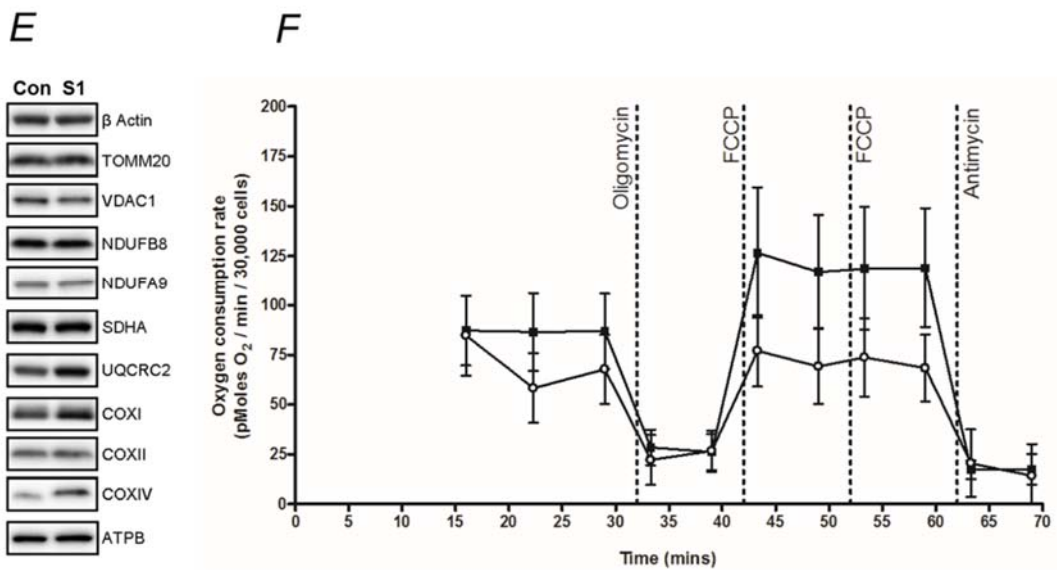
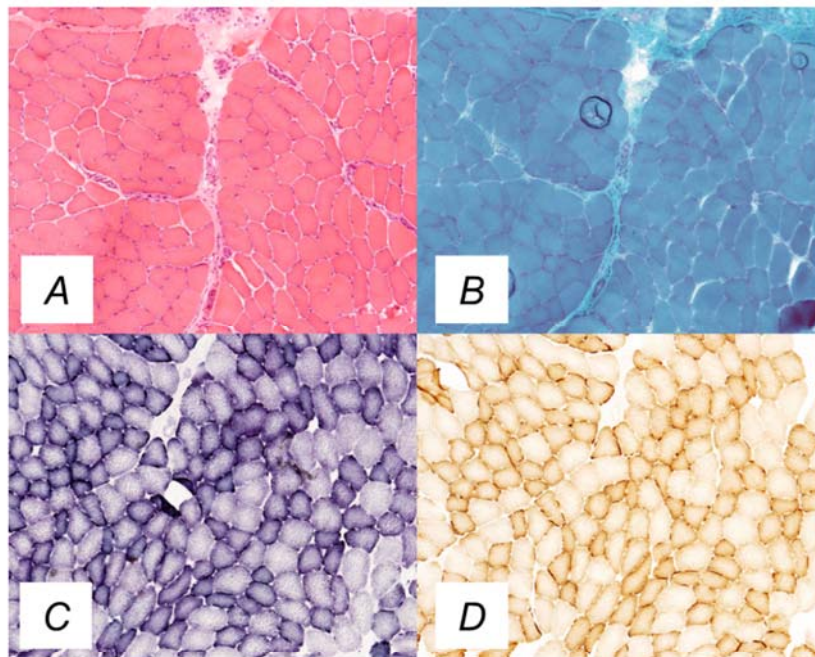


