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Ozone stress as a driving force of sesquiterpene emissions: a suggested parameterization

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Abstract

Sesquiterpenes ($C_{15}H_{24}$) are semi-volatile organic compounds emitted by vegetation and are of interest in atmospheric research because they influence the oxidative capacity of the atmosphere and contribute to the formation of secondary organic aerosols. However, little is known about their emission pattern and no established parameterization is available for global emission models. The aim of this study is to investigate a Central European spruce forest and its emission response to meteorological and environmental parameters, looking for a parameterization that incorporates heat and oxidative stress as the main driving forces of the induced emissions. Therefore, a healthy ca. 80 yr old Norway spruce (*Picea abies*) tree was selected and a dynamical vegetation enclosure technique was applied from April to November 2011. The emissions clearly responded to temperature changes with small variations in the β -factor along the year ($\beta_{\rm spring} = 0.09 \pm 0.01$, $\beta_{\rm summer} = 0.12 \pm 0.02$, $\beta_{\rm autumn} = 0.11 \pm 0.02$). However, daily calculated values revealed a vast amount of variability in temperature dependencies ($(0.02 \pm 0.002) < \beta < (0.27 \pm 0.04)$) with no distinct seasonality.

By separating the complete dataset in 10 different ozone regimes, we found that in moderately or less polluted atmospheric conditions the main driving force of sesquiterpene emissions is the temperature, but when ambient ozone mixing ratios exceed a critical threshold of (36.6 ± 3.9) ppb_v, the emissions become primarily correlated with ozone. Considering the complete dataset, cross correlation analysis resulted in highest correlation with ambient ozone mixing ratios ($CC_{O_3} = 0.63\pm0.01$; $CC_T = 0.47\pm0.02$ at t=0 h for temperature) with a time shift 2–4 h prior to the emissions. An only temperature dependent algorithm was found to substantially underestimate the induced emissions (20% of the measured; $R^2 = 0.31$). However, the addition of an ozone dependent term improved substantially the fitting between measured and modeled emissions (81% of the measured; $R^2 = 0.63$), providing confidence about the reliability of the suggested parameterization for the spruce forest site investigated.

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1 Introduction

Biogenic volatile organic compounds (BVOCs), including monoterpenes (MT), sesquiterpenes (SQT), isoprene and oxygenated VOCs, are reactive plant compounds emitted into the atmosphere (Kesselmeier et al., 1999). They have been hypothesized to function as endogenous antioxidants within plants, protecting them from oxidative damage during the stress-induced accumulation of reactive oxygen species (Vickers et al., 2009). Especially SQT, react readily with atmospheric ozone, with a high potential to form secondary organic aerosol (Hoffmann et al., 1997; Jaoui et al., 2003; Bonn et al., 2008). Their vital role on atmospheric chemistry is stressed out by their ability to influence the oxidative capacity of the atmosphere (Fuentes et al., 2000) and their contribution to secondary organic aerosol production (Hoffmann et al., 1997; Andreae and Crutzen, 1997; Bonn et al., 2003; Kulmala et al., 2004). In order to understand the role of BVOCs in atmospheric chemistry, it is important to quantify their emissions and atmospheric abundance, along with a better understanding of their atmospheric oxidation.

SQT belong to a broad spectrum of VOCs emitted by plant leaves that typically act in multiple roles for plant protection (Niinemets et al., 2004). However, the mechanism by which volatile isoprenoids (isoprene, MT, SQT) protect plants against oxidative stress and probably against a wide range of environmental stresses is still under investigation. In a recent study, Jardine et al. (2012) highlighted the potential function of isoprene production, in protecting plants against reactive oxygen species. Global and regional atmospheric chemistry models typically incorporate empirical algorithms that have been developed to model VOC emission rates and their dependence on photosynthetic active radiation and leaf temperature (Guenther et al., 1995, 2000, 2006), with an exponential relationship between emission rates and leaf temperature. Nevertheless, strong variations of standard emission factors of many plant species with discrepancies of up to an order of magnitude have been reported (Kesselmeier and Staudt, 1999; Niinemets et al., 2011). Fillela et al. (2006) concluded that algorithms which use only

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incident irradiance and leaf temperature as drivers to predict VOC emission rates may be inadequate. As the review of Holopainen and Gershenzon (2010) has shown, the interaction of multiples stresses, both biotic and abiotic, provides a great potential to alter VOC emission. It is most likely that other environmental factors such as drought, ozone and CO₂ concentrations can amplify or attenuate such alterations in the emission responses to temperature and light (Staudt et al., 2011).

The phytotoxicity of ozone has been demonstrated for forest tree species since more than 50 yr (Karnosky et al., 2007 and references cited therein). Current evidence suggests that several global change drivers such as ozone concentration can also affect both constitutive and herbivore-induced BVOCs production (Loreto et al., 2007; Vuorinen et al., 2005; Blande et al., 2007). Ozone causes biochemical and physiological changes leading to the inhibition of photosynthesis and a consequent decrease in plant growth (Guderian et al., 1985), often associated with visible injuries (Loreto et al., 2001; Vollenweider and Gunthardt-Georg, 2005). Plants act as a sink for ozone, through stomatal and nonstomatal processes (Fares et al., 2010) while ozone may apparently induce biosynthesis and emission of volatile isoprenoids, mainly SQT, even in plants that do not naturally emit these compounds (Heiden et al., 1999). In the presence of NOx, however, these BVOCs initiate reactions that lead to ozone formation (Fehsenfeld et al., 1992; Kurpius and Goldstein, 2003). Hence, the dual action of VOCs is depending on the presence of atmospheric pollutants (Loreto and Schnitzler, 2010). The chemical reactivity of atmospheric terpenes led to the hypothesis that volatile terpenes play a similar dual action inside the leaves before they are released into the atmosphere (Hewitt et al., 1990; Loreto et al., 2001). Vegetation is exposed to increasing levels of tropospheric ozone concentrations, particularly in Northern mid-latitudes (Brasseur et al., 1998; Parrish et al., 2009). Usually forests considered to be low level ozone regions due to the excessive sink terms. However, because of the urban areas at the vicinity of the site of interest, highly elevated ozone concentrations are observed during the season. This pollutant penetrates in leaves through stomata and quickly reacts inside leaves, thus making plants valuable ozone sinks, but at the same time

triggers oxidation processes which lead to leaf injuries (Fares et al., 2010). Therefore, atmospheric interactions of ozone and other oxidants with biogenic VOCs appears to be quite complex, especially in forest ecosystems.

In their review, Peñuelas and Staudt (2010) reported a significant amount of results that indicate elevated SQT emissions under ozone stress. However, SQT emissions were treated to date as only temperature driven, with slightly higher temperature dependency from MT. To date, aerosol model studies include only MTs as the biogenic precursor for organic aerosol (Carslaw et al., 2010) as no global SQT emission inventory is available (Duhl et al., 2008).

