## Supporting Information Available

## Tables

Table S1: FP secondary-structure elements in membrane contact during initial membrane binding event that led to stable binding and in final trajectory frame (initial/final).

membrane system	run no.	NTH bound	AH2 bound	CTH bound
endosomal	1	✓ / ✓	✓/✓	$\times/\checkmark$
endosomal	2	✓ / ✓	$\times / \times$	\$   \$
endosomal	3	$ imes/\checkmark$	$\checkmark/ imes$	$\checkmark/ imes$
endosomal	4	5/5	$\times / \times$	$\checkmark/ imes$
outer plasma	1	$\times / \times$	$\times/\checkmark$	✓ / ✓
outer plasma	2	$\times / \times$	$\times / \times$	$\times / \times$
outer plasma	3	×/×	5/5	✓/×
outer plasma	4	$\times / \times$	5/5	<i>√</i> / <i>√</i>

Table S2: Outer plasma membrane-like lipid membrane composition as created by Lorent et al.  $^{35}$ 

Lipid	Acyl chains	Full name	Abundance %
CHOL		Cholesterol	40
PSM	18:1/16:0	N-palmitoyl-D-erythro-sphingosylphosphorylcholine	12
NSM	18:1/24:1	N-nervonoyl-D-oleoyl-sphingosylphosphorylcholine	9.3
LSM	18:1/24:0	N-lignoceroyl-D-oleoyl-sphingosylphosphorylcholine	8
PLPC	16:0/18:2	1-palmitoyl-2-linoleoyl-sn-glycero-3-phosphocholine	14.7
SOPC	18:0/18:1	1-stearoyl-2-oleoylphosphatidylcholine	6.7
PAPC	16:0/20:4	1-palmitoyl-2-arachidonoyl-glycero-3-phosphocholine	5.3
PLA20(PE)	18:0/20:4	1-O-stearoyl-2-O-arachidonoyl-glycero-3-phosphoethanolamine	2.7
SAPS	18:0/20:4	1-stear oyl-2-arachidon oyl-glycer o-3-phosphoser ine	1.3

Table S3:	Late	endosomal	membrane	composition
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Lipid	Acyl chains	Full name	Abundance %
CHOL		Cholesterol	27
PLPC	16:0/18:2	1-palmitoyl-2-linoleoyl-sn-glycero-3-phosphocholine	22
POPE	16:0/18:1	1-palmitoyl-2-oleoyl-glycero-3-phosphoethanolamine	16
BMP	18:1/18:1	Bis(monoacylglycero)phosphate	11.6
POPC	16:0/18:1	1-palmitoyl-2-oleoyl-glycero-3-phosphocholine	11
SSM	18:1/18:0	N-stearoyl-D-erythro-sphingosylphosphorylcholine	7.3
SAPI	18:0/20:4	1-stearoyl-2-arachidonoyl-sn-glycero-3-phosphoinositol	3.65
SAPS	18:0/20:4	1-stearoyl-2-arachidonoyl-sn-glycero-3-phospho-L-serine	1.45

Table S4: Restraints used during energy minimization and equilibration of the FP in aqueous solution. Values in units of  $\rm kJ\,mol^{-1}\,nm^{-2}$ 

	Backbone	Sidechains	Dihedrals
EM	400	40	4
$\mathbf{EQ}$	400	400	4

Table S5: Restraints used during energy minimization and equilibration of the endosomal and outer plasma membrane systems in  $\rm kJ\,mol^{-1}\,nm^{-2}$ 

	Time [ns]	Timestep [fs]	Backbone	Sidechains	Lipids	Dihedrals
EM			4000	2000	1000	1000
$1^{\rm st}$ EQ	0.125	1	4000	2000	1000	1000
$2^{nd} EQ$	0.125	1	2000	1000	400	400
$3^{\rm rd}$ EQ	0.125	1	1000	500	400	200
$4^{\text{th}}$ EQ	0.5	2	500	200	200	200
$5^{\text{th}} \text{ EQ}$	0.5	2	200	50	40	100
$6^{\rm th} {\rm EQ}$	0.5	2	50	0	0	0

## Figures

AAL40400.1/914-953 AAP41037.1/798-837 AAS00003.1/798-837 AAT76147.1/798-837 AAU04646.1/798-837 AAV49730.1/798-837 AAX76521.1/901-940 AAY68297.1/900-939 AAZ41329.1/785-824 ABC70719.1/901-940 ABD75497.1/905-944 ABI96958.1/798-837 ABN10839.1/887-928 ADE34722.1/785-824 ADM33558.1/808-849 AEN19366.1/912-951 AFG25750.1/906-945 AGN52936.1/888-929 AGT17716.1/870-909 AGW27881.1/905-944 AHY61337.1/855-896 AIA62277.1/779-818 AIX09799.1/909-948 AJA91196.1/907-946 AJC98127.1/913-952 AKZ19076.1/784-823 AMN88686.1/905-944 AOL02453.1/909-948 AQN78656.1/909-948 AQZ41285.1/888-929 ARB07438.1/902-941 ARC95203.1/900-939 ARQ84735.1/888-929 ASL68941.1/879-920 ASU45696.1/888-929 ATN37888.1/870-909 ATN37896.1/870-909 ATY74381.1/888-929 ATY74392.1/888-929 ATY74403.1/888-929 ATY74414.1/888-929 AUF40275.1/900-939 AVK87318.1/888-929 AVP25406.1/803-845 AVV61900.1/888-929 AWM99582.1/888-929 AWU59321.1/888-929 AWW13511.1/910-949