**Supplemental Data**

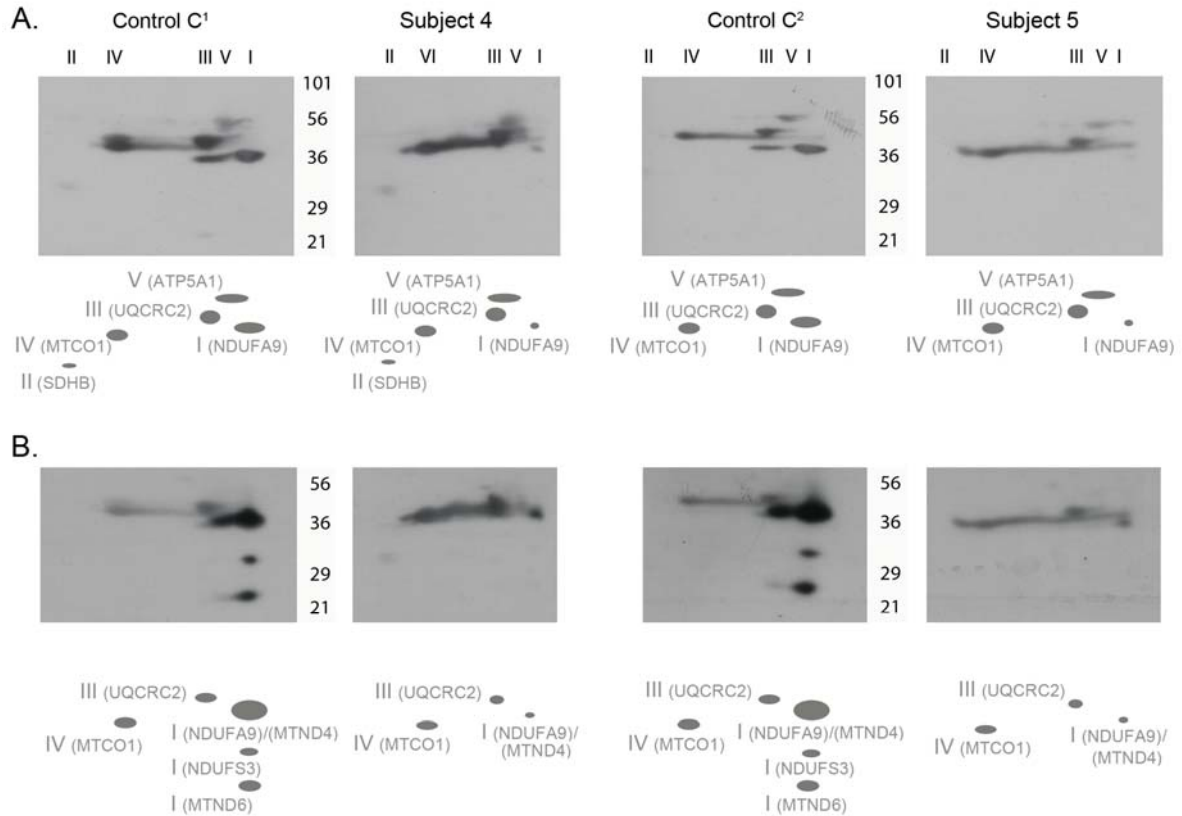
**Biallelic Mutations in *TMEM126B* Cause Severe**

