## 0625

Description of two new Bolivian Cerambycidae and a Costa Rican record for an African prionine (Coleoptera)

James E. Wappes
American Coleoptera Museum 8734 Paisano Pass
San Antonio, TX 78255-3523, USA
Antonio Santos-Silva
Museu de Zoologia
Universidade de São Paulo
Caixa Postal 42.494, 04218-970, São Paulo, SP, Brazil

Maria Helena M. Galileo<br>Museu Anchieta de Ciências Naturais<br>Porto Alegre, RS, Brazil

Date of issue: April 27, 2018

# James E. Wappes, Antonio Santos-Silva and Maria Helena M. Galileo <br> Description of two new Bolivian Cerambycidae and a Costa Rican record for an <br> African prionine (Coleoptera) <br> Insecta Mundi 0625: 1-9 <br> ZooBank Registered: urn:lsid:zoobank.org:pub:18792ACD-91B9-42FC-9CE3-B113BC777585 

## Published in 2018 by

Center for Systematic Entomology, Inc.
P.O. Box 141874

Gainesville, FL 32614-1874 USA
http://centerforsystematicentomology.org/
Insecta Mundi is a journal primarily devoted to insect systematics, but articles can be published on any non-marine arthropod. Topics considered for publication include systematics, taxonomy, nomenclature, checklists, faunal works, and natural history. Insecta Mundi will not consider works in the applied sciences (i.e. medical entomology, pest control research, etc.), and no longer publishes book reviews or editorials. Insecta Mundi publishes original research or discoveries in an inexpensive and timely manner, distributing them free via open access on the internet on the date of publication.

Insecta Mundi is referenced or abstracted by several sources, including the Zoological Record and CAB Abstracts. Insecta Mundi is published irregularly throughout the year, with completed manuscripts assigned an individual number. Manuscripts must be peer reviewed prior to submission, after which they are reviewed by the editorial board to ensure quality. One author of each submitted manuscript must be a current member of the Center for Systematic Entomology.

Guidelines and requirements for the preparation of manuscripts are available on the Insecta Mundi website at http://centerforsystematicentomology.org/insectamundi/

Chief Editor: David Plotkin, insectamundi@gmail.com
Assistant Editor: Paul E. Skelley, insectamundi@gmail.com
Head Layout Editor: Robert G. Forsyth
Editorial Board: J. H. Frank, M. J. Paulsen, Michael C. Thomas
Review Editors: Listed on the Insecta Mundi webpage
Printed copies (ISSN 0749-6737) annually deposited in libraries
CSIRO, Canberra, ACT, Australia
Museu de Zoologia, São Paulo, Brazil
Agriculture and Agrifood Canada, Ottawa, ON, Canada
The Natural History Museum, London, UK
Muzeum i Instytut Zoologii PAN, Warsaw, Poland
National Taiwan University, Taipei, Taiwan
California Academy of Sciences, San Francisco, CA, USA
Florida Department of Agriculture and Consumer Services, Gainesville, FL, USA
Field Museum of Natural History, Chicago, IL, USA
National Museum of Natural History, Smithsonian Institution, Washington, DC, USA
Zoological Institute of Russian Academy of Sciences, Saint-Petersburg, Russia
Electronic copies (Online ISSN 1942-1354, CDROM ISSN 1942-1362) in PDF format
Printed CD or DVD mailed to all members at end of year. Archived digitally by Portico.
Florida Virtual Campus: http://purl.fcla.edu/fcla/insectamundi
University of Nebraska-Lincoln, Digital Commons: http://digitalcommons.unl.edu/insectamundi/
Goethe-Universität, Frankfurt am Main: http://nbn-resolving.de/urn/resolver.pl?urn:nbn:de:hebis:30:3-135240
Copyright held by the author(s). This is an open access article distributed under the terms of the Creative Commons, Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited. http://creativecommons.org/licenses/by-nc/3.0/

Layout Editor for this article: Robert G. Forsyth

# Description of two new Bolivian Cerambycidae and a Costa Rican record for an African prionine (Coleoptera) 

James E. Wappes<br>American Coleoptera Museum<br>8734 Paisano Pass<br>San Antonio, TX 78255-3523, USA<br>wappes@earthlink.net

