

Supplementary Materials

Table S1: Data collection and refinement statistics of reported crystal structures.

Complex	ERK3	CLK1-CAF052
PDB Accession Code	7aqb	7ak3
Beamline	SLS X10SA	Diamond I04
	Data Collection	
Resolution ^a (Å)	50.00-2.25 (2.32-2.25)	47.96-2.50 (2.60-2.50)
Spacegroup	<i>P</i> 4 ₃ 2 ₁ 2	<i>C</i> 2
Cell dimensions	<i>a</i> = 83.1, <i>b</i> = 83.1, <i>c</i> = 181.2 Å	<i>a</i> = 91.0, <i>b</i> = 64.3, <i>c</i> = 78.5 Å
	$\alpha = \beta = \gamma = 90.0^\circ$	$\alpha = \gamma = 90.0^\circ; \beta = 119.3^\circ$
No. unique reflections ^a	31,101 (2,799)	13,502 (1,527)
Completeness ^a (%)	100.0 (100.0)	98.0 (98.1)
<i>I</i> / σ <i>I</i> ^a	10.1 (2.1)	7.5 (1.8)
<i>R</i> _{merge} ^a	0.156 (2.484)	0.061 (0.468)
<i>CC</i> 1/2 ^a	0.857 (0.562)	0.995 (0.880)
Redundancy ^a	10.7 (10.2)	2.3 (2.3)
	Refinement	
No. atoms in refinement (P/L/O) ^b	4,137/-/86	2,688/30/19
B factor (P/L/O) ^b (Å ²)	57/-/51	81/55/69
<i>R</i> _{fact} (%)	22.2	21.08
<i>R</i> _{free} (%)	25.5	25.99
rms deviation bond ^c (Å)	0.008	0.004
rms deviation angle ^c (°)	1.5	1.3
	Molprobrity Ramachandran	
Favour (%)	97.83	94.33
Outlier (%)	0	0.30

^a values in brackets are for high resolution shell. ^b P/L/O indicates protein/ligand in the binding pocket/others (solvent and water molecules). ^c rms indicates root-mean-square.

Table S2. Compounds with highest measured ΔT_m values in the thermal stability based assay against ERK3. The ΔT_m values and standard deviation for follow up are the average of two repeats.

