK); 1 ♀, N. Cape, Fynbos, 2 km W Nieuwoudtvile, near Grasberg [31°21’05” S, 19°05’49” E], 742 m, 23 Aug. 2003, K. Timmermann (RCMK); 1 ♀, N. Cape, Nieuwoudtville, Flower Reserve East [31°21’56” S, 19°08’52” E], 735 m, 11 Sep. 2003, M. Kuhlmann (RCMK); 1 ♀, 2 ♂♂, N. Cape, Nieuwoudtville, Glen Lyon, Renosterveld [31°24’03” S, 19°08’34” E], 700 m, 7 Aug. 2004, M. Kuhlmann (RCMK); 2 ♂♂, N. Cape, Nieuwoudtville, Glen Lyon, Renosterveld [31°24’03” S, 19°08’34” E], 700 m, 11 Aug. 2004, M. Kuhlmann (RCMK); 4 ♀♀, 20 km S Nieuwoudtville, Farm Papkuilsfontein, Fynbos [31°33’16” S, 19°08’31” E], 680 m, 27 Aug. 2010, M. Kuhlmann (SANC, RCMK); 1 ♀, van Rhynsdorp, Cape [31°36’ S, 18°44’ E], 1929, Dr. Brauns (SAMC); 1 ♀, Clanwilliam, Ramskop [32°10’ S, 18°53’ E], 24 Jul. 1985, V.B. Whitehead (SAMC).

Fig. 2. Redivivoides capensis sp. nov., ♀. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
Fig. 3. *Redivivoides capensis* sp. nov., ♂. A. Lateral view. B. Scutum and scutellum. C. Metasoma (dorsal view). D. S6 (ventral view). E. S7 (dorsal view). F. S8, apical end (backscattered electron image). G. Genitalia (dorsal view). H. Genitalia (lateral view). Scale bar: 1 mm.
Description

Female

Body length. 9.0-10.5 mm.

Head. Head slightly wider than long. Integument black except tips of mandible partly dark reddish-brown. Face sparsely covered with long, whitish-grey to brown, erect hairs intermixed with black hairs especially along the inner eye margins and on vertex (Fig. 2B). Clypeus mostly flat, apically almost impunctate; medially covered with medium-sized punctures that become gradually smaller and denser towards the lateral and upper margins; surface between punctures shiny (Fig. 2B). Malar area medially narrow, almost linear. Antenna black, ventrally orange to reddish-brown.

Mesosoma. Integument black. Mesoscutum impunctate, finely shagreened and matt (Fig. 2C). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, on the disc of mesoscutum some black hairs intermixed.

Wings. Yellowish-brown; wing venation dark brown.

Legs. Integument black to dark reddish-brown. Vestiture of femora whitish-yellow, on tibiae and tarsi dark brown to black, scopae dark brown, ventrally hairs white to greyish-yellow.

Metasoma. Integument black, except T1 to a variable extent apically red, T2 red with a black spot anterior-medially, T3 apically with a red margin (Fig. 2D). T1 completely and T2 on disc sparsely covered with a few long erect yellowish-white hairs; T3 – T4 covered with short yellow to black hairs; apical tergal hair band missing on T1, on T2 – T4 broadly white (Fig. 2A, D). Prepygidial fimbria dorsally black, laterally partly white, pygidial fimbria black. Terga impunctate, finely shagreened with a silky shine (Fig. 2D).

Male

Body length. 8.5-10.0 mm.

Head. Head slightly wider than long. Integument black except tip of mandible partly dark reddish-brown. Face sparsely covered with long, whitish-grey to brown, erect hairs intermixed with black hairs especially along the inner eye margins and on vertex. Clypeus mostly flat, apically almost impunctate; medially covered with medium-sized punctures that become gradually smaller and denser towards the lateral and upper margins; surface between punctures shiny. Malar area medially narrow, almost linear. Antenna black, ventrally reddish-brown.

Mesosoma. Integument black. Mesoscutum impunctate, finely shagreened and matt (Fig. 3B). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long white to yellowish-grey erect hairs, on the scutellum black hairs intermixed (Fig. 3A).

Wings. Slightly yellowish-brown; wing venation brown.

Legs. Integument black, tibiae and tarsi partly dark red-brown. Vestiture whitish.

Metasoma. Integument black, except T1 to a variable extend apically red, T2 red with a black spot anterior-medially, T3 either like T2 or in some specimens only apically with a red margin, T4 in some males with red basal and apical margins (Fig. 3C). T1 completely and T2 – T3 on disc sparsely covered with a few long erect yellowish-white hairs; T4 – T6 covered with short black hairs; apical tergal hair band missing on T1, on T2 – T5 narrow and sparsely white (Fig. 3C). Terga impunctate, finely shagreened with a silky shine (Fig. 3C).
TERMINALIA. Genitalia (Fig. 3G-H), S6 (Fig. 3D), S7 (Fig. 3E) and terminal plate of S8 (Fig. 3F) as illustrated.

Distribution
This species is only known from a few localities from Clanwilliam to the Nieuwoudtville area (Fig. 4).

Floral hosts
All specimens collected on the farms Papkuilsfontein and Glen Lyon were collected on Polygalaceae: *Nylandtia scoparia*.

Seasonal activity
July – September.

Fig. 4. Distribution of *Redivivooides capensis* sp. nov. and *R. karooensis* sp. nov. in South Africa.
Redivivoides eardleyi sp. nov.
Figs 5, 9
urn:lsid:zoobank.org:act:543F31E2-5A26-4D78-A07A-674CA7997F19

Diagnosis
Females of *R. eardleyi* sp. nov. can be separated from other *Redivivoides* species by a combination of the following characters: metasomal terga black to brown, T2 sparsely punctate with minute punctures and surface between punctures smooth and shiny, white apical tergal hair bands present (Fig. 5D), prepygidial and pygidial fimbria dark brown to black (Fig. 5D). The male is unknown.