**Complex I Deficiency with a Variable Clinical Phenotype**

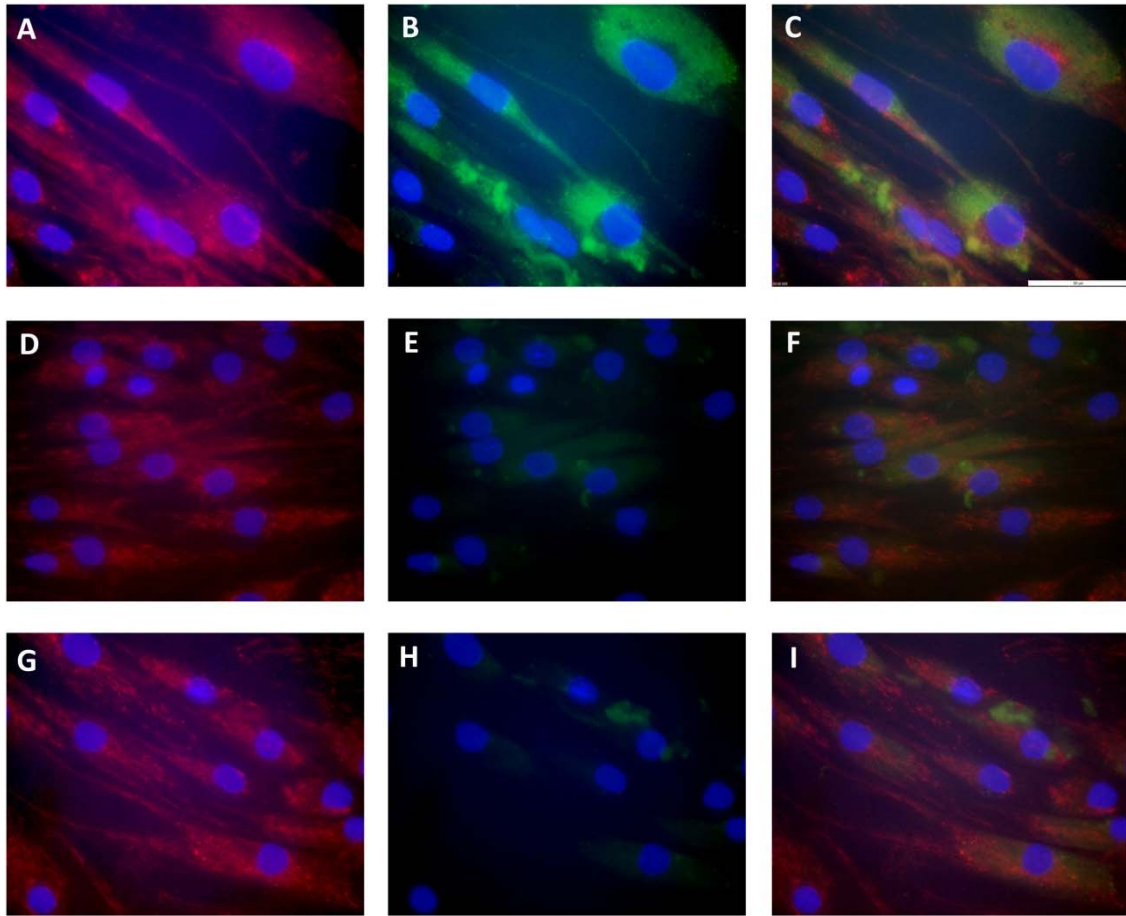
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**Figure S1: Functional characterization of muscle and cells from Subject 1**  
 Histopathological analysis of a serially-sectioned skeletal muscle biopsy from Subject 1 (homozygous p.(Gly212Val) *TMEM126B* variant) showing (A) H&E staining, (B) modified Gomori Trichrome staining, (C) succinate dehydrogenase (SDH) and (D) cytochrome c oxidase (COX) reactions highlighting evidence of subsarcolemmal mitochondrial accumulation. Interestingly, subject fibroblasts did not show a significant OXPHOS defect, either based on immunoblotting of fibroblast mitochondrial proteins for OXPHOS components (E) or micro-scale oxygraphy analysis (Subject 1, n=10, white circles) compared to the combined data of control cell lines (n=5, black squares; Experimental details are described in detail previously<sup>1</sup>) although overall rates of oxygen consumption did appear to be generally decreased (F). Error bars indicate the standard deviation.