Antonio Santos-Silva
Museu de Zoologia
Universidade de São Paulo
Caixa Postal 42.494, 04218-970, São Paulo, SP, Brazil
toncriss@uol.com.br
Maria Helena M. Galileo
Museu Anchieta de Ciências Naturais
Porto Alegre, RS, Brazil
galileomh@yahoo.com


#### Abstract

Two new species of Cerambycidae (Coleoptera) are described from Bolivia: Compsibidion woodleyi Wappes, Santos-Silva and Galileo (Cerambycinae: Neoibidionina): and Drycothaea dozieri Wappes, Santos-Silva and Galileo (Lamiinae: Calliini). Illustrations of the new species are included. Mallodon downesii Hope, 1843 (Prioninae: Macrotomini) is reported for the first time in Costa Rica.


Key words. Bolivia, key, Prioninae, South America, taxonomy.

## Introduction

Martins (1967, 1970) characterized his Division V (currently Compsina) of Ibidionini (currently Neoibidionini) as having procoxal cavities closed behind, with his Division IV (currently Neoibidionina) defined as having procoxal cavities open behind. Oftentimes, the procoxal cavity in some specimens of Neoibidionina is only slightly open appearing closed, and hence leads to improper placement of species.

This is also true in Compsibidion Thomson, 1864 (Neoibidionina), a genus with some of its species having features shared with Heterachthes (Compsina), especially the basal antennomeres thickened and not carinate in males. According to Martins and Galileo (2007) (translated): "Compsibidion includes a great miscellany of species that, for now, must remain together. A division of Compsibidion into new genera, in addition to being based on very variable features, would be premature." Actually, the separation of some species currently placed in Compsibidion from those of Heterachthes is problematic. Although Compsibidion woodleyi Wappes, Santos-Silva and Galileo has the procoxal cavities open posteriorly, they are not widely open and hence we prefer to include it in Compsibidion. Currently, Compsibidion includes 79 species distributed from Mexico to southern South America (Monné 2018a).

Drycothaea Thomson, 1868 is the second largest genus of Calliini, with 27 species distributed from Mexico to southern South America (Monné 2018b). Martins and Galileo (1990) presented a key to South American species of Drycothaea, which included five species. However, Galileo and Martins (1991) transferred several species to Drycothaea, including South American species, making the key obsolete. Later, Galileo and Martins (2010) published the most recent key to species of Drycothaea. The new species, Drycothaea dozieri Wappes, Santos-Silva and Galileo, is the fourth Drycothaea species formally recorded for Bolivia.

Mallodon downesii Hope, 1843, a Prioninae species known as a coffee pest in Africa (Delahaye and Tavakilian 2009) has been found in Costa Rica. Coffee is one of the main agricultural crops in Costa Rica and the introduction of this species into the country is of real concern and a potential problem for coffee growers.

## Materials and Methods

Photographs were taken with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65mm f/2.8 1-5× macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken using measuring ocular Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens.

The collection acronyms used in this study are as follows:
FSCA - Florida State Collection of Arthropods, Gainesville, FL, USA
INBio - Instituto Nacional de Biodiversidad, Santo Domingo de Heredia, Costa Rica
MZSP - Museu de Zoologia, Universidade de São Paulo, São Paulo, São Paulo, Brazil