According to Jardine at al. (2011), only a few studies have attempted to quantify ambient SQT concentrations (Hakola et al., 2006; Bonn et al., 2007; Bouvier-Brown et al., 2009; Boy et al., 2008; Kim et al., 2009, 2010) including the attempt of Kim et al. (2009) to quantify ecosystem-scale SQT emission rates using the vertical gradient technique. Nevertheless, the necessity for further experimental data (see Hakola et al., 2006; Tarvainen et al., 2005, 2007 with remarks) and the need for a parameterization which includes further environmental factors (Loreto et al., 2004; Staudt et al., 2011), is apparent. According to Loreto et al. (2004) and to Holopainen and Gerhenson (2010), only the study of plant volatile emission under realistic, multiple stress regimes will be able to determine the natural occurrence of such VOC reaction products and their importance in plant function. This study provides on-line seasonal measurements of SQT emissions from a spruce tree into the atmosphere, under a variety of stress regimes and attempts to combine temperature and ozone stress in a single parameterization that can be used for future model calculations.

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2 Material and methods

2.1 Site description

Biogenic emissions were investigated from a (13 ± 1) m tall healthy Norway spruce (Picea abies) tree, located at Taunus Observatory $(50^{\circ}\ 13'\ N, 8^{\circ}\ 26'\ E$ and 825 m a.s.l.), at the top of Mt. Kleiner Feldberg; a hill on the Taunus ridge in South-Western Germany (Crowley et al., 2010). The area around the observatory is covered by a coniferous forest, in which Norway spruce is the dominant tree type with smaller contributions of pine. The area up to $100\ km$ to the North of Kleiner Feldberg is only lightly populated and is devoid of major industry. However, Taunus lies on the Northern rim of the relatively heavily industrialised Rhine-Main region. The remote character of the site is influenced mainly by south-easterly winds, where the city of Frankfurt and the Rhine-Main-area including airport and major traffic pathways are located. The wind pattern can result in very high ozone concentrations, providing a variety of atmospheric conditions that the forest ecosystem is experiencing.

5 2.2 Experimental setup

Emission data were obtained using a branch enclosure technique, applied for a healthy branch 5 m a.g.l. The instrumentation was kept inside a monitoring van, situated below the investigated tree. VOCs were measured by a PTR-MS (Proton Transfer Reaction-Mass Spectrometer), while samples have also been analysed with GC-MS (Gas Chromatography-Mass Spectrometry) techniques for identification of individual VOCs. Temperature, relative humidity, ozone (O₃) and photosynthetic active radiation (PAR, for September onward only) were continuously measured inside the plant cuvette. Therefore, a temperature-humidity sensor (Hygrosens Instruments), an ozone analyser (model APOA-350E, Horiba) and two (inside and outside the cuvette) PAR sensors (LI-190SZ, LI-190SL/Li-Cor Inc., Lincoln, NB, USA) have been applied. Meteorological and environmental parameters (radiation, atmospheric pressure, wind velocity

diation, H₂O, NO, NO₂) were measured by HLUG (Hessian Agency for Environment and Geology) and DWD (German Weather Service), 50 m away from the measuring site.

The dynamic branch enclosure was a cylindrical shaped glass cuvette (volume = 15 l), with a lid that can be opened and closed by an electrical compressor. During closure, air was drawn from the chambers to the analysers along the sampling lines

and direction, temperature, O₃, relative humidity, soil moisture, precipitation, global ra-

closure, air was drawn from the chambers to the analysers along the sampling lines and the under-pressure was avoided with replacement by ambient air at equal flow rate (Ruuskanen et al., 2005). The contact between plant surface and the chamber was avoided when possible, while a fan was installed to ensure homogenous mixing of the air in the enclosure. During June (DOY = 167), the first selected branch showed visible damage in the bark after a storm event and strong winds prevailing at the site of interest. For avoiding different stress responses, another branch close to the selected one was used for the continuation of the experiments. During October (DOY = 283) a type of fungi started to develop on the enclosed branch and a third one was selected. All the data up to 3 days after the newly selected branches were excluded from the data analysis. The quantified PAR losses due to the cuvette material range from 25 to 45%, depending on the solar zenith angle.

2.3 PTR-MS measurements

Volume mixing ratios of total MT and total SQT were quantified using a commercial high-sensitivity proton transfer reaction-mass spectrometer (PTR-MS, IONICON, Austria). General information for the analytical technique can be found in Blake et al. (2009) and references therein. The PTR-MS was operated in standard conditions with a drift tube voltage of 600 V and drift tube pressure of 2.3 mbar (E/N = 117Td; E is the electric field strength in V cm⁻¹, N is the buffer gas number density in units of cm⁻³, 1Td = 1Townsend 10⁻¹⁷ cm² V⁻¹). Optimization of PTR-MS conditions resulted in extremely high and sustained primary ion signal ($H_3O^+ = 4 - 7 \cdot 10^7$ cps). The ion source conditions were such that the contribution of O_2^{*+} impurity ions (the major impurity

ions) relative to the sum of proton hydrates at standard drift tube voltage (600 V) was limited to 2%. Dhooghe et al. (2008) showed that O_2^{*+} ions react equally fast with sesquiterpenes as do H_3O^+ ions, so the contribution of product ions originating from O_2^{*+} /sesquiterpene reactions to the sum of all sesquiterpene product ions observed in the mass spectra will therefore not exceed 2%. The dwell time for MT (m/z = 137) was adjusted to 2s, while the dwell time for SQT (m/z = 205) was adjusted to 20s for further increase in the detection efficiency.

The PTR-MS instrument was frequently calibrated with a gas standard (L4763, Ionimed analytic GmbH, Austria). For SQT, calibrations were performed with a permeation oven technique and a β -caryophyllene standard (W225207, Sigma-Aldrich, Inc.). The experimental determined reaction rate constant for MT was $1.8-2.2 \times$ $10^{-9}\,\text{molecule}\,\text{cm}^3\,\text{s}^{-1}$ (2.5 × $10^{-9}\,\text{molecule}\,\text{cm}^3\,\text{s}^{-1};$ Zhao and Zhang, 2004) and for SQT 2.5 – 2.9 × $10^{-9}\,\text{molecule}\,\text{cm}^3\,\text{s}^{-1}$ (3.0 × $10^{-9}\,\text{molecule}\,\text{cm}^3\,\text{s}^{-1};$ Dhooghe et al., 2008; Kim et al., 2010). Reaction of H₃O⁺ ions with many VOCs mainly results in the protonated VOC, but the H₃O⁺/sesquiterpene interaction in a PTR-MS reactor at typical drift-tube conditions is known to proceed through both dissociative and non-dissociative proton transfer, resulting in multiple fragment ion species. This fragmentation results in a decrease of the detection efficiency with respect to other VOCs, which are not or hardly subjected to fragmentation following protonation (Dhoogle et al., 2008). The PTR-MS technique has already been applied for the detection of sesquiterpenes on several occasions and expected product ions and their relative abundances have been reported for some sesquiterpenes at standard PTR-MS conditions (Ezra et al., 2004; Lee et al., 2006; Demarcke et al., 2009; Kim et al., 2009, 2010). The relative abundance of parent SQT ion signal quantified as 32±2% and comes in line with the previous values reported. According to Kim et al. (2009) when only m/z 205+ is considered, the uncertainty increases to 30 %.