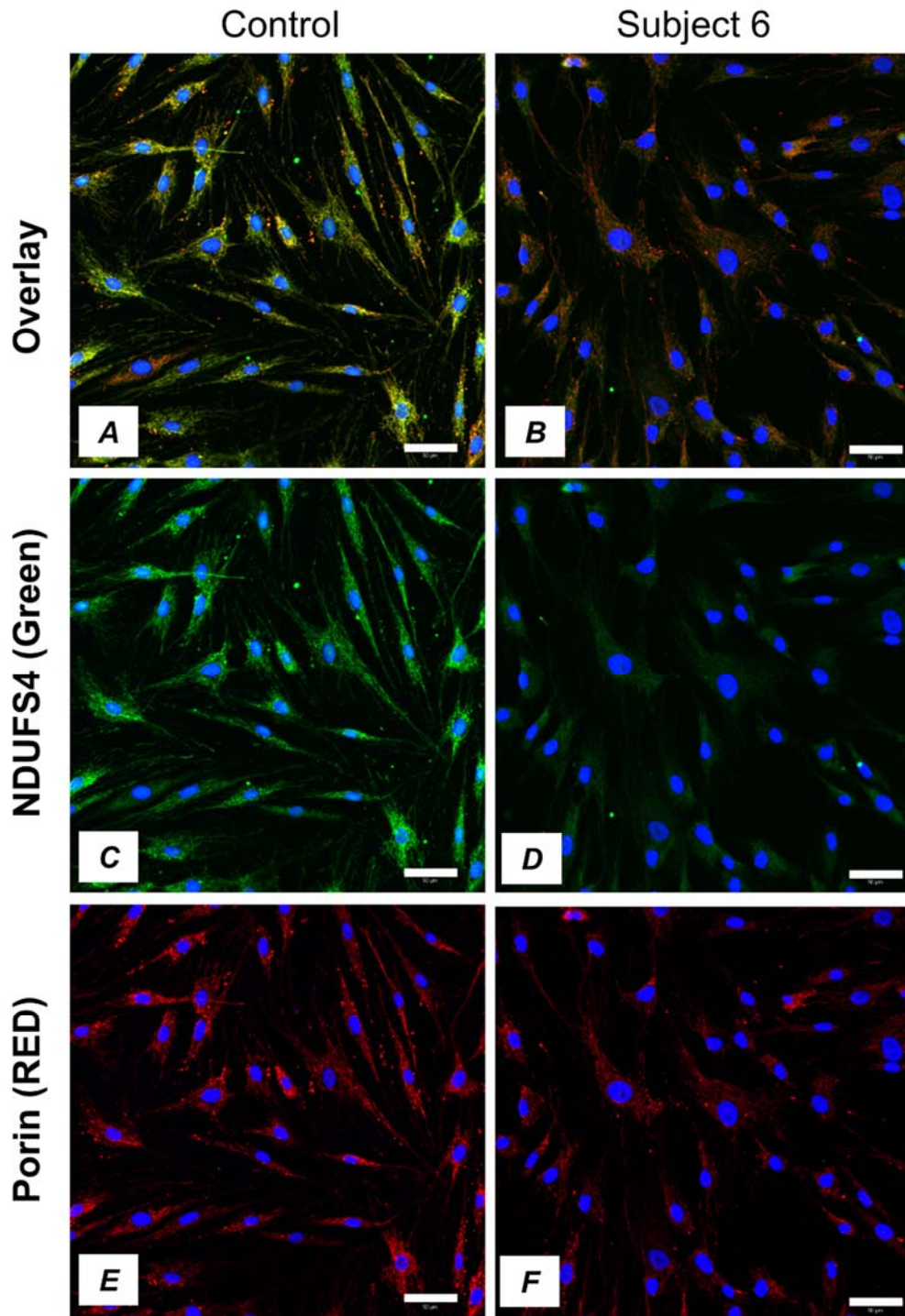


**Figure S2: Western blotting following 2D BN-PAGE/tricine SDS-PAGE separation of isolated skeletal muscle mitochondria from Subjects 4 and 5**  
 Experiments were performed according to the procedures previously described in detail<sup>2</sup>. **(A)** A mixture of the following antibodies was used to evaluate the abundance of the five OXPHOS protein complexes: complex I (NDUFA9), complex II (SDHB), complex III (UQCRC2), complex IV (MTCO1) and complex V (ATP5A1). **(B)** Following stripping of the antibodies, the nitrocellulose blot was reprobed using antibodies for complex I (MTND4, NDUFA9, NDUFS3 and MTND6), for complex III (UQCRC2) and for complex IV (MTCO1). An almost complete absence of signal with antibodies directed to the different complex I subunits is observed in skeletal muscle mitochondrial isolates from Subjects 4 and 5, highlighting a severe disturbance in the assembly of this OXPHOS complex.



**Figure S3: Cultured skin fibroblasts from Subjects 4 and 5 show severely decreased TMEM126B immunofluorescence**

Double immunofluorescent staining of fibroblasts<sup>3</sup> from a control (**A-C**) and from Subject 4 (**D-F**) and Subject 5 (**G-I**) was performed, using MitoTracker Red CMXRos (Invitrogen) shown in red (panels **A**, **D** and **G**) and rabbit polyclonal anti-TMEM126B (AV49321, Sigma; 30µg/ml 2h room temperature) visualized with donkey anti-rabbit AlexaFluor488 (Invitrogen) shown in green (panels **B**, **E** and **H**). Cell nuclei were counterstained with dapi shown in blue. The overlays (panels **C**, **F** and **I**) demonstrate a reduction of TMEM126B staining in cells from both subjects (Scale bar = 50um).



**Figure S4: Cultured skin fibroblasts from Subject 6 express a complex I defect**  
 Immunofluorescence staining of fibroblasts obtained from Subject 6 (homozygous p.(Gly212Val) *TMEM126B* variant) and a control was performed<sup>4</sup> using the following primary antibodies: mouse monoclonal anti-NDUFS4 antibody (1:100; Abcam, Cambridge, UK) shown in green (panels **C** and **D**) and rabbit polyclonal anti-VDAC1 (1:500; Abcam, Cambridge, UK) shown in red (panels **E** and **F**), with the overlay (panels **A** and **B**) clearly demonstrating strong staining of the NDUFS4 (complex I) protein in the control and absence in the patient cell line (Scale bar = 50um).

## References

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