Etymology
Named after Connal D. Eardley, Pretoria, who collected this species and to honour his outstanding contribution to African bee taxonomy.

Type material (3 specimens)

Holotype
♀, 13 Sep. 2011, C. Eardley (SANC). SOUTH AFRICA: N. Cape, 6 km E Pofadder, 29°06′26″ S, 19°27′13″ E.

Fig. 5. *Redivivoides eardleyi* sp. nov., ♀. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
Paratypes
SOUTH AFRICA: 1 ♀, same date as holotype (RCMK); 1 ♀, N. Cape, 21.6 km E Springbok [29°34’ S, 18°04’ E], 1006 m, 13 Sep. 2011, L. Packer (LPCT).

Description
Female
BODY LENGTH. 11.0 mm.

HEAD. Head wider than long. Integument black. Face sparsely covered with long, whitish-grey, erect hairs, along the inner eye margins and on vertex intermixed with black hairs (Fig. 5B). Clypeus convex in profile, apically impunctate; medially covered with fine punctures that become gradually smaller and denser towards the clypeal margins; surface between punctures smooth and shiny (Fig. 5B). Malar area mediately narrow, almost linear. Antenna black.

MESOSOMA. Integument black. Mesoscutal disc between punctures smooth and shiny; disc densely (i = 0.5-1.0 d) and finely punctate (Fig. 5C). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-white erect hairs, on the disc of mesoscutum intermixed with black hairs.

WINGS. Yellowish-brown; wing venation reddish brown.

LEGS. Integument black. Vestiture whitish to brown, scopae yellowish to dark brown.

METASOMA. Integument black, apical margins of T2 – T4 narrowly reddish-brown (Fig. 5D). T1 with a few long erect whitish hairs; discs of T2 – T4 sparsely covered with very short erect white to dark brown hairs; apical tergal hair bands on T1 – T4 broad and white; prepygidial and pygidial fimbriae black (Fig. 5A, D). T1 almost impunctate, polished and shiny, T2 – T4 shiny, with very fine, superficial and sparse punctuation that becomes progressively denser on apical tergae (Fig. 5D).

Male
Unknown.

Distribution
There is only a single record of this species from the Karoo in the summer rainfall area (Fig. 9).

Floral hosts
Unknown.

Seasonal activity
September.

Redivivoides kamieskroonensis sp. nov.
Figs 6, 9

urn:lsid:zoobank.org:act:C65B0CFE-D36A-4C57-BE42-7009CE0F357D

Diagnosis
Females of R. kamieskroonensis sp. nov. can be separated from other Redivivoides species by their extensively and brightly red metasomal terga (Fig. 6D) in combination with the head that is distinctly wider than long (Fig. 6B). Unlike other species the propodeal triangle is weakly shagreened. The male is unknown.
Etymology
Named after the town Kamieskroon in NW South Africa, in the vicinity of which this species was discovered.

Type material (2 specimens)

Holotype

Paratype
SOUTH AFRICA: 1 ♀, NCP, Dassiefontein Farm, 14 road km E Kamieskroon [30°09’31” S, 17°59’96” E], 760 m, 16-17 Sep. 2001 (CUIC).

Description
Female
BODY LENGTH. 12.0 mm.

Fig. 6. Redivivoides kamieskroonensis sp. nov., ♀. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
HEAD. Head much wider than long. Integument black except tips of mandible partly dark reddish-brown. Face sparsely covered with long, whitish-grey to brown, erect hairs intermixed with black hairs (Fig. 6B). Clypeus almost flat, slightly convex in profile in the lower part and slightly concave in the upper part, apico-medially and apical margin impunctate; clypeus otherwise densely (i ≤ d) covered with small punctures that become gradually smaller and denser towards the clypeal margins; surface between punctures shiny (Fig. 6B). Malar area medially about 1/4 to 1/5 as long as width of mandible base. Antenna black, ventrally reddish-brown.

MESOSOMA. Integument black. Mesoscutal disc between punctures smooth and shiny; disc densely (i = 0.5-1.0 d) and finely punctate (Fig. 6C). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish to whitish-grey erect hairs, on the disc of mesoscutum intermixed with black hairs.

WINGS. Yellowish-brown; wing venation dark reddish brown.

LEGS. Integument black, partly dark red-brown. Vestiture dark brown, scopae blackish-brown (Fig. 6A).

METASOMA. Integument mostly black, red are the apical part of T1, all of T2 except for a small black spot anterior-medially, T3 except for a large black spot in the same position and apical margin of T4 (Fig. 6D). T1 and T2 on disc with long erect yellowish-white hairs; T3 – T4 covered with mostly blackish and very short erect hairs; apical tergal hair bands on T1 – T4 medially broad, consisting of short sparse white hairs; prepygidial and pygidial fimbriae black (Fig. 6A, D). Terga impunctate, finely shagreened with a silky shine (Fig. 6D).

Male
Unknown.

Distribution
This species is only known from a single locality near Kamieskroon (Fig. 9).

Floral hosts
Unknown.

Seasonal activity
September.