## Taxonomy

## Cerambycinae

## Neoibidionini

## Neoibidionina

## Compsibidion woodleyi Wappes, Santos-Silva and Galileo, new species

(Fig. 1-4)
Description. Male. Dorsal side of head and pronotum black; ventral side of head dark brown, almost black, except yellowish-brown centrobasal area; scape dark brown, almost black; antennomeres dark brown, gradually lighter toward distal segments; ventral side of prothorax dark brown; ventral side of meso- and metathorax mostly reddish-brown, darkened on some areas, gradually lighter toward apex of metaventrite. Elytra black, gradually brown after middle, with small, oblique pale-yellow macula on about end of basal third; coxae mostly reddish-brown; femoral peduncle light reddish-brown, and femoral club brown; tibiae blackish basally, dark reddish-brown on remaining surface; tarsi brown on tarsomeres I-II, yellowish-brown on remaining tarsomeres; abdominal ventrites brown, gradually lighter toward ventrite V, except narrow, transverse yellowish band at apex of ventrites I-IV.
Head. Frontal plate longitudinally depressed centrally; finely punctate centrally, slightly transversely striate laterally; remaining surface of frons finely vermiculate centrally, rugose-punctate laterally; with short, sparse, decumbent pale-yellow setae. Area between antennal tubercles and upper eye lobes rugose-punctate; area between apex of upper eye lobes and prothorax finely, moderately sparsely punctate; with short, sparse, decumbent pale-yellow setae, and a few long, erect pale-yellow setae between upper eye lobes. Area behind upper eye lobes finely, moderately sparsely punctate (less so toward lower eye lobe); with a few long, erect pale-yellow setae close to eyes, glabrous on remaining surface. Area behind lower eye lobes very finely, sparsely punctate; with long, erect pale-yellow setae close to eye, glabrous on remaining surface. Genae minutely punctate close to lower eye lobe, smooth toward apex; with short, decumbent pale-yellow setae except on glabrous distal area. Antennal tubercles rugosepunctate frontally, finely, sparsely punctate on remaining surface, except nearly smooth apex; with short, decumbent, sparse pale-yellow setae. Median groove distinct from frontal plate to area between upper eye lobes. Gulamentum smooth and glabrous between prothorax and eyes, coarsely, moderately abundantly punctate on remaining surface (less so centrally); with short, sparse, decumbent pale-yellow setae interspersed with long, erect pale-yellow setae. Distance between upper eye lobes 0.51 times length of scape; in frontal view, distance between lower eye lobes 0.79 times length of scape; upper eye lobes with four rows of ommatidia. Antennae 2.0 times elytral length, reaching elytral apex at basal third of antennomere IX. Scape basally curved in lateral view, gradually widened toward apex; with short, sparse, decumbent pale-yellow setae interspersed with long, erect pale-yellow setae. Antennomere III tumid, not carinate dorsally, with decumbent, sparse pale-yellow setae interspersed with long, erect pale-yellow setae; antennomere IV tumid, slightly narrower than III, not carinate, with setae as on III; remaining


Figures 1-10. Illustrations of Bolivian Cerambycidae. 1-4) Compsibidion woodleyi new species, holotype male. 1) Dorsal habitus. 2) Ventral habitus. 3) Lateral habitus. 4) Head, frontal view. 5-9) Drycothaea dozieri new species, holotype female. 5) Head, frontal view. 6) Mesosternal process. 7) Dorsal habitus. 8) Ventral habitus. 9) Lateral habitus. 10) Drycothaea viridescens, male, mesosternal process.
antennomeres slender, especially from VI, with pale-yellow pubescence not obscuring integument, and long, erect pale-yellow setae at apex. Antennal formula (ratio) based on antennomere III: scape $=0.67$; pedicel $=0.14 ; \mathrm{IV}=0.63 ; \mathrm{V}=0.79 ; \mathrm{VI}=0.86 ; \mathrm{VII}=0.81 ; \mathrm{VIII}=0.77 ; \mathrm{IX}=0.75 ; \mathrm{X}=0.71 ; \mathrm{XI}=0.88$.
Thorax. Prothorax subcylindrical, slightly rounded laterally, with basal constriction distinct, 1.2 times longer than wide. Pronotum with slightly elevated central gibbosity, between basal constriction and middle; coarsely, partially confluently punctate on each side of central gibbosity, finely, sparsely punctate on area of basal constriction, smooth on central gibbosity, and nearly smooth on remaining surface; with pale-yellow pubescence in area of basal constriction, sides of basal half, nearly all area close to central gibbosity (slightly sparser than basally), and wide transverse band on each side of distal half; remaining surface with very sparse, decumbent pale-yellow setae except glabrous central gibbosity; with long, erect, sparse pale-yellow setae. Sides of prothorax nearly smooth except a few coarse punctures centrally near pronotum, and finely striate distal area; with pale-yellow pubescence near pronotum, gradually shorter, less conspicuous toward ventral side, except striate area with sparse pubescence; with long, erect, sparse pale-yellow setae near pronotum. Prosternum with pale-yellow pubescence in basal half, with long, decumbent pale-yellow setae interspersed; remaining surface nearly glabrous except sparse, erect, long pale-yellow setae. Prosternal process centrally about $1 / 4$ width of procoxal cavity. Procoxal cavities slightly open posteriorly. Metaventrite with sparse pale-yellow pubescence centrally, slightly denser laterally. Mesoventral process somewhat tab-shaped at sides of distal area, distinctly emarginate centrally at apex. Mesanepisternum, mesepimeron, and metanepisternum with moderately dense paleyellow pubescence not obscuring integument. Metaventrite with coarse, sparse, shallow punctures on sides of basal third; with pale-yellow pubescence not obscuring integument, distinctly sparser centrally; with long, erect, sparse pale-yellow setae. Scutellum with pale-yellow pubescence in distal half, interspersed with a few long, nearly decumbent golden setae. Elytra. Surface very slightly rugose; minutely, sparsely punctate; with coarse, sparse punctures from which emerges a long, erect golden seta (sub-aligned in basal half); with minute, sparse, decumbent pale-yellow setae; apex rounded at outer side, somewhat rounded projection at sutural angle. Legs. Femora clavate; with pale-yellow pubescence not obscuring integument, with long, erect, sparse pale-yellow setae interspersed. Tibiae slightly, gradually widened toward apex; with pale-yellow pubescence not obscuring integument, with long, erect pale-yellow setae interspersed. Metatarsomere I about as long as II-III together.

