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Reporting Summary

Statistics

X Life sciences

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For all statistical analyses	s, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.					
n/a Confirmed						
☐ ☐ The exact samp	\times The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement					
A statement on	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly					
The statistical te	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.					
A description of	all covariates tested					
A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons						
A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)						
For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.						
For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings						
For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes						
Estimates of effe	ect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated					
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.					
Software and co	de					
Policy information about	availability of computer code					
	stepOne Software v2.3; EthoVision XT 8; PyroMark ID 1.0 (database version 23.2); PyroMark CpG Software 1.0.11; HEKA Patchmaster x90.4; NIKON Imaging Software NIS-Elements 4.5					
,	Graphpad Prism v7; StepOne Software v2.3; EthoVision XT 8; PyroMark ID 1.0 (database version 23.2); PyroMark CpG Software 1.0.11; HEKA Fitmaster 2x90.4; Wavemetrics IGOR Pro 8					
	algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. position in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.					
Data						
- Accession codes, unique - A list of figures that ha	clude a <u>data availability statement</u> . This statement should provide the following information, where applicable: ue identifiers, or web links for publicly available datasets					
No restrictions on data avai	lability					
Field-specif	ic reporting					

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Ecological, evolutionary & environmental sciences

Behavioural & social sciences

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

For behavioral experiments, a justifiable number of mice was used. Sample size evaluation based upon: Dell R.B., Holleran S., Ramakrishnan R. (2002), ILAR J. 2002;43(4):207-13.

For in vitro experiments, no statistical methods or calculations were used to predetermine sample size. The sample size was determined to be sufficient based on the size and consistency of the measurable differences between the groups.

Data exclusions

In general, outliers were identified with the Grubbs' test (Graphpad Prism). In the circadian rhythm experiments, several mice were excluded due to insufficient tracking in the light phase (low contrast between mouse and background). Tracking in the dark phase was overall successful (high contrast). One locus coeruleus sample due to insufficient tyrosine hydroxylase expression was excluded from analysis (Fig. 2c)

Replication

Mice from different litters and biological replicates were used in this study, unless stated otherwise.

Behavioral analysis: Experiments without injections of mice were performed in 7 individual sessions (Fig.1), experiments with injections of mice in 9 individual sessions (Fig.5)

RT-qPCR analysis: 3 technical replicates of one single sample were arithmetically averaged and integrated as one biological replicate in statistical analysis.

Randomization

Mice were initially genotyped and randomly divided into experimental groups with equal amount of mice per group. Male mice used in behavioral experiments were age-matched. Several breeding-dependent litters were utilized.

Blinding

Y-maze experiments and analyses were performed blinded in terms of genotype and treatment.

Circadian rhythm experiments were not performed blinded, but with an equal distribution of mice of different groups per run, which were analyzed simultaneously (restricted number of experimental setups). Several sessions were performed with all groups represented within each session.

Electrophysiological experiments were performed blinded.

In vitro experiments were not performed blinded. If investigators were not blinded to genotypes and treatment, the reported data were not subjective but rather computer-produced quantitative measurements.

Behavior experiments after AAV injection were performed blinded.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Ma	terials & experimental systems	Methods	
n/a	Involved in the study	n/a	Involved in the study
	Antibodies	\boxtimes	ChIP-seq
\boxtimes	Eukaryotic cell lines	\boxtimes	Flow cytometry
\boxtimes	Palaeontology	\boxtimes	MRI-based neuroimaging
	Animals and other organisms		•
\boxtimes	Human research participants		
\boxtimes	Clinical data		

Antibodies

Antibodies used

All antibodies are commercially available:

Antigen/Supplier/Cat#

- 1. H3K27ac/ Abcam/ ab4729
- 2 H3K4me1/ Active motif/ 39297
- 3. Histone H3/ Abcam/ ab1791
- 4. Rabbit IgG/ Diagenode/ C15410206
- 5. Neuron-specific nuclear protein NeuN/ Abcam/ ab104225
- 6. AlexaFluor 488 anti-mouse/ Abcam / ab150077

7. tyrosine hydroxylase/ Millipore/ MAB318

- 8. GFP/Life Technology/ A11122
- 9. Alexa Fluor 488 anti-rabbit/ Thermo Fisher Scientific, Invitrogen/ A11008
- 10. Alexa Fluor 568 anti-mouse/ Thermo Fisher Scientific, Invitrogen/ A11004

Validation

Validation for the antibodies are presented in relevant citations below or can be found on the manufacturer's website.

- 1. Nat Commun. 2018 Jan 11;9(1):161. doi: 10.1038/s41467-017-02536-7.
- 2. Cell. 2017 Oct 19;171(3):557-572.e24. doi: 10.1016/j.cell.2017.09.043.
- 3. Nat Commun. 2018 Jan 4;9(1):53. doi: 10.1038/s41467-017-02242-4.
- 4. Elife. 2018 Oct 4;7. pii: e33337. doi: 10.7554/eLife.33337.
- 5. J Neurosci. 2014 Oct 8;34(41):13586-99. doi: 10.1523/JNEUROSCI.5069-13.2014.
- 6. Sci Rep. 2017 Jan 25;7:41223. doi: 10.1038/srep41223.
- 7. J Biol Chem. 2003 Mar 14;278(11):9928-37. Epub 2003 Jan 2.

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

C57BL/6J and Mus musculus castaneus (Cast/Ei) mouse strains were used in this study. Details concerning genotype, sex, age and cross-breeding are described in the methods section or within individual experiments. For Kcnk9 knockout mouse generation, please see link: J Neurosci. 2009 Feb 25;29(8):2528-33. doi: 10.1523/JNEUROSCI.5764-08.2009. Heterozygous maternal Kcnk9 KO mice, homozygous Kcnk9 mice and WT were used in this study.

Wild animals

This study does not involve wild animals.

Field-collected samples

This study does not involve samples collected from the field.

Ethics oversight

All experimental procedures were performed in accordance with institutional animal welfare guidelines and were approved by the ethical committee of the state government of Rhineland-Palatinate, Germany.

Note that full information on the approval of the study protocol must also be provided in the manuscript.