Redivivoides karooensis sp. nov.
Figs 4, 7, 8
urn:lsid:zoobank.org:act:00C81E1F-78C5-406D-8132-0CC6ABD24EB9

Diagnosis
Females of R. karooensis sp. nov. can be separated from other Redivivoides species by a combination of the following characters: metasomal terga black to brown, T2 densely punctate with large punctures and surface between punctures smooth and shiny, white apical tergal hair bands present (Fig. 7D), prepygidial and pygidial fimbriae dark brown to black (Fig. 7D). These characters also apply to males (Fig. 8C) but the genitalia and S6 – S8 (Fig. 8D-H) should be checked to avoid potential confusion with the unknown male of R. eardleyi.

Etymology
Named after the Karoo, the arid region in western and central South Africa where this species was found.
Type material (22 specimens)

Holotype

Paratypes

Fig. 7. Redivivoides karooensis sp. nov., ♀. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
Description

Female

**Body Length.** 9.5-10.5 mm

**Head.** Head slightly wider than long. Integument black except mandibles and labrum largely dark reddish-brown. Face sparsely covered with long, yellowish-grey, erect hairs intermixed with black hairs especially along the inner eye margins and on vertex (Fig. 7B). Clypeus slightly convex, apically narrowly impunctate; medially densely covered with medium-sized punctures that become gradually smaller and denser towards the lateral and upper margins; surface between punctures shiny or superficially shagreened (Fig. 7B). Malar area medially narrow, almost linear. Antenna black, ventrally orange to reddish-brown.

**Mesosoma.** Integument black. Mesoscutal disc between punctures smooth and shiny; disc densely (i = 0.5–1.0 d) and finely punctate (Fig. 7C). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, black hairs intermixed on mesoscutum (Fig. 7C).

**Wings.** Yellowish-brown; wing venation brown.

**Legs.** Integument black to dark reddish-brown. Vestiture of femora whitish-yellow, on tibiae and tarsi dark brown to black, scopae very sparse, dark brown to whitish-grey.

**Metasoma.** Integument black, apical margins of terga narrowly translucent reddish-brown (Fig. 7D). T1 apically and T2 on disc sparsely covered with long erect yellowish-white hairs; T3 – T4 covered with short yellow to black hairs; apical tergal hair band missing on T1, on T2 – T4 broadly white (Fig. 7A, D). Prepygidial and pygidial fimbriae black. T1 almost impunctate, polished and shiny, T2 – T4 shiny, with very fine, superficial and sparse punctation that becomes progressively denser on apical terga (Fig. 7D).

Male

**Body Length.** 8.5-10.5 mm.

**Head.** Head slightly wider than long. Integument black except tip of mandible partly dark reddish-brown. Face densely covered with long, yellowish-brown, erect hairs intermixed with black hairs along the inner eye margins and on vertex. Malar area medially narrow, almost linear. Antenna black, ventrally yellowish to reddish-brown.

**Mesosoma.** Integument black. Mesoscutal disc between punctures smooth and shiny or superficially shagreened; disc densely (i = 0.5–1.0 d) and finely punctate (Fig. 8B). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, on scutellum black hairs intermixed.

**Wings.** Yellowish-brown; wing venation brown.

**Legs.** Integument black, tibiae and tarsi partly dark red-brown to yellowish-brown. Vestiture yellowish-brown.

**Metasoma.** Integument black, apical margins of terga narrowly translucent reddish-brown (Fig. 8C). T1 – T3 covered with long erect yellowish-white hairs; T4 – T6 covered with short black hairs; apical tergal hair band missing on T1, on T2 – T5 narrow and sparse, white (Fig. 8A, C). T1 almost impunctate, polished and shiny, T2 – T4 shiny, with very fine, superficial and sparse punctation that becomes progressively denser on apical terga (Fig. 8C).
TERMINALIA. Genitalia (Fig. 8G-H), S6 (Fig. 8D), S7 (Fig. 8E) and terminal plate of S8 (Fig. 8F) as illustrated.

Distribution
The species is known from a few places in southern Namaqualand and low lying areas west of the Cedarberg Mountains (Fig. 4).

Floral hosts
Scrophulariaceae: *Phyllopodium heterophyllum*, *Phyllopodium pumilum*, *Phyllopodium* spec., *Polycarena* spec.

Seasonal activity
July – September.

*Redivivoides namaquaensis* sp. nov.
Figs 9, 10, 11
urn:lsid:zoobank.org:act:AEF68F53-B181-4D12-8D1C-980031A57F2E

Diagnosis
Males and females of *R. namaquaensis* sp. nov. can be separated from other *Redivivoides* species by their black to brown metasomal terga in combination with dark brown to black prepygidial fimbria and orange-brown pygidial fimbria (Figs 10D, 11C).

Etymology
Named after Namaqualand, the arid region in NW South Africa where this species occurs.

Type material (71 specimens)

**Holotype**

**Paratypes**
3219BB [32°10’ S, 19°50’ E], 3 Oct. 1985, V.B. Whitehead, *Polycarena* (SAMC, RCMK); 2 ♀♂, 29.7
km NW Sutherland [32°13’ S, 20°30’ E], 27 Sep. 1985, V.B. Whitehead (SAMC); 1 ♀, Wuppertal, 2
km N of, 3219AA [32°15’ S, 19°12’ E], 29 Aug. 1990, V.B. Whitehead, *Polycarena* (SAMC); 1 ♂,
*racemosa* (SAMC); 1 ♀, Farm Kanolfontein, 3220AD [32°20’ S, 20°20’ E], 22 Sep. 1985, K. Steiner,
*Polycarena* (SAMC); 1 ♂, Piketberg, Hartebeesrivier, 3218DC [32°50’ S, 18°40’ E], 23 Aug. 1991, V.B.
Whitehead, ?*Polycarena* (SAMC); 1 ♂, Sauer, Suurfontein, 3218DC [32°50’ S, 18°40’ E], 1 Sep. 1994,

**Description**

**Female**

**Body length.** 10.0-11.5 mm.

**Head.** Head slightly wider than long. Integument black except mandible and sometimes labrum partly
dark reddish-brown. Face sparsely covered with long, whitish-grey, erect hairs intermixed with black
hairs along the inner eye margins and on vertex (Fig. 10B). Clypeus mostly flat, apical margin almost

![Distribution of Redivivoides eardleyi sp. nov., R. kamieskroonensis sp. nov. and R. namaquaensis
sp. nov. in South Africa.](image-url)
impunctate; medially densely covered with medium-sized punctures that become gradually smaller and denser towards lateral and upper margins; surface between punctures shiny (Fig. 10B). Malar area medially narrow, almost linear. Antenna black, ventrally dark orange to reddish-brown.