. 1	0	20	30	40
SAIEDLLFS	VKLSDVGF			
<mark>s</mark> fi <mark>ed</mark> llfn <mark>k</mark>				
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>				
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>	VTLADAGFN	MKQ <mark>YGEC</mark> LO	3 D I N A R	DLICAQKF
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>	VTLADAGFN	MKQ <mark>YGEC</mark> L <mark>O</mark>	3 D I N A R	DLICAQKF
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>	V T L A D A G F I	MKQ <mark>YGEC</mark> LG	D I N A R	DLICAQKF
<mark>S</mark> LL <mark>ED</mark> LLFN <mark>K</mark>	VKL S <mark>DVG</mark> F	V E A Y N N C T G	🗟 <mark>G S</mark> E I <mark>R</mark>	DLLCVQSF
SAIEDLLFDK	(V <u>k</u> ls <mark>dvg</mark> fv	V <mark>Q</mark> A <mark>Y</mark> N N C T G	<mark>3</mark> <mark>G</mark> A E I R	
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>	VTLADAGFN	MKQ <mark>Y</mark> GDCLO	3 <u>DV</u> SAR	DLICAQKF
<mark>S</mark> LL <mark>ED</mark> LLF <mark>N</mark> K	(VKLS <mark>DVG</mark> F)	VEAYNNCTO	6 G <mark>S</mark> E I <mark>R</mark>	DLLCVQSF
<mark>S</mark> FF <mark>ED</mark> LLF <mark>D</mark> K				
<mark>S</mark> FI <mark>ED</mark> LLF <mark>N</mark> K				
SAIEDLLF <mark>D</mark> K				
<mark>S</mark> FI <mark>ED</mark> LLFN <mark>K</mark>				
SALSDLLYNK				
S A I E D L L F D K				
S A I EDVL F DK				
SAIEDLLFDK				
SAIEDLLFDK				
SFFEDLLFDK				
SAIEDLLFNK				
SFIEDLL <mark>Y</mark> NK SAIEDLLFDK				
SAIEDILFD				
SAIEDLLFDK				
SFIEDLLYNK				
SFFEDLLFDK				
SAIEDLLFDK				
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SAIEDLLFDK				
SLL EDLLF NK				
SAIEDLLFDK				
SAIEDLLFDK	VTIADPGYN	M <mark>QGY</mark> D <mark>DC</mark> M -	QQ <mark>GPA</mark> SA <mark>R</mark>	DLICAQYV
<mark>ST</mark> I <mark>ED</mark> LLFN <mark>K</mark>	(V <mark>T</mark> IA <mark>DPGY</mark> N	M <mark>QGY</mark> D <mark>EC</mark> M -	QQ <mark>GPP</mark> S <mark>A</mark> R	DLICAQFV
<mark>S</mark> AI <mark>ED</mark> LLF <mark>D</mark> K	(V <mark>T</mark> IA <mark>DPGY</mark> M	M <mark>QGY</mark> D <mark>DC</mark> M -	- QQ <mark>GP</mark> ASA <mark>R</mark>	DLICAQYV
<mark>S</mark> AI <mark>ED</mark> LLF <mark>D</mark> K	(VKL S <mark>DVG</mark> F )	VEAYNNCTO	<mark>3</mark> <mark>G</mark> QE <mark>V</mark> L	DLLCVQSF
<mark>S</mark> AI <mark>ED</mark> LLF <mark>D</mark> K				
<mark>S</mark> AI <mark>ED</mark> LLF <mark>D</mark> K				
<mark>S</mark> AI <mark>ED</mark> LLF <mark>D</mark> K				
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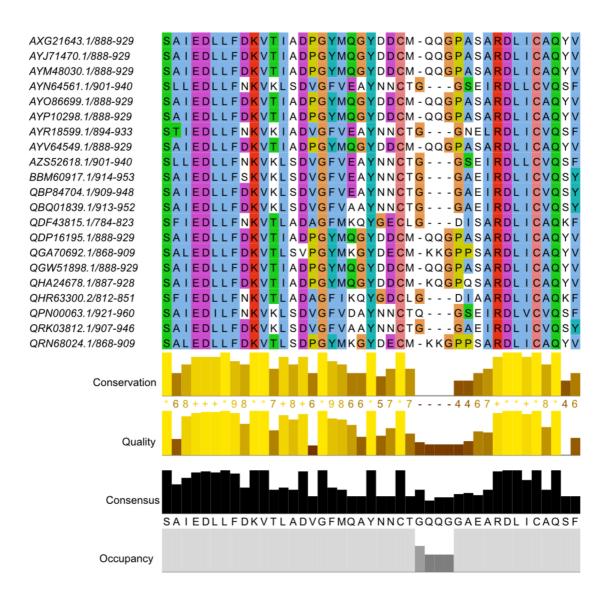


Figure S1: Alignment of the FP region of betacoronaviruses. Complete S protein sequences of the genus betacoronavirus were selected from the NCBI virus database<sup>40</sup> and aligned using Clustal Omega.<sup>22</sup> Only the FP region is shown. The fully conserved cysteines are at positions 25 and 39, respectively. Multiple entries from the same species were deleted. Coloring according to ClustalX.