Another major issue when measuring SQT are transfer line (tube) losses. For minimizing this effect the tube that was transferring the air to the instruments was manufactured from glass (inner diameter = $3 \, \text{mm}$, total length = $4.1 \, \text{m}$) and heated up to

 $70\,^{\circ}\text{C}$. Glass is an inert material that reduces memory effects due to BVOC adsorption/desorption on surfaces and was found to be the favorite of all materials tested for SQT measurements (Helmig et al., 2003). Tubing losses were quantified prior to the measurements ($(9\pm1)\,\%$ for SQT, $(7\pm1)\,\%$ for MT) in the laboratory, using a β -caryophyllene (W225207, Sigma-Aldrich, Inc.) and a pinene (L4763, Ionimed analytic GmbH) standards. The flow rate was continuously maintained at 1.8 Imin⁻¹, resulting a very short total residence time in the line downstream of the enclosure (2.2 s).

2.4 Emission calculation

As reported before, the dynamic enclosure was automatically operated with an electrical compressor. The chamber remained open for the most of the time and was closing three times per hour. Closing time adjusted to be 3 min in a 20 min cycle. The mass to charge ratios (m/z) that were continuously monitored during a PTR-MS cycle were: 21 (H_3O^+) , 30 (NO^+) , 32 $(O_2^{\bullet+})$, 37 $(H_2O-H_3O^+)$ with a dwell time of 50 ms each. For the purposes of this study m/z ratios of 137 $(MT-H^+)$, 81 $(major\ MT\ fragment)$, 205 $(SQT-H^+)$ and 149 $(major\ SQT\ fragment)$ were monitored with the aforementioned dwell times of 2 s for the parent ion mass of MT (m/z=137) and 20 s for the parent ion mass of SQT (m/z=205). The time resolution of one PTR-MS measurement was 36 s, resulting in five measurements during a single closed cuvette period. This facilitates a linear fit to the mixing ratios measured, in order to obtain the mean emission during the closure period.

Enclosure approaches cannot be used to estimate emission rates without the exclusion or corrections of ozone and disturbance effects during the studies (Jardine et al., 2011). In order to correct for ozone destruction during the sampling process, selected branch emissions were studied by GC-MS technique at the Finnish Meteorological Institute in Helsinki and compared with the chemical analysis performed earlier at Mainz University (Münz, 2010) for the site of interest. The VOC samples for GC-MS analysis performed at the Finnish Meteorological Institute in Helsinki, were collected from branch enclosures (101 Tedlar bags) on adsorbent tubes filled with Tenax-TA and

Carbopack-B. Emissions from three different branches (2 during spring and one during autumn) were studied. The samples were analyzed in the laboratory, using a thermal desorption instrument (Perkin-Elmer TurboMatrix 650, Waltham, USA) attached to a gas-chromatograph (Perkin-Elmer Clarus 600, Waltham, USA) with DB-5MS (60 m, 0.25 mm, 1 μ m) column and a mass selective detector (Perkin-Elmer Clarus 600T, Waltham, USA). The system was calibrated using liquid standards injected on Tenax TA-Carbopack B adsorbent tubes and analyzed together with the samples. More detailed description on GC-MS analysis can be found from Hellén et al. (2012) and from Munz (2010). Both measurement methods revealed the presence of β -caroyphyllene, α -farnesene, longicyclene and α -humulene while the method applied at the Finnish Meteorological Institute additionally found aromadendrene and longipinene. These values might vary at maximum by ± 10 % along the year, but due limited amount of samples analyzed, a clear conclusion about the relative abundances along the season cannot be driven.

Ozonolysis reaction rate constants were calculated based on these results. On average MT consisted of 28% β -pinene, 25.2% α -pinene, 17.5% limonene, 14.2% β -phellandrene, 9.3% camphene and 5.8% myrcene, while SQT were treated as 35% β -caryophyllene, 29% α -farnesene, 11% aromadendrene, 18% longicyclene, 3% α -humulene and traces of β -farnesene and longipinene. Note that the fraction of β -caryophyllene included longifolene as well, which could not be separated. Based on the work of Münz (2010) this is mainly β -caryophyllene and therefore was treated as such for further analysis. The above approach resulted in the two following mean reaction rate coefficients with respect to ozone: $k_{\text{SQT}+\text{O}_3} = (6.6\pm2.1)\times10^{-15}\,\text{cm}^3\,\text{molecule}^{-1}\,\text{s}^{-1}$; $k_{\text{MT}+\text{O}_3} = (9.4\pm0.9)\times10^{-17}\,\text{cm}^3\,\text{molecule}^{-1}\,\text{s}^{-1}$. Since several of the SQT-ozone reaction rates are not well known, the uncertainty range is therefore increased.

In a review of SQT emissions from vegetation, β -caryophyllene was found to be the most frequently reported SQT emitted and the most abundant SQT within many emission profiles (Duhl et al., 2008). Ozonolysis rate constant for β -caryophyllene is $1.16\times10^{-14}\,\mathrm{cm}^3\,\mathrm{molecule}^{-1}\,\mathrm{s}^{-1}$ (Shu and Atkinson, 1994) and while several SQTs have

high reported ozonolysis rate constants similar to β -caryophyllene (e.g., α -humulene, $1.2 \times 10^{-14}\,\mathrm{cm^3}$ molecule $^{-1}\,\mathrm{s^{-1}}$, others are nearly two orders of magnitude lower if sterically hindered (e.g., α -copaene, $1.6 \times 10^{-16}\,\mathrm{cm^3}$ molecule $^{-1}\,\mathrm{s^{-1}}$, Shu and Atkinson, 1994). The ozonolysis rate constant that is used for all SQT has been reported to be $3.3 \times 10^{-15}\,\mathrm{cm^3}$ molecule $^{-1}\,\mathrm{s^{-1}}$ (Bouvier-Brown et al., 2008) and is in the same order of magnitude but smaller than the one applied in this study, because of the different SQT mixture found.