**Mesosoma.** Integument black. Mesoscutal disc between punctures smooth and shiny; disc densely (i = 0.5-1.0 d) and finely punctate (Fig. 10C). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, on the mesoscutum black hairs intermixed (Fig. 10C).

**Wings.** Yellowish-brown; wing venation brown.

**Legs.** Integument black to dark reddish-brown. Vestiture of femora whitish-yellow, on tibiae and tarsi dark brown to black, scopae yellowish-brown.

**Metasoma.** Integument dark brown to black, apical margins of terga narrowly translucent reddish to yellowish-brown (Fig. 10D). T1 – T4 covered with erect yellowish-white hairs, becoming progressively shorter from T1 to T4; apical tergal hair band sparse on T1, on T2 – T4 broadly white (Fig. 10A, D). Prepygidial fimbria dark brown to black, pygidial fimbria orange-brown. Terga impunctate, finely shagreened and matt (Fig. 10D).

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**Fig. 10.** *Redivivoides namaquaensis* sp. nov., ♂. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
Male

**BODY LENGTH**: 9.0-10.0 mm.

**HEAD**: Head slightly wider than long. Integument black except tip of mandible partly dark reddish-brown. Face densely covered with long, whitish-grey, erect hairs intermixed with black hairs along the inner eye margins and on vertex. Malar area mediastically narrow, almost linear. Antenna black, ventrally dark brown.

**MESOSOMA**: Integument black. Mesoscutal disc between punctures smooth or superficially shagreened and shiny; disc densely (i = 0.5 d) and very finely punctate (Fig. 11B). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long white to yellowish-grey erect hairs, partly intermixed with a few black hairs.

**WINGS**: Yellowish-brown; wing venation brown.

**LEGS**: Integument black to reddish-brown. Vestiture whitish to yellowish-brown.

**METASOMA**: Integument dark brown to black, apical margins of terga narrowly translucent reddish to yellowish-brown (Fig. 11C). T1 – T3 densely covered with long, erect yellowish-brown hairs, on T3 intermixed with black hairs; T4 – T6 covered with short black hairs; apical tergal hair band missing on T1, on T2 – T5 narrow and sparse, white; apical fringe of T7 orange (Fig. 11A, C). Terga impunctate, finely and densely shagreened, matt (Fig. 11C).

**TERMINALIA**: Genitalia (Fig. 11G-H), S6 (Fig. 11D), S7 (Fig. 11E) and terminal plate of S8 (Fig. 11F) as illustrated.

**Distribution**

This species has the centre of its distribution on the Bokkeveld Plateau and in the Roggeveld Mountains with a few records southwest of that area (Fig. 9).

**Floral hosts**

Asteraceae: *Arctotheca calendula*, *Arctotheca* spec., *Othonna* spec.; Scrophulariaceae: *Hemimeris racemosa*, *Phyllopondium heterophyllum*, *Polycarena* spec., *Selago pinguicola*, *Selago* spec..

**Seasonal activity**

July – October.

*Redivivoides simulans* Michener, 1981

Figs 12, 13, 14

*Redivivoides simulans* Michener, 1981: 121-122, figs 80, 82-84 (p. 43), 97 (p. 48), Holotype ♀ (type locality: Clanwilliam, South Africa) (Snow Entomological Museum, University of Kansas, Lawrence).

**Diagnosis**

Females of *R. simulans* can be separated from other *Redivivoides* species by a combination of the following characters: metasomal terga at least partly red (at least on a narrow stripe on the apical part of the discs of T1 – T3), metasomal terga with surface between punctures smooth and shiny (Fig. 12D), scutum at least on the disc between punctures smooth and shiny (Fig. 12C), and head about as long as wide (Fig. 12B). These characters also apply to males (Fig. 13C) but genitalia and S6 – S8 (Fig. 13D–H) should be checked to avoid potential confusion with the unknown male of *R. kamieskroonensis*.
Both sexes of *R. simulans* were described in detail by Michener (1981).