Abdomen. Ventrites with yellowish-white pubescence not obscuring integument, slightly denser laterally, interspersed with long erect, sparse pale-yellow setae, except in glabrous yellowish area of ventrites I-IV. Apex of ventrite V, widely, slightly emarginate centrally.
Dimensions in mm (holotype male). Total length 7.00; prothoracic length 1.60; basal prothoracic width 1.20 ; distal prothoracic width 1.20 ; max. prothoracic width 1.30 ; humeral width 1.65 ; elytral length 4.65 .

Type material. Holotype male from BOLIVIA, Santa Cruz: Refugio Los Volcanes ( 4 km N Bermejo; $18^{\circ} 10^{\prime} \mathrm{S} / 63^{\circ} 60^{\prime} \mathrm{W} ; 1,045-1,350 \mathrm{~m}$ ), 31.X.2013, J. Wappes \& K. Kuckartz col. (FSCA).
Etymology. Named for Norm Woodley, Hereford, AZ, who has so kindly assisted the authors with numerous reviews, consultation on difficult taxonomic problems and for his valued friendship.
Remarks. Compsibidion woodleyi sp. n. shares the characters of thick antennomere III, which is not carinate, with a number of other species of Compsibidion. However, it differs from them as follows: 1. C. angulare (Thomson, 1867) - outer angle of elytral apex unarmed, and pronotum lacking pubescent bands (C. angulare has elytral apex with outer angle spined and pubescent pronotal bands); 2. C. balium Napp and Martins, 1985 - general color dark brown, almost black, outer elytral apex unarmed (C. balium is generally colored orangish-yellow and the outer elytral apex has a long spine); 3. C. clivum Martins, 1971 - thicker body, antennomere III widened from near the base, then parallel-sided, and elytral apex distinctly wider (C. clivum has body slender, antennomere III gradually widened toward apex, and elytral apex distinctly narrower); 4. C. concisum Napp and Martins, 1985 - base and apex of the elytra similarly colored, and elytral apex unarmed (C. concisum has base and apex of the elytra with distinctly different color, and elytral apex with outer spine); 5. C. decemmaculatum (Martins, 1960) - antennomere IV thick, elytral setae not distinctly thick, elytral apex unarmed at outer angle ( $C$.
decemmaculatum has antennomere IV slender, elytral apex slightly concave, with outer angle slightly projected, and elytral setae distinctly thick); 6. C. fairmairei (Thomson, 1865) - body robust, and elytral apex wide, with outer angle rounded (C. fairmairei with body slender, and elytral apex narrower, with outer angle not rounded); 7. C. guanabarinum (Martins, 1962) - elytra mostly dark brown, with outer angle unarmed (C. guanabarinum has large light areas on elytra, and outer angle of apices spiniform); 8. C. inflatum Martins and Galileo, 2014 - antennomere III parallel-sided almost from the base, elytral setae about as long as diameter of antennomere III, and prothorax slightly wider than length of antennomere III (C. inflatum has antennomere III gradually widened from base, with elytral setae distinctly longer than diameter of antennomere III, and prothorax distinctly wider than length of antennomere III); 9. C. inornatum (Martins, 1962) - elytra with light macula in basal half, and elytral apex unarmed (C. inornatum without macula, and with long spine at outer elytral apex); 10. C. megarthron (Martins, 1962) - elytra nearly entirely dark, and