Based on the above approach, the measured data was corrected for ozone reaction losses by considering the individual reaction rate constants for the aforementioned SQT, weightened by their individual contribution to the emissions. Further corrections have been applied for deposition $(k_{dep} = (2.64 \pm 0.23) \times 10^{-5} \text{ s}^{-1})$ and dilution $(k_{\rm dil} = (2.9 \pm 0.2) \times 10^{-3} \, {\rm s}^{-1})$ losses for both compounds. Deposition rates were calculated by the exponential decline in a closed cuvette for a three day period (no sources), while dilution rates were calculated considering the total flow that was drawn from the chambers to the analysers and as mentioned before, was replaced with ambient air at equal flow rate. The last data points with open cuvette were used to quantify the ambient mixing ratios of total MT and SQT. With respect to the corrections, the deposition and ozone loss rate had a minor impact on MT concentrations, indicating that the dilution is the major loss factor. For SQT however, the situation was different with the ozone reactions to be the dominating loss procedure while the deposition and dilution losses were substantially smaller. The general pattern assumed to apply for the entire season and possible changes in individual contribution would induce further uncertainties, that would not however impact the measured by PTR-MS total SQT concentrations in a critical way.

Biogenic emissions are usually quantified in ngg(dw)⁻¹ h⁻¹, where g(dw) is the needle dry weight mass in g. After the experiments the branch was cut and the needles were dried in order to measure their mass. During the growing period, frequent length measurements were conducted and the dry biomass was extrapolated from

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these measurements. The data up to 2 h after any length measurement were excluded because of the observed stress response of touching the branch.

3 Results

3.1 Driving forces

Under natural conditions, plants rarely experience single stress factors one by one, but are much more likely to be exposed to multiple stresses simultaneously. The role of volatiles in protection against thermal stress has been relatively well studied, while ozone was found to trigger the emissions of herbivore induced plant volatiles (Vuorinen at al., 2004) under controlled laboratory experiments. Light and drought stress have been considered as environmental factors that can also influence biogenic emissions. At this section we present the effects that these stress factors can have in SQT emissions. In a peculiar season, which was characterized by a warm and dry spring (precipitation anomaly < 25%; DWD), a wet summer (precipitation anomaly 126–150%; DWD) and an extremely dry autumn (precipitation anomaly < 50%; DWD), SQT emissions were analyzed and studied in order to discern the importance of different driving forces.

3.1.1 Temperature effect

SQT emissions clearly responded to temperature changes. The analysis of emission rates as a function of temperature was performed similar to Guether et al. (1995), as exponential function between temperature and SQT emissions:

$$E_{\text{SQT},T} = E_{\text{s}(T)} \cdot \exp(\beta \cdot (T - T_{\text{s}})) \tag{1}$$

Here, $E_{SQT,T}$ is the emission rate (in $ngg(dw)^{-1}h^{-1}$) observed at temperature T. $E_{s(T)}$ is the standard emission potential at $T_s = 30$ °C (Guether et al., 1995) and β (°C⁻¹) is an 7672

empirical coefficient called β -factor, that is based on the best fit curve between temperature and emission rate. T (°C) is the temperature inside the cuvette that on average was about one degree Celsius above ambient temperature. Applying our data to the above equation, we found that SQT emissions are correlated with ambient temperature

$$E_{SOT,T} = (347.2 \pm 70.4) \operatorname{ng} \operatorname{g(dw)}^{-1} \operatorname{h}^{-1} \cdot \exp((0.11 \pm 0.02)^{\circ} \operatorname{C}^{-1} \cdot (T - 30^{\circ} \operatorname{C}))$$
 (2)

As it is demonstrated in Fig. 1a, SQT emissions rise exponentially with the temperature, but with varying temperature dependency (slope = β -factor) and standardized emission potential at $30\,^{\circ}$ C ($E_{\rm s}$) during the year. The emission potential was found to be highest during spring $(E_{s(T),spring} = (533.8 \pm 61.7) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1})$ and declined during summer and autumn (352.9 \pm 56.1) and (175.9 \pm 23.7) ngg(dw)⁻¹ h⁻¹, respectively), with the latest values to be thrice lower than the spring ones. On contrary, β -factor showed different behaviour during the season, with higher values observed during summer $(\beta_{\text{summer}} = (0.12 \pm 0.02)^{\circ} \text{C}^{-1})$, followed by autumn ones $(\beta_{\text{autumn}} = (0.11 \pm 0.02)^{\circ}\text{C}^{-1})$. Temperature dependency found to be minimum during spring time $(\beta_{\text{spring}} = (0.09 \pm 0.01)^{\circ}\text{C}^{-1})$, mainly because of the low β -factors observed during May (Fig. 2), where a severe draught stress was present and possibly the tree was not able to extract water of the soil anymore, closing its stomata to prevent further water loss.

The exponential relationship between emissions and temperature has been used to describe MT emissions during the last two decades (Guenther et al., 1993) while recent reports (Tarvainen et al., 2005; Helmig et al., 2006, 2007; Hakola et al., 2006) have shown SQT with slightly stronger temperature dependence than MT. For this particular site MT emissions were correlated with the enclosure temperature as shown in Eq. (3):

$$E_{\text{MT},T} = (885.5 \pm 149.6) \,\text{ng} \,\text{g(dw)}^{-1} \,\text{h}^{-1} \cdot \exp((0.11 \pm 0.02)^{\circ}\text{C}^{-1} \cdot (T - 30^{\circ}C)) \tag{3}$$

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The basal emission potential $E_{\mathrm{M}}(T)$ was substantially higher for MT, but the β -factor was identical with the one for SQT, when using the complete dataset. As it is shown in Sect. 3.4, the β -factor is influenced by ambient ozone concentrations but only in the case of SQT.

3.1.2 Oxidative stress

Ambient ozone concentrations varied during the season notably, proving a wide range of oxidative stress that the ecosystem experienced. Total ambient mixing ratios varied between 3.6 and (96.2 ± 0.3) ppb_v, with a variation between 3 and (84.5 ± 0.3) ppb_v inside the cuvette. The 50% of annual measurements were situated between 23.2 to (57.1 ± 0.3) ppb_v. In general, spring values were (7.4 ± 0.3) ppb_v higher than the corresponding fourteen year averaged mixing ratios for this site. Contrariwise, the wet and cold summer resulted in average (8.5±0.3) ppb_v lower ozone mixing ratios. Autumn values were in line with the previous years, indicating an anomaly of only (1.0 ± 0.3) ppb_v.

Assuming a solely dependency between ambient ozone stress and emission, an exponential relationship ($R^2 = 0.50$; $R^2 = 0.38$ for linear) between ambient ozone mixing ratios and SQT emissions has been observed (Fig. 1b), with the ozone dependency (slope of the fitting) to be almost constant during the season. The direct relationship observed between SQT emission rates and ambient ozone mixing ratios can be explained by the fact that the selected branch was in the lowermost level of the canopy (Jardine et al., 2011). Furthermore, the benefits of our experimental setup along with the aforementioned corrections applied minimized any type of SQT emission losses. Detailed equations of the exponential relationship between SQT emissions and ozone can be found in the caption of Fig. 1, while the reader is referred to Sect. 4.1 for a detailed formulation of the assumed relationship.