**Material examined**

Specimens examined (56 specimens): SOUTH AFRICA: 2 ♀♀, 40 km N Port Nolloth [28°54’3” S, 16°44’3” E], 11 Sep. 2001 (CUIC, RCMK); 1 ♀, 1 ♂, Skurfkop St., 3118AB [31°10’ S, 18°20’ E], 18 Aug. 1983, V.B. Whitehead, white *Hermannia* prostrate (SAMC); 1 ♀, 1 ♂, Farm Sarrisam, 3017DA [30°40’ S, 17°40’ E], 14 Aug. 1993, K. Steiner, *Hermannia* sp. (SAMC); 1 ♀, Ouberg Pass, 27 km SE Vanrhynsdorp, Fynbos [31°48’07” S, 18°55’00” E], 380 m, 9 Aug. 2011, M. Kuhlmann (RCMK); 2 ♀♀, 2 ♂♂, near Ratelfontein [31°56’ S, 18°31’ E], 21 Sep. 2007, C. Eardley (SANC); 1 ♀, Koinonia [32°01’42” S, 18°39’34” E], 24 Sep. 2011, C. Eardley (SANC); 2 ♀♀, 1 ♂, Ratelfontein / Olaf Bergfontein [32°02’ S, 18°35’ E], 7 Oct. 1995, F.W., S.K. & R.W. Gess, visiting yellow fls. semi-prostrate *Hermannia* sp., Sterculiaceae, 95/96/273 (AMGS); 2 ♀♀, Trawal, 15 km SSW [32°02’ S, 18°35’ E], 21 Sep. 2007, T.L. Griswold (EMUS); 1 ♀, 1 ♂, S Skurfkop [32°03’26” S, 18°40’27” E], 24 Sep. 2011, C. Eardley (SANC); 1 ♀, Lamberts Bay [32°04’21” S, 18°22’12” E], 160 m, 13 Sep. 2005, C. Eardley (SANC); 5 ♀♀, 1 ♂, 5 km N Clanwilliam [32°08’1” S, 18°51’3” E], 94 m, 20 Sep. 2001 (CUIC); 2 ♀♀, 11 km N Clanwilliam [32°05’08” S, 18°49’25” E], 21 Sep. 2011, C. Eardley (SANC); 1 ♀, 20 mi. W. of Clanwilliam [32°09’ S, 18°36’ E], 19 Sep. 1966, C.D. Michener (SAMC); 1 ♂, 9 km W Clanwilliam [32°10’11” S, 18°48’21” E], 22 Sep. 2011, C. Eardley (SANC); 1 ♀, 1 ♂, Clanwilliam

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**Fig. 12.** *Redivivoides simulans* Michener, 1981, ♀. A. Lateral view. B. Head. C. Scutum and scutellum. D. Metasoma (dorsal view). Scale bar: 1 mm.
Additional material

Additional records from the database of the South African Museum, Cape Town (material in SAMC, no data on number or sex of specimens available): SOUTH AFRICA: Goegap Reserve, 4 km from tarred road on circular drive, Springbok [29°40′ S, 18°00′ E], 13 Aug. 1993, Whitehead, *Zygophyllum*;
Distribution

This is the most common and widespread of all Redivivoides species and has been found in most parts of the winter rainfall area (Fig. 14).
Floral hosts

Seasonal activity
(June) July – October.

*Redivivoides variabilis* sp. nov.
Figs 15, 16, 17

Diagnosis
Females and males of *R. variabilis* can be separated from other *Redivivoides* species by their black to brown metasomal terga which are smooth and shiny between punctures and that lack white apical hair bands (Figs 15E-F, 16E). This is the only *Redivivoides* species where terga have an intense oily bluish shine.

Etymology
The species is named after the colour variation of the females.

Type material (31 specimens)

**Holotype**
SOUTH AFRICA: N.C. Prov., Grootvlei, 3017BB, 17°50’E, 30°10’S.

**Paratypes**
SOUTH AFRICA: 1 ♀, 1 ♂ same date as holotype (SAMC); 1 ♀, 1 ♂, Springbok [29°40’ S, 17°52’ E], 7 Sep. 1966, C.D. Michener (SAMC); 3 ♀♂,Namaqualand, Springbok, Goegap Nature Res. (hills) [29°41’ S, 17°59’ E], 8-10 Sep. 1992, F.W. & S.K. Gess, on yellow fls. of *Lebeckia spinescens* Harv. (Papilionaceae), 92/93/41, (AMGS); 2 ♀♂, Dassiefontein Farm, 14 km E Kammsiekroon (sic) 30°09’ S, 17°59’ E], 16-17 Sep. 2001, C. Eardley (SANC); 1 ♀, Skilpad, 3017BB [30°10’ S, 17°50’ E], 19 Aug. 1999, K. Steiner, *Nemesia anisocarpa* (SAMC); 1 ♀, Kamieskroon, Dassiefontein, 3017BB [30°10’ S, 17°50’ E], 7 Sep. 1986, K. Steiner, *Lotononis* (SAMC); 1 ♀, Farm Grootvlei, 3017BB [30°10’ S, 17°50’ E], 24 Aug. 1990, K. Steiner, Legume (SAMC); 1 ♂, Grootvlei, 20 km S Kamieskroon [30°10’ S, 17°50’ E], 10 Aug. 1984, V.B. Whitehead, yellow legume (SAMC); 2 ♀♂, 8 km E Karkams, 3017BD [30°20’ S, 17°50’ E], 10 Aug. 1985, V.B. Whitehead, *Nemesia & Lachenalia* (SAMC); 1 ♀, 6.2 km NE of Karkams, 3017BD [30°20’ S, 17°50’ E], 4 Sep. 1986, K. Steiner, *Nemesia* (SAMC); 1 ♀, Garies, Welkom Farm, 3018CA [30°40’ S, 18°10’ E], 30 Sep. 1988, V.B. Whitehead, *Lotononis serpens* (SAMC); 1 ♀, N. Cape, Nieuwoudtville, Flower Reserve East [31°21’56” S, 19°08’52” E], 735 m, 10 Aug. 2004, M. Kuhlmann (RCMK); 1 ♀ (black form), N. Cape, Nieuwoudtville, Flower Reserve East [31°21’56” S, 19°08’52” E], 735 m, 6 Sep. 2002, M. Kuhlmann (RCMK); 1 ♀ (black form), 1 ♂, N. Cape, Nieuwoudtville, Flower Reserve East [31°21’56” S, 19°08’52” E], 735 m, 3 Sep. 2003, M. Kuhlmann, on *Oxalis* spec. (RCMK); 1 ♀, Nieuwoudtville, Farm Glenlyon, 3117AC [31°21’ S, 19°08’ E], 14 Aug. 1997, V.B. Whitehead, *Lotononis serpens* (SAMC); 1 ♂, Nieuwoudtville Flower Reserve [31°22’2” S, 19°08’ E], 2 Aug. 1984, V.B. Whitehead, *Lotononis serpens* (SAMC); 2 ♀♂, W. Cape, 24 km N of Garies [30°23’ S, 17°54’ E], 777 m, 25 Sep. 2011, L. Packer (LPCT, RCMK); 1 ♂, N. Cape, Nieuwoudtville, Glen
Description

Female

Body length. 12.0-13.0 mm.