elytral apex unarmed (C. megarthron nearly entirely light, and elytral apex with long spine at outer angle); 11. C. monnei Martins, 1969 - general color dark brown, almost black, and elytra with a single, oblique light macula directed toward scutellum (C. monnei with general color light reddish-brown, and elytra with two light maculae, the anterior one elliptical and not directed toward scutellum); 12. C. paradoxum Martins, 1971 - elytra nearly entirely dark, and antennomere IV short and thick (C. paradoxum with elytra entirely lighter, and antennomere IV long and slender); 13. C. psydrum Martins, 1969 - elytra nearly entirely dark, with apex wide and unarmed (C. psydrum has elytra mostly light, and elytral apex narrow, and projected at outer angle); 14. C. punga Martins and Galileo, 2007 - antennomere IV thick, and elytral apex unarmed (C. punga has antennomere IV slender, and elytral apex spiniform at outer angle); 15. C. reichardti (Martins, 1962) - elytra mostly dark brown (C. reichardti has basal half of the elytra reddish and distal half black); 16. C. singulare (Gounelle, 1909) - antennomere IV thick, and outer elytral apex unarmed (C. singulare has antennomere IV not thick, and outer elytral apex spiniform); 17. C. sphaeriinum (Bates, 1870) - antennomere IV thick, and outer elytral apex unarmed (C. sphaeriinum has antennomere IV not thick, and outer elytral apex spiniform); 18. C. taperu Martins and Galileo, 2007 - pronotum without dense white pubescent macula, and elytra nearly entirely dark, with outer angle rounded (C. taperu with dense pubescent macula, and elytra with large light areas); 19. C. trichocerum (Martins, 1962) - elytra nearly entirely dark, elytral apex with rounded outer angle (C. trichocerum nearly entirely reddish, and outer elytral apex with long spine); 20. C. trinidadense (Gilmour, 1963) - antennomere III about as wide as IV, and elytral apex rounded at outer angle (C. trinidadense has antennomere III distinctly wider than IV, and elytral apex spiniform); 21. C. unifasciatum (Gounelle, 1909) - antennomere IV thick, and elytral apex unarmed (C. unifasciatum has antennomere IV slender, and elytral apex with spine at outer angle); 22. C. varipenne Martins, 1969 - antennomere IV thick, and outer elytral apex rounded (C. varipenne has antennomere IV slender, and elytral apex spiniform).

Currently, 17 species of Compsibidion are only known from females. Of these, five lack spines on the outer elytral apex as in the holotype male of C. woodleyi sp . n. Typically, this kind of sexual dimorphism (elytral apex armed in female and unarmed in male) does not occur in Compsibidion. Of the five species with unarmed outer elytral apex, only C. achiraensis Santos-Silva, Galileo and Wappes, 2017 and C. amboroensis Santos-Silva, Galileo and Wappes, 2017 could even remotely be confused with the new species (the general appearance of the other three are dramatically different). Compsibidion woodleyi sp. n. differs from both these species by the light elytral macula placed at basal third, smaller, and ending away from the suture (placed at about middle, larger, and ending nearer the suture in $C$. achiraensis and C. amboroensis), and elytral apex almost truncate (uniformly rounded in C. achiraensis and C. amboroensis).