In order to investigate possible light dependencies on SQT emissions, we separated our data in two individual categories (day and night) and plotted our results according to Eq. (1). Emission potential and temperature dependency were almost identical for both cases (Fig. 1c). Among the studies that have examined the light dependency of SQT emissions, there have been contrary findings, with evidence that some emissions are solely temperature-controlled while others are also affected by light (Duhl et al., 2008). However, each of the studies that did not observe light dependencies in SQT emissions was conducted in field conditions under ambient light and temperature regimes and comes in line with our finding. As pointed out from Jardine et al. (2011) SQTs are thought to mainly be produced in the cytosol via the MVA pathway, while MTs are thought to be mainly produced in chloroplasts via the MEP pathway. The di-

rect connection to recently assimilated carbon may result in a strong light dependence on de novo MT production/emission whereas de novo SQT production/emission may

3.1.4 Drought stress

be more temperature dependent.

3.1.3 Radiation effect

Relative humidity (RH) was found to be anticorrelated with SQT emissions (Fig. 1d). However, the effect of this parameter on SQT emissions was getting important only when RH was below 50 %. It is more likely that emissions were unaffected by mild drought conditions but were affected substantially during extended drought periods (May). Only 7.6% of our data recorded RH values below 50%. Under these conditions, SQT emissions rise substantially during spring and summer (Fig. 1d). Autumn conditions resulted in only few data points below this limit and therefore basal emission potential and relative humidity dependency were substantially lower. In every case SQT emissions were suppressed, when ambient RH conditions were high. The amount of released SQT almost never exceeded the value of 100 ng (g(dw)h)-1, when RH was maximum. This result indicates that small RH value can induce emissions but only

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under severe drought conditions, while water stress could negatively affect plant responses.

3.2 Daily emissions

A clear mean daily cycle for SQT emissions has been observed. Emission generally peaked at midday (1:00-4:00 p.m.) for all months but the rise was starting two hours earlier (8:00 a.m.) for all the months before August. A vast amount of daily derived β -factors has been obtained from regression analysis between enclosure temperature and SQT emissions (Fig. 2a). Calculated β-factors ranged for 0.02 to 0.27 (°C⁻¹) with no distinct seasonal pattern. Highest values were observed during April and early summer, while the lowest ones were observed during the wet August (precipitation anomaly = 130-150 %, DWD). This can be an indication for the importance of water vapour and a detailed discussion about this issue can be found in Sect. 4.3. Autumn variations cover the whole spectrum, producing difficulties in understanding any possible seasonality of temperature driven emissions. During the growing period (end of April/beginning of May) β -factors ranged from 0.03 to 0.07 °C⁻¹, suggesting that temperature effect was weak, when new twigs were growing and other processes were taking place inside the leaves governing the emissions. The wide spectrum of β -factors indicates that temperature stress should not be considered as the single stress factor inducing SQT emissions, and that other environmental and biotic factors can amplify the temperature driven emissions.

Daily calculated emission potentials $E_{s(T)}$ can be seen in Fig. 2b. Highest $E_{s(T)}$ were observed during spring and an exponential decline was observed while moving towards the end of the year. Our results show a decline of SQT $E_{s(T)}$ for every next month, apart from the transition from October to November. If SQT are stored inside some pools and used from the tree when required, the extremely dry November (historic record for Germany since 1900, DWD) can explain this rise in the basal emission rates. During the growing period $E_{s(T)}$ was dramatically reduced as was the β -factor. An explanation for

the reported low values could be that newly grown leaves have different SQT capacities and therefore react different to environmental factors until they become mature.

3.3 Correlation coefficients

The vast variations that have been extrapolated from the aforementioned regression analysis emphasize the need to investigate in detail the possible driving forces for SQT emissions. Figure 3 displays the three observed major driving forces and how they correlate with SQT emissions in time. Accounting in total 16200 emission data points, temperature showed a correlation of (0.47 ± 0.02) and RH an anticorrelation $-(0.53 \pm 0.01)$, indicating that RH can suppress SQT emissions.

Interestingly, one can observe a stronger dependency (0.63 ± 0.01) with ambient ozone concentrations, with the maximum correlation values 2–4 h (flat behaviour) before the emission start. Nevertheless it should be kept in mind that ozone and temperature are not independent parameters. It is not the first time that a delay in ozone triggering emissions has been reported. However Sharma et al. (1994) fumigated *thaliana* leaves and observed 3 h delay in the synthesis of phenylpropanoid compounds while Heiden et al. (1999) reported enchased SQT concentrations 5 h after fumigating an ozone resistant variety of Nicotiana tabacum with 170 nmol mol⁻¹ O_3 . This time frame can vary depending on the tree species and on the amount of ozone concentration that it is exposed. High ozone concentrations (> 60 ppb_v) resulted in maximum correlation coefficients 4–5 h before the emission, while low ozone concentrations resulted in poor correlation coefficients (0.24 ± 0.03) without a time delay. It might be that low ozone concentrations are still correlated with the emissions, because of their strong dependency on temperature. This indicates that ambient ozone concentrations can be substantially harmful for Norway spruce above a critical threshold.

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3.4 Ozone regimes

For investigating possible thresholds of the ozone concentrations, we separated our ozone data in ten different regimes with the same probability in between the limits of 10% (Gaussian distribution). Our aim was to investigate the strength of temperature and oxidative stress under different ozone conditions.

Correlation coefficients were calculated individual for each regime for both temperature and ozone concentrations (Fig. 4). In clear conditions, when ozone concentrations were below (22 \pm 0.3) ppb $_{v}$, temperature was much stronger correlated with SQT emissions. Nevertheless, the values were substantially changing, when we were taking account more polluted regimes. Temperature found to be the predominant driving force for the first four regimes (upper O_3 regime limit = (36.6 ± 0.3) ppb_v) with decreasing strength. On contrary correlation coefficients between SQT emissions and ozone were small for the first three regimes with the tendency to increase. For ozone concentrations between (36.6 and 52.2 ± 0.3) ppb, the strength of these two driving forces was relative stable, with the indication that oxidative stress was more important than the heat stress. This spectrum of ozone volume mixing ratios is representative for the oxidative stress that the ecosystem at Mt. Kleiner Feldberg is experiencing (43.3% probability for the last 11 yr, HLUG), although common extremes during July and August were not achieved due to weather conditions in 2011. The last regime includes concentrations above (58.5 \pm 0.3) ppb $_{\rm v}$ O $_{\rm 3}$ and one can clearly see a steep rise in ozone correlation coefficient. When ozone is substantially high, it displays the predominant driving force and triggers SQT emissions with the reported delay.

