

9. Supplementary Material

9.1. Optional reformatting of read names after *flexbar*

```
990 zcat <sampleX.fasta.gz> |  
  
    awk '{ if (FNR%4==1) {  
          if (NF==1) {  
995             print;  
          } else {  
              n=split($NF,v,"-");  
              printf "%s_%s", $1, v[n];  
              for (i=2; i<NF; i++) {  
1000                 printf "_%s", $i;  
              }  
              printf "_";  
              for (i=1; i<(n-1); i++) {  
1005                 printf "%s_", v[i];  
              }  
              print v[n-1];  
          }  
          } else {  
              print  
1010          }  
      }' | gzip > <sampleX.fasta.gz>
```

9.2. Supplementary Data Files

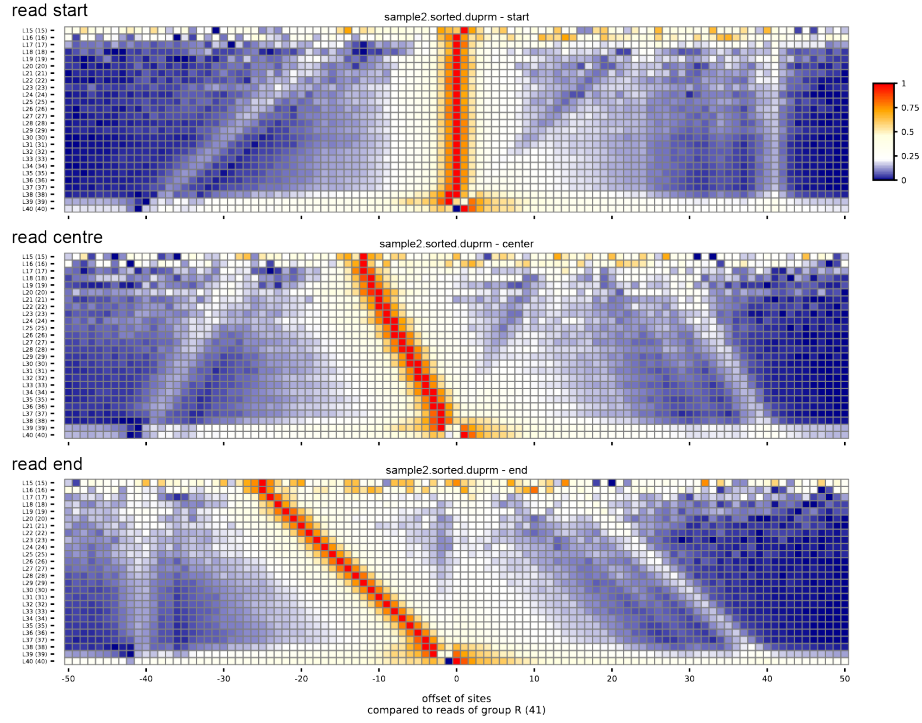
Supplementary Data 1: Bash code for all steps from raw reads

1015 **until peak calling.** This file provides the **bash** code for basic read processing, conversion into crosslink events and peak calling as described in Chapters 3-5.1. Details on the input files, preset variables and external tools required to run this code are listed in Chapters 3.1 and 2.1.

Supplementary Data 2: R code for postprocessing of PureCLIP out-
1020 **put.** This file provides the R code to postprocess the output of PureCLIP peak
calling as described in Chapter 5.2. Running this code requires a bed file with
'crosslink sites' output by PureCLIP (`PureCLIP.crosslink_sites_short.bed`)
and bw files with crosslink events (`sampleX.strand.bw`). The code to obtain
these files is described in Supplementary Data 1 and Chapter 4.2.

1025 **Supplementary Data 3: R code for reproducibility and downstream**
analysis. This file provides the R code to reproducibility analyses and assign-
ment of genes and transcript regions as described in Chapters 6.1 and 6.2.
Running this code requires a bed file with binding sites (e.g. curated PureCLIP
output, see Chapter 5), a gtf file with gene/transcript annotations, and bw
1030 files with crosslink events in the individual replicates (`sampleX.strand.bw`, see
Chapter 4.2).

9.3. *Supplementary Figure*



Supplementary Figure 1: High-resolution read overlap heatmaps for sample 2 in our dataset, generated with iCLIPPro. x-axis shows distance of start, centre and end positions of iCLIP reads that were trimmed to a given length (y-axis) relative to the start positions of reads that were not trimmed as reference positions (41-nt length, see Figure 5). In absence of read-end constraints, most reads starts align at the reference positions, irrespective of their length [9], whereas read centres and ends form a diagonal reflecting the read length after trimming. For more details on the underlying analysis steps and the visualisation, see [20] and <http://www.biolab.si/iCLIPPro/doc/>.