Compsibidion woodleyi sp. n. can be included in the alternative of couplet " 4 " from Santos-Silva et al. (2017):
$4^{\prime}(2) . \quad$ Elytral apex wide, nearly truncate; with light-colored elytral macula placed at basal third; Bolivia . . . . . . . . . . . . . C. woodleyi Wappes, Santos-Silva and Galileo, new species

4(4'). Upper eye lobes with 5 rows of ommatidia; elytra at least partially with moderately abundant, decumbent pubescence; Bolivia
C. amboroensis Santos-Silva, Galileo and Wappes, 2017

- Upper eye lobes with 3 rows of ommatidia; elytra with very sparse, decumbent pubescence; Bolivia
C. achiraensis Santos-Silva, Galileo and Wappes, 2017


## Lamiinae

## Calliini

## Drycothaea dozieri Wappes, Santos-Silva and Galileo, new species

(Fig. 5-9)
Description. Female. Integument mostly dark brown; anteclypeus reddish-brown; distal half of labrum yellowish-brown; most of mouthparts reddish-brown, with palpi almost black except yellowish apex of each segment; antennomere III dark reddish-brown basally, gradually darkening to darker brown apex; antennomere IV reddish-brown basally, brownish on wide central area, dark brown on apex; antennomeres V-X light reddish-brown, gradually slightly darkened on wide central area, dark brown on apex (this latter area gradually smaller and less conspicuous toward X); antennomere XI light reddish-brown; apex of abdominal ventrites I-IV narrowly yellowish-brown. Pubescence pale-yellow (more whitish or golden depending on angle of light source).
Head. Frons coarsely, sparsely punctate; pubescence moderately abundant, not obscuring integument; with long, erect dark seta emerging from nearly all punctures, and close to eyes. Area between antennal tubercles with sculpturing, pubescence, and erect setae as on frons. Area between upper eye lobes with row of punctures, slightly coarser than on frons, on each side of central groove; pubescence as on frons; with a long, erect dark seta emerging from some punctures. Remaining surface of vertex minutely punctate; pubescence sparser than on frons; without erect setae. Area behind eyes minutely punctate (less so behind lower eye lobes); pubescent close to eye, nearly glabrous close to prothorax; with a few long, erect dark setae close to lower eye lobe. Genae very finely, transversely striate; with sparse pubescence close to eye, glabrous toward apex. Antennal tubercles impunctate; pubescence as on frons; with a few long, erect dark setae. Median groove distinct from clypeus to area between antennal tubercles, just indicated from this point to prothorax. Postclypeus coarsely, sparsely punctate in center, slightly rugose in remaining wide central area, smooth laterally; pubescence as on frons in wide central area, gradually sparser toward anteclypeus, glabrous in smooth area; with long, erect, sparse dark setae in pubescent area (one very long on each side). Labrum coplanar with anteclypeus in basal 2/3, inclined at distal third; minutely, abundantly punctate, with interspersed slightly coarser punctures at apex of basal 2/3; with sparse pubescence interspersed with long, erect dark setae directed forward; with fringe of golden setae at apex. Gulamentum smooth, glabrous toward prothorax, finely punctate, sparsely pubescent, with a few long, erect setae toward mouthparts. Distance between upper eye lobes 0.39 times length of scape; in frontal view, distance between lower eye lobes 0.78 times length of scape. Antennae 1.4 times elytral length, reaching elytral apex at base of antennomere XI; scape slightly widened toward apex; moderately coarsely, sparsely punctate; pubescence not obscuring integument, interspersed with long, erect dark setae; antennomeres III-X with long, erect dark setae ventrally (gradually shorter, sparser toward X), and long, erect dark setae at dorsal apex (shorter toward X); Antennal formula (ratio) based on antennomere III: scape $=0.93$; pedicel $=0.22 ; \mathrm{IV}=0.95 ; \mathrm{V}=0.82 ; \mathrm{VI}=0.73 ; \mathrm{VII}=0.67 ; \mathrm{VIII}=0.58$; $\mathrm{IX}=0.53 ; \mathrm{X}=0.42 ; \mathrm{XI}=0.47$.