Head. Head slightly wider than long. Integument black except median part of mandible and sometimes labrum partly dark reddish-brown. Face sparsely covered with long, whitish-grey (in the dark form all black, Fig. 15B), erect hairs intermixed with black hairs along the inner eye margins and on vertex...
Clypeus mostly flat, apically almost impunctate, punctures here large and scattered becoming rapidly smaller and denser towards the upper edge; surface between punctures shiny (Fig. 15C). Malar area medially narrow, almost linear. Antenna black, ventrally sometimes partly dark reddish-brown.

**Mesosoma.** Integument black, tegulae partly yellowish-brown. Mesoscutal disc between punctures smooth and shiny; disc densely \((i = 0.5–1.0 \ d)\) and finely punctate (Fig. 15D). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, on the disc of mesoscutum and scutellum some black hairs intermixed (in the dark form all black, Fig. 15B).

**Wings.** Yellowish-brown; wing venation brown.

**Legs.** Integument black to yellowish-brown. Vestiture yellowish-brown, scopae yellowish-brown. In the dark form hairs of femora and most of tibiae black or dark brown, on tarsi yellowish-brown, scopae darker yellowish-brown (Fig. 15B).

**Metasoma.** Integument black, except apical tergal margins partly narrowly brownish translucent; terga with intense oily bluish shine (Fig. 15E-F). T1 apically sparsely covered with a few long erect yellowish hairs; T2 – T4 covered with short yellowish-white hairs; apical tergal hair band missing on T1, on T2 – T4 very sparse and only laterally developed (Fig. 15A-E). In the dark form all hairs black, no apical tergal hair bands (Fig. 15B-F). Prepygidial fimbria dorsally black, laterally partly yellowish white (in dark form all black), pygidial fimbria black. Terga densely but finely punctate, smooth and shiny between punctures (Fig. 15E-F).

**Male**

**Body length.** 12.0–13.0 mm.

**Head.** Head slightly wider than long. Integument black except tip of mandible partly dark reddish-brown. Face densely covered with long, whitish-grey to yellowish-brown, erect hairs intermixed with black hairs along the inner eye margins and on vertex. Malar area medially narrow, almost linear. Antenna black, ventrally dark reddish-brown.

**Mesosoma.** Integument black, tegulae partly yellowish-brown. Mesoscutal disc between punctures smooth and shiny; disc densely \((i = 0.5–1.0 \ d)\) and finely punctate (Fig. 16B). Mesoscutum, scutellum, metanotum, mesepisternum and propodeum covered with long yellowish-brown erect hairs, on the disc of mesoscutum and scutellum some black hairs intermixed (Fig. 16B).

**Wings.** Yellowish-brown; wing venation brown.

**Legs.** Integument black, tibiae and tarsi partly yellowish-brown. Vestiture yellowish-white.

**Metasoma.** Integument black, except apical tergal margins partly narrowly brownish translucent; T1 – T3 with intense oily bluish shine, T4 – T5 less so (Fig. 16A, C). T1 completely and T2 on disc covered with long erect yellowish-white hairs; T2 – T3 densely covered with short erect yellowish-white either yellowish-white or black on T4 and black on T5 – T6; apical tergal hair band missing on T1, on T2 – T5 sparse and narrow, yellowish-white to white (Fig. 16A, C). Terga densely but finely punctate, smooth and shiny between punctures (Fig. 16C). T7 with pygidial plate that is reduced to a narrow longitudinal, slightly elevated and shiny ridge.

**Terminalia.** Genitalia (Fig. 16G-H), S6 (Fig. 16D), S7 (Fig. 16E) and terminal plate of S8 (Fig. 16F) as illustrated.
Distribution

The species has been found in most parts of Namaqualand to the northern parts of the Roggeveld Mountains in the southeast (Fig. 17). The colour form with females all black is only known from the higher grounds of the south-eastern part of the species' range while the females with light brown pilosity mostly occur in lower areas and the north (Fig. 17). Colour variation is also known from Rediviva gigas Whitehead & Steiner, 1993 and R. parva Whitehead & Steiner, 2001. In R. gigas there is no obvious geographical pattern of colour variation but the black form of R. parva seems to be concentrated in the north and in lower lying areas in the west of its range (Whitehead & Steiner 2001).

Floral hosts


Seasonal activity

August – September.