In Fig. 5 we obtained the β -factors and emission potentials ($E_{\rm s,T}$), derived from regression analysis between enclosure temperature and calculated emissions at each regime. For intercomparison reasons we also demonstrate the behaviour of total MT emissions, analysed with the same approach. Figure 5a shows a different behaviour on β -factors for SQT and MT, respectively. For MT $\beta_{\rm MT}$ was relative constant through the complete ozone spectrum and close to the empirical value of 0.09 °C⁻¹ which is

usually used in biogenic emission modelling studies (Guenther et al., 1995, 2006). On contrary, β_{SQT} is showing a different behaviour at different regimes, with a possible threshold above (36.6 ± 3.9) ppb_v. SQT β -factor is slightly higher than MTs one (Tarvainen et al., 2005; Helmig et al., 2006, 2007; Hakola et al., 2006), but for ambient ozone mixing ratios fall inside the first two regimes (below (28.2 ± 0.3) ppb_v). When oxidative stress was becoming stronger, the temperature dependency expressed by β -factor was dropping down to very small values. Interestingly, minimum β_{SQT} were calculated for the regimes that fall inside the range of ozone concentrations that have the highest probability to occur at this site.

Emission potentials showed a similar behaviour. $E_{\rm s,MT}$ was almost constant for the majority of ozone regimes, with the highest values to be observed at the edges of very clear and very polluted conditions. Regression tree analysis (Breiman et al., 1984) between ozone concentrations and MT emissions showed that MT are emitted in larger amounts when ambient ozone concentrations are above (54 ± 4) ppb_v. Since MT are not considered to be critical for oxidative stress, we assume that the high temperatures associated with high ozone are the main reason for this observed rise. On contrary, $E_{\rm s,SQT}$ displayed a dynamical well along the ozone spectrum and the lowest values were once more obtained for the range with the most representative ozone concentrations for the ecosystem at this site. For the regimes above 47.7 ppb_v, $E_{\rm s,SQT}$ rise linearly with ozone, stressing out the importance of this environmental parameter as a driving force of SQT emissions.

4 Discussion

4.1 SQT emission algorithm

The relationship between plant stress and VOCs has been studied for several years in many ways, but stresses have usually been considered as single, independent factors. The review of Holopainen and Gershenzon (2010) has shown that the interaction

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of multiples stresses has great potential to alter VOC emission and that additive effect of stresses on VOC emissions might also be expected for combination of abiotic stresses, including high temperature and ozone, both of which induce higher VOC emission rates when applied singly. BVOC emissions are typically modelled with either temperature-dependent or temperature- and light-dependent algorithms (e.g., Guenther et al., 1995). Most emission models assume that the majority, if not all, of MTs are emitted as a function of temperature, but due to limited studies, SQT data is either eliminated from global models or vast assumptions have to be made about their emission drivers (Bouvier-Brown et al., 2008). Nevertheless, the involvement of plant volatiles in destroying ozone may have a significant impact on the interpretation of the role of such emissions (Jardine et al., 2011). Previous studies estimated that total ozone fluxes to plant canopies in mid-latitude forests can be dominated by gas-phase chemistry and not by stomatal uptake as generally assumed (Fowler et al., 2001; Kurpius and Goldstein, 2003).

Our results showed a clear correlation between SQT emissions and ambient ozone concentrations, suggesting that this environmental parameter should not be neglected in SQT emission model studies. We suggest a similar parameterization with the one that is used for MT (Guenther et al., 1993) but with an additional term that includes the observed exponential relationship between ozone and SQT emissions:

$$E_{SQT,T} = E_{SQT,T} + E_{SQT,O_2}$$
 (4)

 $E_{\text{SOT},T}$ is the emission term that describes the effect of heat stress on SQT emissions

$$E_{\text{SQT},T} = E_{\text{s},T} \cdot \exp(\beta \cdot (T - 30)) \tag{4a}$$

and E_{SQT,O_3} is the additional term that is used to describe the oxidative stress:

$$E_{\text{SQT},O_3} = E_{\text{s},O_3} \cdot \exp(\sigma \cdot ([O_3] - 40 \text{ ppb}_{\text{v}})) \tag{4b}$$

25 In detail, SQT emissions can be paramerizised as:

$$E_{\text{SQT}} = E_{\text{s,T}} \cdot \exp(\beta \cdot (T - 30)) + E_{\text{s,O}_3} \cdot \exp(\sigma \cdot ([O_3] - 40 \text{ ppb}_{\text{v}}))$$

$$(5)$$

where β (°C⁻¹) is an empirical coefficient called β -factor, T is the temperature inside the cuvette (in °C), $E_{s,T}$ (ng(g(dryweight)h)⁻¹) is the emission potential at $T_s = 30$ °C (Guether et al., 1993), E_{s,O_3} (ng(g(dryweight)h)⁻¹) is the emission potential at 40 ppb_v mixing ratio of ozone, σ (ppb_v⁻¹) is an empirical coefficient called σ -factor (sigma factor) and [O₃] is the ambient ozone mixing ratio in ppb_v. We chose to use the value of 40 ppb_v O₃ for the standard basal emission potential E_{s,O_3} due to oxidative stress, because of the critical threshold observed and presented at Sect. 3.4. Below this value, the second term of Eq. (3) has a minor contribution in total SQT emissions but in higher concentrations, the importance of this pollutant as a driver of SQT emissions is magnified because of the exponential term in the equation.

SQT emissions with representative values for the site of interest (Fig. 6) are in form of Eq. (5):

$$E_{SQT} = (347.2 \pm 70.4) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp((0.11 \pm 0.02)^{\circ} \text{C}^{-1} \cdot (T - 30^{\circ} \text{C}))$$

$$+ (52.5 \pm 6.9) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp((0.05 \pm 0.01) \, \text{ppb}_{v}^{-1} \cdot (([O_{3}] - 40) \, \text{ppb}_{v}))$$
 (6)

Thus, ozone stress has a minor impact in vegetation under clear conditions (Sects. 3.3 and 3.4) and therefore $E_{\rm SQT,O_3}$ should be small in remote forests. Nevertheless, we observed (Fig. 5) that SQT emission responces to temperature can attenuate under higher ozone concentrations. This trend has already been hypothesized by Staudt et al. (2011), according to whom it is most likely that other environmental factors such as drought, ozone and $\rm CO_2$ concentrations can amplify or attenuate such alterations in the emission responses to temperature. An empirical formulation of SQT emissions, such the one that we suggest, cannot explain all the details of the emission profile of these compounds but it broadens our understanding in the tree responses under natural conditions.

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4.2 Intercomparison between emission algorithm and observations

The algorithm that considers temperature as the only driving force (Eq. 1) found to substantially underestimate SQT emissions during the year (Fig. 7). On contrary, when we consider additionally the oxidative stress, the fitting between the measured and calculated values (Eq. 5) improves considerably. Nevertheless, a so far neglected issue when modelling VOC emissions are the observed changes (Fig. 1, Sect. 3.1) at empirical coefficients (β -, and σ -factor) and basal emission potentials ($E_{s,T}$, E_{s,O_3}) along the season. Using Eq. (4) with the aforementioned parameters individual for each season, the linear fit between observations and the suggested emission algorithm is close to 1:1 line. In any case the model overestimates the measured emissions but not in a critical way. This deviation is most likely attributed to the effect of RH, which was excluded from the algorithm even if a tentative effect on SQT emissions was observed (Fig. 1d). A detailed explanation about the role on RH on SQT emissions is discussed at the last section of this chapter.