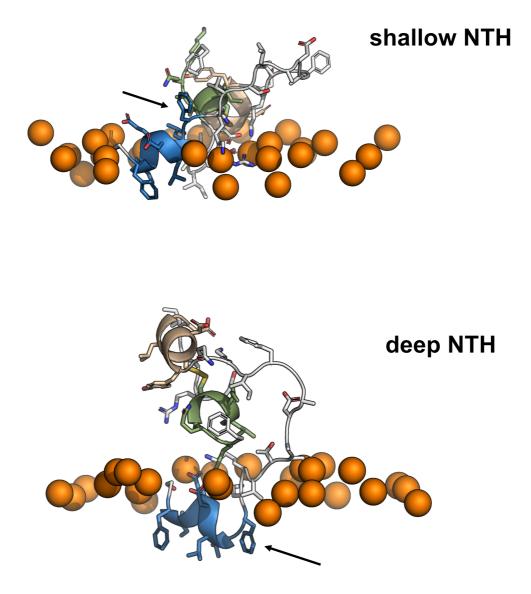


Figure S2: **Deep and shallow NTH insertion.** Arrows indicate the F823 position not inserted (top) and inserted (bottom) into the membrane interface. Colors as in Figure 1 (NTH: blue, AH2: beige, CTH: green). The upper membrane boundary is indicated by phosphate headgroups of nearby lipids (orange spheres).

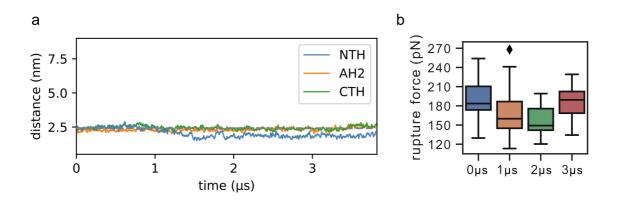


Figure S3: Simulation starting with all three helices bound and adjusted outerleaflet density. (a) Distances of the centers of mass of the three helices and the center of mass of the membrane in which the lipid density of the outer leaflet was adjusted to accommodate the bound FP. The average phosphate position of the outer leaflet is indicated by a gray dotted line. (b) Box-plots showing the peak rupture forces of 20 replica pulling simulations each for pulling simulations started with structures of the simulation taken at 0, 1, 2, and 3  $\mu$ s (box: interquartile range; median: horizontal line; diamond: outlier).

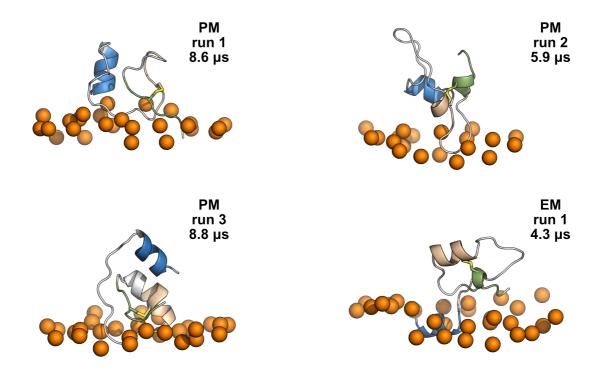


Figure S4: Snapshots of MD simulations in which the AH2 and CTH partially unfolded to form flexible membrane bound structures. (Left) FP interactions with the mimetic of the outer plasma membrane (PM). (Right) FP interactions with the mimetic of the endosomal membrane (EM). Time points are indicated. Colors as in Figure 1 (NTH: blue, AH2: beige, CTH: green). The upper membrane boundary is indicated by phosphate headgroups of nearby lipids (orange spheres).

## Videos

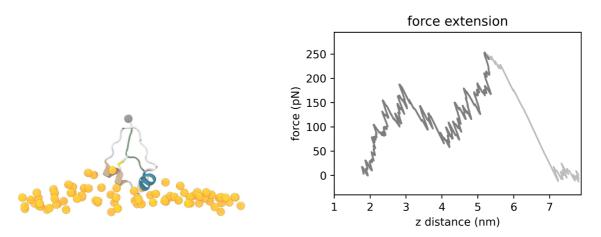


Figure S5: Movie of FP pulling. (Left) Trajectory of the pulling simulation that resulted in the highest rupture force of about 250 pN. The FP is shown in the same colors as used in Figure 1. The gray sphere indicates the C-terminus, to which the pulling force is applied. Lipid headgroup phosphates are shown as yellow spheres. (Right) Force extension curve corresponding to the trajectory. X-axis shows the distance of the C-terminal carbon (gray sphere) to the center of mass of the membrane. Y-axis shows the force acting on the Cterminal carbon.