Key to Redivivoides species

Females

1. Metasomal terga extensively and brightly red (Figs 2D, 6D, 12D), rarely only on a narrow stripe of the apical part of the discs of T1 – T3 ................................................................. 2
   – Metasomal terga black or brown (Figs 5D, 7D, 10D, 15E-F), sometimes apical tergal margins narrowly translucent dark reddish brown ................................................................. 4

2. Scutum impunctate, shagreened and matt (Fig. 2C) ........................................ R. capensis sp. nov.
   – Scutum finely punctate and at least on disc between punctures smooth and shiny (Figs 6C, 12C) … 3

3. Metasomal terga finely shagreened and with a dull shine (Fig. 6D); head distinctly wider than long (Fig. 6B) ............................................................................................... R. kamieskroonensis sp. nov.
   – Metasomal terga between punctures smooth and shiny (Fig. 12D); head only slightly wider than long (Fig. 12B) ................................................................................ R. simulans Michener, 1981

4. Metasomal terga finely shagreened and dull (Fig. 10D); prepygidial and pygidial fimbria yellowish brown (Fig. 10D) ................................................................. R. namaquaensis sp. nov.
   – Metasomal terga between punctures smooth and shiny (Figs 5D, 7D, 15E-F); prepygidial and pygidial fimbria dark brown to black (Figs 5D, 7D, 15E-F) ........................................... 5

5. Metasomal terga without white apical tergal hair bands (Fig. 15E-F); terga with intense oily bluish shine ................................................................. R. variabilis sp. nov.
   – Metasomal terga with white apical tergal hair bands (Figs 5D, 7D); terga without oily bluish shine … 6

6. Disc of T2 sparsely punctate (i = 2-3 d); punctures minute (Fig. 5D); head and clypeus as in Fig. 5B ............................................................................................... R. eardleyi sp. nov.
   – Disc of T1 densely punctate (i = d); punctures larger (Fig. 7D); head and clypeus as in Fig. 7B ................................................................................ R. karooensis sp. nov.
Males

The males of *R. eardleyi* sp. nov. and *R. kamieskroonensis* sp. nov. are unknown.

1. Metasomal terga extensively and brightly red, rarely only on a narrow stripe of the apical part of the discs of T1 – T3 (Figs 3C, 13C) ............................................................. 2
   – Metasomal terga black or brown, sometimes apical tergal margins narrowly translucent dark reddish brown (Figs 8C, 11C, 16C) ............................................................. 3

2. Scutum impunctate, shagreened and matt (Fig. 3B); S6 – S8 and genitalia as in Fig. 3D-H ......................... *R. capensis* sp. nov.
   – Scutum finely punctate and at least on disc between punctures smooth and shiny (Fig. 13B); S6 – S8 and genitalia as in Fig. 13D-H ............................................. *R. simulans* Michener, 1981

3. Metasomal terga finely shagreened and dull (Fig. 11C); prepygidial and pygidial fimbria yellowish brown (Fig. 11C); S6 – S8 and genitalia as in Fig. 11D-H ...................... *R. namaquaensis* sp. nov.
   – Metasomal terga between punctures smooth and shiny at least on T1 – T2 (Figs 8C, 16C); prepygidial and pygidial fimbria brown to black (Figs 8C, 16C); S6 – S8 and genitalia different .... 4

4. Metasomal terga with oily bluish shine; S6 – S8 and genitalia as in Fig. 16D-H ........................................ *R. variabilis* sp. nov.
   – Metasomal terga without oily bluish shine; S6 – S8 and genitalia as in Fig. 8D-H ................................. *R. karooensis* sp. nov.

Discussion

Redefinition of the genus *Redivivoides*

Within the Melittinae Michéz *et al.* (2009), based on a phylogenetic study, recognize two tribes; the Macropidini comprises genera with two submarginal cells (*Afrodasypoda, Macropis, Promelitta* and the fossil taxa *Eomacropis* and *Paleomacropis*) and the Melittini, comprising genera with three submarginal cells (*Melitta, Rediviva, Redivivoides*). In the Melittini *Melitta* is characterized by a well-defined, large and dull propodeal triangle (except *Melitta schultzei*; Michéz & Eardley 2007) while in *Rediviva* and *Redivivoides* this is ill-defined, small and shiny, although in some species superficially sculptured.

In both *Macropis* (Fig. 1A-B) and *Rediviva* (Fig. 1C-D) the females can easily be recognized by their broadened hind tibia and basitarsus with the scopa consisting of densely plumose, velvety pubescence for transporting floral oil, while the elongate fore tarsi (extreme in some species) are a synapomorphy of *Rediviva* (Michener 1981, 2007). In contrast the females of *Redivivoides* have short front legs with simple hairs and a slender hind tibia, as in *Melitta* (Michener 1981). The scopa consists of sparse simple bristles (more dense and usually with at least a few shortly branched hairs in *Melitta*; Fig. 1G) with the anterior margin of the scopa shining, unlike in other genera (Fig. 1E) (Michener 1981). The hind basitarsus of *Redivivoides* females is slightly broadened, about 2-2.5 times as long as its maximum width with the posterior margin curved and without an apical process (Fig. 1F) while in *Melitta* it is slender, three or more times as long as broad, parallel-sided and with a small apical process (Fig. 1H) (Michener 1981). In *Redivivoides* a penicillus is generally missing while it is present in all *Melitta* species known to me and modified into a brush-like structure used for lining the wall of brood cells in *Rediviva* and *Macropis* (Schäffler & Dötterl 2011).

As in females, male *Melitta* can be recognized by the large and dull propodeal triangle (except *Melitta schultzei*) (ill-defined, small and shiny in *Rediviva* and *Redivivoides*), S7 having a large plate that is truncate or emarginate apically (sometimes with hair patches at posterior lateral angles) and with lobes
or processes reduced to small sclerotic structures like in *Melitta budensis* (Michez & Eardley 2007). In *Rediviva* and *Redivivoides* S7 in turn has a small disc with the apex bifid and/or with membranous lobes and the gonostyli are relatively long (Michener 1981, 2007).