Coupled with the presented linear fit between the measured SQT emissions and the modeled ones, a satisfactory fit was observed across the complete timeline of the measurements (Fig. 8). There are only two periods during the year, where a significant mismatch is apparent, while elsewhere the algorithm can efficiently capture SQT emissions. One period is during the growing season at the end of April – beginning of May. Twigs length was measured continuously during this period and dry mass weight was derived from these measurements. New grown leaves have different pools and therefore different capabilities in emitting SQTs (Guenther et al., 2006). The second period that an overestimation of SQT emission was observed is located at the second half of August but a clear explanation about this observation cannot be provided. We assume that, as Holopainen and Gershenzon (2010) stated at their review, that the co-occurrence of multiple stresses may significantly limit energy or nutrient supplies, and so it may not be possible for a plant to respond to every stress factor impinging on it at a single time. Additionally, SQT are usually stored before emissions and these

4.3 The neglected role of relative humidity

The controversial role of water stress in affecting the physiological response of plants when exposed to O_3 has been reported by Vitale et al. (2008). Ribas et al. (2005) found that a severe water stress could negatively affect the O_3 responses of Q. ilex, while more studies (Karlsson et al., 2002; Khan and Soja, 2003) emphasized the ability of water vapour to decrease the negative impact of O_3 in several plant species. These findings seem to explain the observation that our model overestimates SQT emissions under 100 % of RH (Fig. 9). This could also explain the observed deviations presented in Fig. 7, despite the linearity between modelled and measured emissions.

storage pools are expected to decrease during the year. This applies especially at intense stress levels and only moderate production rates may expected thereafter.

Our findings suggest that this parameter is influencing SQT emissions but with a significant impact only under specific conditions. This might indicate different storage pools of sesquiterpenes within a single tree, which are used dependent on ambient conditions. Hence, further developments and field studies are essential in order to achieve a better understanding on SQT emissions from vegetation in ambient atmosphere.

5 Conclusions

In this study we found that SQT emission rates were clearly correlated with temperature, with an exponential function between temperature and emissions, similar to the one that is used for MT (Guenther et al., 1995). However, a similar link has been observed to ambient ozone mixing ratios, with even higher correlations coefficients than for temperature, when considering the annual dataset. A critical threshold above which ozone is the major driving force of SQT emissions was derived. We postulate a new parameterization which includes both, i.e. the heat and the oxidative stress for a future

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application in global simulation models. Bäck et al. (2011) found that in contrary with MT, SQT emissions do not defer significantly between the investigated coniferous trees (Scots Pine) and therefore smaller uncertainties would be induced when applying the algorithm. However, more measurements are necessary in order to confirm the validly of this observation in other ecosystems. Finally, we observed the ability of water vapor to decrease the stress responses of the investigated tree under 100 % RH and therefore this parameter should not be neglected in any studies dealing with SQT emissions from vegetation.

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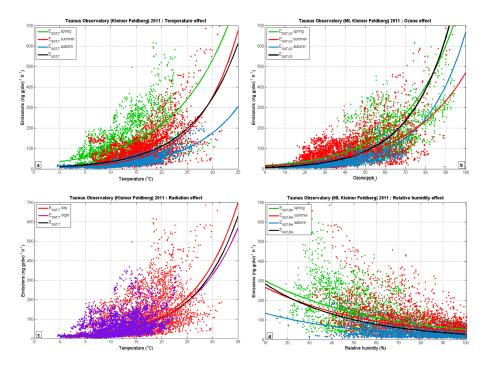










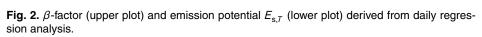


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Fig. 1. Emission responses to different stress factors. (a) (temperature): $E_{SQT,T} =$ $(347.2 \pm 70.4) \, \text{ng} \, \text{g(dw)}^{-1} \, \text{h}^{-1} \cdot \text{exp}((0.11 \pm 0.02)^{\circ} \text{C}^{-1} \cdot (T - 30^{\circ} \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})), \\ E_{\text{SQT},T}(\text{spring}) = (533.8 \pm 0.02)^{\circ} \, \text{C}^{-1} \cdot (T - 30^{\circ} \, \text{C})$ 61.7) $\operatorname{ngg}(\operatorname{dw})^{-1} \operatorname{h}^{-1} \cdot \exp((0.09 \pm 0.01)^{\circ} \operatorname{C}^{-1} \cdot (T - 30^{\circ} \operatorname{C})), E_{\operatorname{SQT},T}(\operatorname{summer}) = (352.9 \pm 0.01)^{\circ} \operatorname{C}^{-1} \cdot (T - 30^{\circ} \operatorname{C})$ 56.1) ngg(dw)⁻¹ h⁻¹ · exp((0.12 ± 0.02)°C⁻¹ · (T – 30°C)), $E_{SQT,T}$ (autumn) = (175.5 ± 61.7) $\operatorname{ngg}(\operatorname{dw})^{-1}\operatorname{h}^{-1}\cdot \exp((0.11\pm0.02)^{\circ}\operatorname{C}^{-1}\cdot(T-30^{\circ}\operatorname{C})).$ **(b)** (ozone): $E_{SQT,O_3} = (7.1 \pm$ 1.3) $\operatorname{ngg(dw)}^{-1} \operatorname{h}^{-1} \cdot \exp((0.05 \pm 0.01) \operatorname{ppb}_{v}^{-1} \cdot [O_{3}]), E_{\operatorname{SQT,O}_{3}}(\operatorname{spring}) = (16.4 \pm 1.9) \operatorname{ngg(dw)}^{-1} \operatorname{h}^{-1} \cdot [O_{3}]$ $exp((0.04 \pm 0.01) ppb_v^{-1} \cdot [O_3]), E_{SQT,O_3}(summer) = (14.2 \pm 3.1) \, ngg(dw)^{-1} \, h^{-1} \cdot exp((0.04 \pm 0.01) \, ngg(dw)^{-1}) \, h^{-1} \cdot exp((0.04$ $0.01) \, ppb_v^{-1} \cdot [O_3]), E_{SQT,O_3}(autumn) = (4.5 \pm 0.7) \, ng \, g(dw)^{-1} \, h^{-1} \cdot exp((0.05 \pm 0.01) \, ppb_v^{-1} \cdot 1) \, dv$ $[O_3]). \quad \textbf{(c)} \quad (radiation): \quad E_{SQT,T}(day) = (365.1 \pm 77.1) \, ng \, g(dw)^{-1} \, h^{-1} \cdot exp((0.12 \pm 0.02)^{\circ} C^{-1} \cdot exp((0.12 \pm 0.02))^{\circ} C^{-1}$ $(T-30^{\circ}\text{C})$, $E_{\text{SQT},T}(\text{night}) = (327\pm57.4) \, \text{ng} \, \text{g(dw)}^{-1} \, \text{h}^{-1} \cdot \exp((0.11\pm0.02)^{\circ}\text{C}^{-1} \cdot (T-30^{\circ}\text{C}))$. (d) (rel. humidity): $E_{SQT,RH} = (409.1 \pm 84.1) \text{ ng g(dw)}^{-1} \text{ h}^{-1} \cdot \exp(-(0.03 \pm 0.01) \cdot \text{RH[\%]}), E_{SQT,RH} (\text{spring}) = 0.001 \cdot \text{RH[\%]}$ $(355.2 \pm 45.2) \text{ ngg(dw)}^{-1} \text{ h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), E_{\text{SOTRH}}(\text{summer}) = (338.9 \pm 0.01) \cdot \text{RH[\%]}$ $62.7) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 31.8) \, \text{ngg(dw)}^{-1} \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 0.01) \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{RH[\%]}), \\ E_{\text{SQT,RH}}(\text{autumn}) = (163.9 \pm 0.01) \, \text{h}^{-1} \cdot \exp(-(0.02 \pm 0.01) \cdot \text{h}^{-1}) \,$ $\exp(-(0.02 \pm 0.01) \cdot RH[\%]).$



