

Marker	1a	2a	2b	3a	3b	3c	3d	3e	3f	3g	3h	4a	5a	5b	5c	5d	5e	6a	6b	7a	8a	8b	9a	9b	10a
	E	Y2	X2	F2	X2	F2	Y4	X2	D2	D2	D2	H2	C2	B2	B2	C	(GGLTR4a)	B2	C	C	B2	B2	(GGERV- 18LTR)	(GGERV- 18LTR)	D2
CR1-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
<i>Taeniopygia guttata</i>																									
<i>Anas crecca</i>																									
<i>Cairinia moschata</i>																									
<i>Alectura lathami</i>	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
<i>Crax fasciolata</i>	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	
<i>Crax alector</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
<i>Numida meleagris</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
<i>Callipepla squamata</i>																									
<i>Colinus virginianus</i>																									
<i>Rollulus rouloul</i>																									
<i>Gallicus gallus</i>																									
<i>Gallus lafayettii</i>																									
<i>Coturnix japonica</i>																									
<i>Africopavo congensis</i>																									
<i>Pavo cristatus</i>																									
<i>Pavo muticus</i>																									
<i>Tragopan caboti</i>																									
<i>Perdix perdix</i>																									
<i>Chrysolophus amherstiae</i>																									
<i>Chrysolophus pictus</i>																									
<i>Tetrao tetrix</i>																									
<i>Tympanuchus cupido</i>																									
<i>Meleagris gallopavo</i>																									

Table S2. Presence (+) and absence (-) patterns. 1a-10a correspond to the analyzed markers. Row 2 denotes the CR1 subtypes or LTR elements (in brackets) present in the (+) state. For empty cells no data was obtained.

Table S3. Oligonucleotides used for PCR amplifications

primer	(5' - 3')
1a-F	TGCCCATCTTGATGATAATGAGAG
1a-R	TCTCGCCTGCAATGTAAGTGAC
2a-F (5b)	CATTCCAGTGTCTGATGAAGCAG
2a-R (5b)	GTCAGAAGTAGGCCACCATAGACAC
2b-F (8a)	GGAGGGACCTGGCAGCAC
2b-R (8a)	CAACACCGTGGACAGCATTG
3a-F	AAAGTTCGCTACGTGAAGCTG
3a-R	CTTCCTTGGTTTCTCATCCAC
3b-F	AAGGTGACCCAGGTATTGAAG
3b-R	TTCTCACCTTTCTCCTTCAG
3c-F	CCACCACAGGGAGCTTCAG
3c-R	GGTTTCAACAAATCTCCTGGAC
3d-F	CCTCACAGCGAGTAACCATGAC
3d-R	GAACCAAGAAAATACATTGCTGC
3e-F	GGCTTAATGATTTCATTAGAAG
3e-R	ATTCCAAGATAGATTGAACCTCAG
3f-F	CTCTAACAAATGAGCGAGAACCG
3f-R	AGGTGATCCAGTCCAGCATATC
3g-F	GTGACAGGAGTCCAGGTGATCTG
3g-R	GGAGCCACCACTGTGATGCAG
3h-F	CACGTGGAGATGAAGCAGAAC
3h-R	CTCCAGGTAGTTGATGCCTTCATAG
4a-F	CATCAGACACCTCCTGGGTCA
4a-R	GGCTGCTCAGCTTGCTGAC
5a-F (10a)	ATACCGCCTGTTCACACTGCT
5a-R	CGTACATTGCAAAGGATATCACAG

(10a)	
5a-Fa (10a)	TCATAGATGAGCTGAAACAG
5a-Ra (10a)	GAAATCATCGATTTGTGG
5c-F	GAGTTGGAGTGGTGATCGAAGC
5c-R	TTTTCTGTACCCCACGCATTAC
5c-Fa	GTGATCGAAGCTACGTAAG
5c-Ra	TTACCATAACACATATGCCTAG
5d-F	TGACACTGTTTATGTACACATCAAC
5d-R	TCCAGAATGAGAGAAGTAGCAGC
5e-F	CCAGTAATCCAGGATCTCTGTCAC
5e-R	GGACTATCCGTGACAAGTTGC
6a-F	CGTGTATTTCTGGTGCACAGC
6a-R	ATGGTTTGCTGACAGCTATGC
6a-Fa	GAGTAAAATGACATGATTGC
6a-Ra	AGAACCCATACGGATCTC
6b-F	CTACAGTAACACTCAGCATGTCTGC
6b-R	GTCCCATATCAGCAATCTGTACTC
7a-F	GCAGGCACAGTTGGTAAAG
7a-R	TGATTCATTCGTCTCAGCTC
7a-Fa	CTGACTCTGAATTGGTCTCAG
7a-Ra	TATCCAAGACCATGACCG
7a-Fb	TCCAGAATTGCAGTAGCTG
7a-Rb	AGCCTCTGAGCCAATTG
8b-F	CCGTGAGCGCAACAAGACG
8b-R	CGGTGATGCCAGAGAACTTCTC
9a-F	TGCATAATAAGTCATCTGAGCTTC
9a-R	CGTCAAGCAGAGATTACCAGAC
9b-F	TCTGGCTTCTTTCTAATAGCAATG
9b-R	TGCTGATTTTGAAAACTTGATAGA

Primer names correspond to markers in Table S2. (F) denotes forward primers, (R) denotes reverse primers.