Thorax. Prothorax 1.5 times wider than long (including lateral tubercles); lateral tubercles moderately large, conical, nearly acute at apex, placed at about middle. Pronotum with large, subcircular, slightly distinct gibbosity on each side of distal half, and elongate, slightly elevated gibbosity at center of basal half; coarsely, sparsely punctate except on smooth central gibbosity; pubescence moderately abundant, not obscuring integument; with long, erect dark setae emerging from some punctures. Sides of prothorax
smooth between base and lateral tubercle, coarsely punctate close to pronotum between lateral tubercle and narrow, smooth distal area; pubescence moderately abundant, not obscuring integument; with long, erect dark setae emerging from some punctures. Prosternum very finely rugose, with interspersed fine punctures; pubescence not obscuring integument; with long, erect dark setae toward prosternal process; prosternal process noticeably narrowed centrally. Ventral side of meso- and metathorax with pubescence not obscuring integument, slightly sparser centrally. Mesoventral process slightly longitudinally elevated centrally; apex emarginate. Scutellum with yellow pubescence, denser toward margins. Elytra. Coarsely, abundantly punctate (slightly finer, sparser toward apex); pubescence abundant, not obscuring integument; with long, erect, dark setae emerging from punctures; apex rounded. Legs. Femora moderately finely, sparsely punctate; pubescence abundant, not obscuring integument; with a few long, erect dark setae. Tibiae slightly, gradually widened toward apex; with abundant, long erect, dark and yellowish setae, especially dorsally on meso- and metatibiae. Metatarsomere I slightly shorter than II-III together.

Abdomen. Pubescence abundant, not obscuring integument; apex of ventrite V truncate.
Dimensions in mm (holotype female). Total length 7.55; prothoracic length 1.45; basal prothoracic width 1.70 ; distal prothoracic width 1.50 ; max. prothoracic width 2.15 ; humeral width 2.70 ; elytral length 5.45.
Type material. Holotype female from BOLIVIA, Santa Cruz: Potrerillo del Guendá (350-400 m; 17 ${ }^{\circ} 40^{\prime} \mathrm{S}$ / 63²7${ }^{\circ}$ W), 7-9.IX.2012, J. Wappes, P. Skelley and T. Bonaso col. (FSCA).
Etymology. Named to honor B. K. "Byrd" Dozier, an avid lifetime collector of beetles (especially the Buprestidae and Cerambycidae), whose most favorite collection locality in Bolivia is Potrerillo del Guenda, the type locality of this species (Note: in February of this year Byrd celebrated his 91st Birthday!)

Remarks. Drycothaea dozieri sp. n. differs from D. angustifrons (Breuning, 1943) by the distance between upper eye lobes wider than width of one upper lobe (at most equal to width of an upper lobe in D. angustifrons), and tubercle of mesoventral process not forming elevated keel (forming elevated keel in D. angustifrons). The new species differs from D. viridescens (Buquet, 1857) by the mesoventral process without an elevated central tubercle (Fig. 6) (tubercle present in D. viridescens (Fig. 10)).

Drycothaea dozieri sp. n. can be included in the alternative of couplet " 22 " from Galileo and Martins (2010):

22'(21). Mesoventral process without distinct elevated tubercle; Bolivia $\qquad$ . . . . . . . . . . . . . . . . . . . . D. dozieri Wappes, Santos-Silva and Galileo, new species

- Mesoventral process with distinct elevated tubercle ...................................... 22

22(22'). Elytral punctures separated, those of row close to suture, at central area, separated by distance equal to twice the diameter of a puncture; punctures of distal area of the elytra sparse; Venezuela
D. estola (Lameere, 1893)

- Elytral punctures of the row close to suture, at central area, separated by distance equal to diameter of a puncture; punctures of distal area of the elytra not distinctly sparse; Brazil (Bahia, Espírito Santo, Rio de Janeiro, São Paulo, Santa Catarina), Paraguay, Argentina (Misiones)
D. viridescens (Buquet, 1857)


## Mallodon downesii Hope, 1843

(Fig. 11)
Mallodon downesii Hope 1843: 366; Tavakilian and Chevillotte 2018: website (cat.).
In 2009, a male specimen of Mallodon downesii was collected in a light trap in Costa Rica: "La Ceiba Manzanillo". The place is a private reserve near "Refugio de vida silvestre Gandoca-Manzanillo" and Puerto Viejo, in the southern Caribbean Zone of Costa Rica, province of Limón. As INBio has a very large collection of insects from Costa Rica, we sent photographs of the specimen to Angel Solís asking if he was aware of other specimens of this species from the country. He made an extensive search of the INBio collection but was not able to locate additional specimens. Since it is impossible to know how the