β - factor



July

June

July

Taunus Observatory (Mt. Kleiner Feldberg) 2011 : Daily values

August

August

September

0.3

0.25

0.05

2700 2400 2100

> 300 April

Es (ng g(dw) 1 h-1)

April

β - factor (K⁻¹)



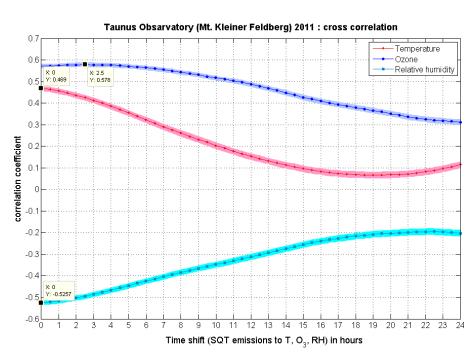


Fig. 3. Cross-correlation of SQT emission and temperature (red), ozone (blue) and relative humidity (cyan), with a time shift between the emissions and the individual stress factors.



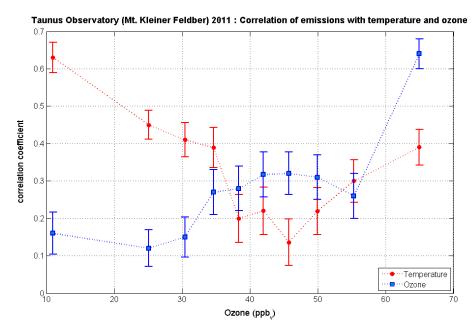


Fig. 4. Correlation coefficients for temperature and ozone calculated individually for ten different ozone regimes. In moderately or less polluted atmospheric conditions SQT emissions were better correlated with temperature, while in more polluted atmospheric conditions the emissions were better correlated with ambient ozone mixing ratios, indicating a critical threshold in the parameter that is driving the emissions.

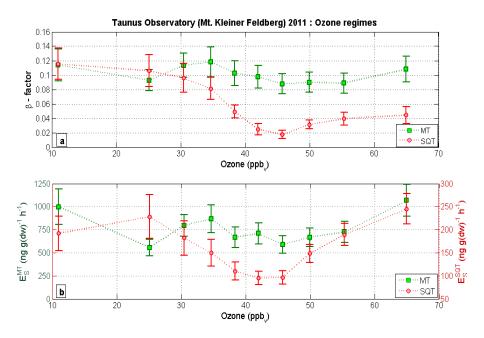


Fig. 5. β -factor (upper plot) and emission potential $E_{s,T}$ (lower plot) for MT (green squares) and SQT (red cycles), derived from regression analysis for ten individual ozone regimes with the same probability. eta_{MT} and emission potential ($E_{\mathrm{s,MT}}$) were almost constant along the complete ozone spectrum with no significant variation is visible. On contrary, β_{SQT} display smaller values after the fourth ozone regime. The respective emission potential $(E_{s,SQT})$ is displayed as a dynamical well along the ozone spectrum, with the lowest values to be observed for the range with the most representative ozone concentrations for the site.



Temperature (°C) Fig. 6. Pattern of the derived SQT emissions, parameterized for the site of interest.

Sesquiterpene emissions

Ozone (ppb,

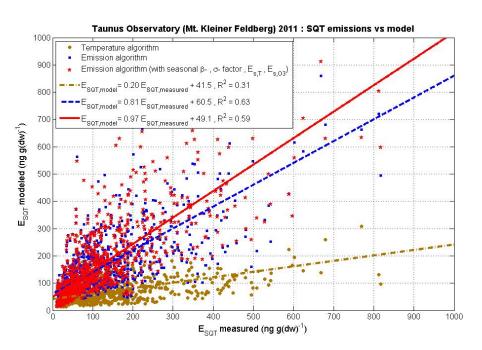


Fig. 7. Comparison between measured and calculated SQT emissions. An only temperature dependent algorithm substantially underestimates the measured emissions (gold dashed line). The addition of the ozone depended term (blue dashed line) is improving the fitting between measured and modeled emissions, while the best fit (red line) is observed when the seasonal β - and σ -factors and emission potentials ($E_{s,T}$, E_{s,O_3}) are considered in the algorithm.



Discussion Paper | Discussion Paper



Discussion Paper | Discussion Paper

Discussion Paper

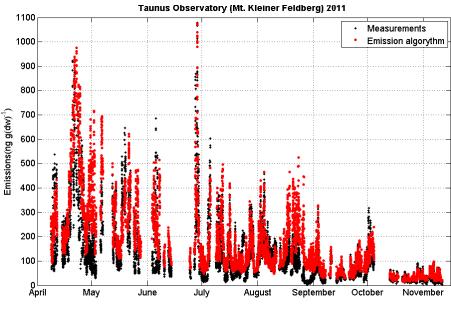


Fig. 8. Temporal pattern of the measurements compared with the new emission algorithm.

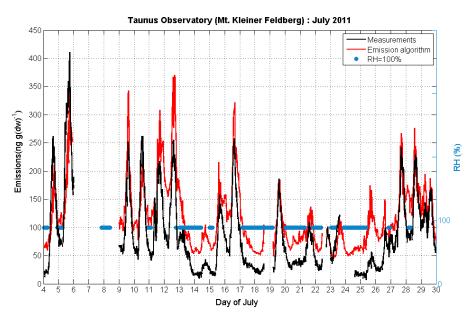


Fig. 9. Example of how RH can influence SQT emission rates derived from Eq. (5). In case of RH = 100 %, the algorithm fails, since the thin water layer created above the needle surface acts in protective way for the involved stresses.