Compound Name	ΔT_m (°C) Initial Screen	ΔT_m (°C) Follow Up	SMILES
CAF078	3.83	3.3 ± 0.2	<chem>CN1CCN(c2ccc(Nc3nccc(-c4cnn5ncccc45)n3)cc2C#N)CC1</chem>
AT9283	3.08	3.1 ± 0.1	<chem>C1CC1NC(Nc1c[nH]nc1c1nc2cc(CN3CCOCC3)ccc2[nH]1)=O</chem>
GW779439X	2.84	2.3 ± 0.3	<chem>CN1CCN(CC1)c1ccc(cc1C(F)(F)F)Nc1nccc(c2cnn3c2ccc3)n1</chem>
CAF052	2.81	2.5 ± 0.3	<chem>CN1CCN(c2ccc(Nc3nccc(-c4cnn5ncccc45)n3)cc2F)CC1</chem>
GW814408X	2.55	2.5 ± 0.2	<chem>COc1cccc(c1)-c1c[nH]c2c(N\N=C\c3ccncc3)ncnc12</chem>
Milcilib	2.11	2.5 ± 0.3	<chem>CNC(=O)c1nn(C)c2c1C(C)(C)Cc1nc(Nc3ccc(N4CCN(C)CC4)cc3)nc1-2</chem>
Canertinib	2.05	3.1 ± 0.3	<chem>C=CC(=O)Nc1cc2c(Nc3ccc(F)c(Cl)c3)nenc2cc1OCCCN1CCOCC1</chem>
XL147	1.96	1.2 ± 0.6	<chem>CC(C)(C(=O)NC1=CC(=CC=C1)S(=O)(=O)NC2=NC3=CC=CC=C3N=C2NC4=C(C=CC(=C4)OC)Cl)N</chem>
CD532	1.7	1.0 ± 0.1	<chem>C(=O)(NC1=CC=C(C=C1)NC2=NC=CC(=N2)NC3=NNC(=C3)C4CCCC4)NC5=CC=CC(=C5)C(F)(F)F</chem>
IKK-16	1.67	1.7 ± 0.4	<chem>O=C(C1=CC=C(NC2=NC=CC(C3=CC4=CC=CC=C4S3)=N2)C=C1)N5CCCC(N6CCCC6)CC5</chem>
CAF045	1.61	1.4 ± 0.5	<chem>CN1CCN(c2ccc(Nc3nccc(-c4cnn5ncccc45)n3)cc2)CC1</chem>
GSK1070916	1.45	0.1 ± 0.3	<chem>CCn1cc(c2ccnc3c2cc(c2cccc(CN(C)C)c2)[nH]3)c(c2ccc(cc2)NC(N(C)C)=O)n1</chem>
GW5074	1.36	n/a	<chem>C=C1C(Nc2ccc(cc12)I)=O)c1cc(c(c1)[Br])O[Br]</chem>
GW743024X	1.32	1.2 ± 0.3	<chem>CC(=CC=C(NC(=O)C(C=CO1)=C1)C1)C=1C(C=CC1C(=O)NCC(C2)C2)=CC=1</chem>
GW784307A	1.3	n/a	<chem>[Cl].COC(C=CC=C1N(N=C2)C3=C2C(NN=CC(=CC=C2)C(=O)NCCN(C)C)C=C2)=NC=N3)=C1</chem>
IMD0354	1.21	n/a	<chem>C1=CC(=C(C=C1Cl)C(=O)NC2=CC(=CC(=C2)C(F)(F)F)C(F)(F)F)O</chem>
Sorafenib	1.2	0 ± 0.5	<chem>CNC(=O)c1cc(Oc2ccc(NC(=O)Nc3ccc(Cl)c(C(F)(F)F)c3)cc2)ccn1</chem>
CGK733	1.17	n/a	<chem>O=C(NC(NC(=S)Nc1ccc(F)c([N+](=O)[O-])c1)C(Cl)(Cl)Cl)c1cccc1)c1cccc1</chem>
Butein	1.15	n/a	<chem>C1=CC(=C(C=C1C=CC(=O)C2=C(C=C(C=C2)O)O)O)O</chem>
Quizartinib	1.12	n/a	<chem>O=C(NC1=NOC(C(C)(C)C)=C1)NC2=CC=C(C=C2)C3=CN4C(SC5=CC(OCCN6CCOCC6)=CC=C45)=N3</chem>
GSK1326255A	1.12	-0.3 ± 0.3	<chem>CCCN1CCC(CC1)c1ccc(Nc2nc(Nc3cc(F)ccc3C(N)=O)c3cc[nH]c3n2)c(OC)c1</chem>
SB-250715	1.1	n/a	<chem>FC(=CC=C1C(N=C2)=C(N2C(CCNC2)C2)C(C=CN=C2OC(C=CC(OCO3)=C34)=C4)=N2)C=C1</chem>
Biofocus 093_0089_0063	1.07	0.2 ± 0.2	<chem>CN(C)c1ccc(Nc2cc(-c3cccc3Oc3cccc3)nc3cnn23)cc1</chem>
GW305178X	1.04	1.1 ± 0.2	<chem>NS(=O)(=O)C(C=CC(NN=C(C(=O)N1)C(=C1C=C1)C2C1=NC=CC=2)=C1)=C1</chem>
GW580496A	1.04	0.8 ± 0.4	<chem>[Cl].CS(=O)(=O)CCNCC(=CC=C1C(C=CC(N=CN2)=C3C=2NC(C=CC(OCC(C=CC=C2)=C2)=C2[Br])=C2)=C3)O1</chem>
ALW-II-49-5	1.04	0 ± 0.2	<chem>Cc1ccc(C(=O)Nc2cccc(C(F)(F)F)c2)cc1Nc1ccc2occc(=O)c2c1</chem>
CNX-774	1.04	n/a	<chem>CNC(=O)C1=NC=CC(=C1)OC2=CC=C(C=C2)NC3=NC=C(C(=N3)NC4=CC(=CC=C4)NC(=O)C=C)F</chem>
ChemDiv Y200- 4004	1.03	-0.3 ± 0.3	<chem>Cc1ccc(C(=O)Nc2cccc(C(F)(F)F)c2)cc1Nc1ccc2occc(=O)c2c1</chem>
Biofocus 040_0252_0062	1.02	-0.3 ± 0.3	<chem>Cc1ccc(C(=O)Nc2cccc(C(F)(F)F)c2)cc1Nc1ccc2occc(=O)c2c1</chem>