The differentiation of *Rediviva* and *Redivivoides* males is problematic. Michener (1981, 2007) suggested that the broad, subtruncate apex of the male S7, with its large laterally attached, vertical, membranous hairy lobes (Fig. 13E) is a unique apomorphy of *Redivivoides*. However, these characters seem to be species specific for *R. simulans* and they are either absent (*R. karooensis, R. namaquaensis*) (Figs 8E, 11E) or reduced (*R. capensis, R. variabilis*) (Figs 3E, 16E) in the other four species (the males of *R. eardleyi* and *R. kamieskroonensis* are unknown). Michener (1981) mentioned the weak pygidial plate in male *R. simulans* as a character differentiating *Redivivoides* from both *Melitta* and *Rediviva*. A pygidial plate is also present in male *Macropis* and in *R. variabilis*, where it is reduced to a narrow longitudinal, slightly elevated and shiny ridge but it is missing in all the other three species making it unsuitable as a differentiating character.

Generally, the morphology of S7, S8 and the genitalia of *Redivivoides* males (Figs 3, 8, 11, 13, 16) reflect much of the morphological diversity known from the much more species-rich *Rediviva* as illustrated in Whitehead & Steiner (2001) and Whitehead et al. (2008). Although well characterized as species, the obvious lack of apomorph characters defining the males of *Redivivoides* at the genus level make their identification difficult and indicate the close relationship of both genera as suggested by Michener (1981) and Michez et al. (2009).

**Origin of oil-collecting**

The origin of oil-collecting in bees has been much debated and this behaviour has evolved independently at least five times in the Centridini, Ctenoplectrini, Exomalopsini, Tetrapediini and Melittinae (Buchmann 1987; Vogel 1974, 1986, 1990; Renner & Schaefer 2010). Within the Melittinae floral oil is collected by bees of the genera *Macropis* and *Rediviva* (Michener 2007). Melittids are the most basal bees (Danforth et al. 2006) and, thus, they can be a key-group for understanding the evolution of oil-collecting in bees in general. Michener (1981) and later Michez et al. (2009), based on the results of a phylogenetic study, discussed whether oil-collecting has evolved only once in melittids or if there are two independent origins in *Macropis* and *Rediviva*.

In both scenarios the phylogenetic position of *Redivivoides* is crucial to understanding the evolutionary history of early bees. In Michez et al.’s (2009) analysis *Redivivoides* arose from *Rediviva*, making the latter paraphyletic, indicating a loss of oil-collecting in *Redivivoides*. However, only two of the currently 24 described *Rediviva* species and only *R. simulans* of the now seven known *Redivivoides* species were included in the study. A broader approach including more species is required to confirm the paraphyly of *Rediviva*.

If *Redivivoides* evolved from *Rediviva* the question is why *Redivivoides* is only present in the winter rainfall region in the west of South Africa whereas *Rediviva* also occurs in the summer rainfall area in the east of the country. Either *Redivivoides* evolved in the winter rainfall region and has not spread to the east of South Africa or it became extinct there. An interesting parallel case is the 11 of 35 described species of the *Colletes fasciatus*-group in western South Africa that have a reduced scopa, which is unique in the genus (Kuhlmann 2006, 2007).

Based on the present taxonomic revision there is now additional evidence that *Redivivoides* species might have lost their oil-collecting structures, as presumably happened in some Neotropical *Centris* species (Michener 2007). With the exception of *Redivivoides* all non oil-collecting genera of the Melittinae have a slender hind basitarsus and a dense scopa with at least some branched long hairs. In contrast, the
hind tibia and basitarsus of oil-collecting *Macropis* and *Rediviva* is very broad and the scopa consists of densely plumose, short pilosity for oil-collecting embedded in a matrix of sparse, long, simple, emergent hairs (Fig. 1A-D) (Michener 1981), with the latter reminiscent of *Redivivoides* (Fig. 1E-F). Thus, the broadened and posteriorly curved hind basitarsus lacking an apical process (present in *Melitta*, *Macropis* and in modified form in some *Rediviva*), the sparse scopa consisting of only simple hairs in *Redivivoides* and the lack of a penicillus (present in *Melitta* and modified into a brush-like structure in *Rediviva* and *Macropis*) might be remnants indicating a loss of oil-collecting structures. This view is supported by the phylogenetic position of *R. simulans*, arising from *Rediviva*, rendering *Rediviva* paraphyletic (Michez et al. 2009), the similar structure of male genitalia, S7 and S8 and a study of fore and mid leg anatomy by Steiner & Cruz (2006) suggesting that *Redivivoides* has descended from an oil-collecting ancestor.

The close relationship of *Redivivoides* and *Rediviva* is also reflected in flower visitation. Most *Rediviva* collect floral oil on Scrophulariaceae (Whitehead & Steiner 2001; Whitehead et al. 2008) and pollen from this plant family is also a major constituent of female scopal pollen loads (Kuhlmann unpubl.) but it is rarely used by other Melittidae (Michez et al. 2008). Whilst no information is available on flower visitation of *R. eardleyi* sp. nov. and *R. kamieskroonensis* sp. nov. and only a single observation for *R. capensis* sp. nov. all the other *Redivivoides* species were found visiting Scrophulariaceae with *R. karooensis* sp. nov. and *R. namaquaensis* sp. nov. obviously having a preference for this family. Although Kuhlmann & Eardley (2012) showed that Scrophulariaceae are visited and their pollen collected by a range of bee species it is usually only a minor component in scopal pollen loads (Kuhlmann unpubl.). Thus, the shared preference for Scrophulariaceae even though only based on a small number of observations can be seen as another indicator for the common ancestry of both genera.

However, a robust phylogeny including more species of *Rediviva* and *Redivivoides* is required to better understand the evolution of oil-collecting and phylogeography of both genera.

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References


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