Figure 11. Mallodon downesii, male, dorsal habitus of the specimen collected in Costa Rica.
specimen was introduced into Costa Rica, it is considered an incidental introduction.
According to Delahaye and Tavakilian (2009), Mallodon downesii occurs in: "Afrique du Sud, Angola, Bénin, Botswana, Burundi, Cameroun, République Centrafricaine, Comores, Côte d’Ivoire, Ethiopie, Gabon, Gambie, Guinée, Guinée-Bissau, Kenya, Liberia, Madagascar, Mali, Mayotte, Mozambique, Namibie, Nigeria, Ouganda, République Populaire du Congo, République Démocratique du Congo, Rwanda, São Tomé, Sénégal, Sierra Leone, Tanzanie, Togo, Zambie, Zimbabwe." We were not able to find a work citing this species as occurring outside of Africa.

Full bibliographic references for Mallodon downesii are available in Tavakilian and Chevillotte (2018).
Material examined. COSTA RICA, Limón: La Ceiba Manzanillo ("cebo luminoso"), 1 male, 17.IV.2009, José Esteban Durán col. (MZSP).

## Acknowledgments

We express our sincere thanks to José Rafael Esteban-Durán (Spain) for sending the specimen of Mallodon downesii for study, and to Angel Solís (INBio) for examining the INBio collection for other specimens. Careful reviews by Robert Androw, Cambria, PA and Don Thomas, Weslaco, TX are also very much appreciated by the authors.

## Literature Cited

Delahaye, N., and G. L. Tavakilian. 2009. Note sur Mallodon downesii Hope, 1843, et mise en synonymie de M. plagiatum Thomson, 1867 (Coleoptera, Cerambycidae). Bulletin de la Société entomologique de France 114(1): 39-45.
Galileo, M. H. M., and U. R. Martins. 1991. Revisão da tribo Calliini (Coleoptera, Cerambycidae, Lamiinae). Giornale Italiano di Entomologia 5: 243-262.
Galileo, M. H. M., and U. R. Martins. 2010. Gênero Drycothaea (Coleoptera, Cerambycidae, Lamiinae): chave para espécies, nova combinação, espécies novas. Papéis Avulsos de Zoologia 50(5): 69-75.
Hope, F. W. 1843. On some new insects from Western Africa. The Annals and Magazine of Natural History 11: 364-369.

Martins, U. R. 1967. Monografia da tribo Ibidionini (Coleoptera, Cerambycinae). Parte I. Arquivos de Zoologia 16(1): 1-320.
Martins, U. R. 1970. Monografia da tribo Ibidionini (Coleoptera, Cerambycinae). Parte IV. Arquivos de Zoologia 16(4): 879-1149.
Martins, U. R., and M. H. M. Galileo. 1990. Notas sobre Calliini (Coleoptera, Cerambycidae, Lamiinae). IV. As espécies sulamericanas do gênero Drycothaea Thomson, 1868. Revista Brasileira de Entomologia 34(3): 607-613.
Martins, U. R., and M. H. M. Galileo. 2007. Tribo Ibidionini, Subtribo Ibidionina, p. 177-330. In: U. R. Martins (org.). Cerambycidae Sul-Americanos (Coleoptera). Taxonomia. Sociedade Brasileira de Entomologia; Curitiba. v. 9, ii + 349 p.
Monné, M. A. 2018a. Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part I. Subfamily Cerambycinae. Available at http://cerambyxcat.com/. (Last accessed March 2018.)
Monné, M. A. 2018b. Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part II. Subfamily Lamiinae. Available at http://cerambyxcat.com/. (Last accessed March 2018.)
Santos-Silva, A., M. H. M. Galileo, and J. E. Wappes. 2017. Descriptions, notes and reassignments in Neoibidionini (Coleoptera: Cerambycidae: Cerambycinae) with a new genus, three new species and keys to species of Brechmoidion Martins, 1969, Compsibidion Thomson, 1864 and Rhysium Pascoe, 1866. Insecta Mundi 560: 1-21.
Tavakilian, G. L., and H. Chevillotte. 2018. Titan: base de données internationales sur les Cerambycidae ou Longicornes. Version 3.0. Available at: http://titan.gbif.fr/. (Last accessed January 2018.)

Received March 24, 2018; accepted March 31, 2018.
Review editor Jiri Zidek.