Biofocus 032_0099_0280	1.01	0.6 ± 0.4	<chem>Cc1ccc(C(=O)Nc2cccc(C(F)(F)F)c2)cc1Nc1ccc2occc(=O)c2c1</chem>
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Abbreviations: ΔT_m : melting temperature shift; n/a: not available.

Table S3. Top hits of cellular NanoBRET screening. Displayed are the average and standard deviation of a single dose measurement of both 10 and 5 μ M. For comparison the ΔT_m values measured in the DSF assay of each inhibitor are shown.

Name of Compound	Norm.BRET 10 μ M (%)	Norm.BRET 5 μ M (%)	ΔT_m ERK3 ($^{\circ}$ C)	SMILES
AT9283	10.6 \pm 1.5	16.1 \pm 3.9	3.1 \pm 0.1	<chem>N(C(=O)NC1C(=NNC=1)C2NC3C(N=2)=CC(=CC=3)CN4CCOCC4)C5CC5</chem>
JNJ-7706621	17 \pm 0.1	23.1 \pm 0.1	0.2 \pm 0.1	<chem>C1(=CC=C(C=C1)NC2N=C(N(N=2)C(C3=C(C=CC=C3F)F)=O)N)S(N)(=O)=O</chem>
IKK-16 (IKK Inhibitor VII)	19.6 \pm 5.7	28.8 \pm 2.4	1.7 \pm 0.4	<chem>C1(C=CC(=CC=1)NC2N=C(C=CN=2)C3=CC4C(S3)=CC=CC=4)C(=O)N5CCC(CC5)N6CCCC6</chem>
Milciclib (PHA- 848125)	26.8 \pm 0.5	40.9 \pm 0	2.5 \pm 0.3	<chem>C12CC(C3=C(C(=NC(=NC=1)NC4=CC=C(C=C4)N5CCN(CC5)C)2)N(N=C3C(NC)=O)C)(C)C</chem>
TWS119	28.1 \pm 5.5	54.2 \pm 6.4	0.6 \pm 0.2	<chem>C1=CC=C(C=C1O)OC2=NC=NC3=C2C=C(N3)C4=CC=CC(=C4)N</chem>
GSK1070916	31.3 \pm 0.1	56.8 \pm 2.8	0.1 \pm 0.3	<chem>C1=CN=C2C(=C1C3=CN(N=C3C4=CC=C(C=C4)NC(N(C)C)=O)CC)C=C(N2)C5=CC=CC(=C5)CN(C)C</chem>
WHI-P154	36.2 \pm 6.3	50.8 \pm 4.5	-0.1 \pm 0.2	<chem>N1C=NC(=C2C=C(C(=CC=12)OC)OC)NC3C=C(C(=CC=3)O)Br</chem>
TG101209	36.9 \pm 0.7	46.6 \pm 2.2	0.3 \pm 0.2	<chem>C1=C(C=C(C=C1)NC2=NC(=NC=C2)NC3=CC=C(C=C3)N4CCN(CC4)C)S(NC(C)(C)C)(=O)=O</chem>
AEE788 (NVP- AEE788)	38.8 \pm 3.1	40 \pm 1.6	0.4 \pm 0.3	<chem>C1=NC(=C2C(=N1)NC(=C2)C3=CC=C(C=C3)CN4CCN(CC4)CC)N[C@@H](C5=CC=CC=C5)C</chem>
AG-1478 (Tyrphostin AG- 1478)	41.7	73.5 \pm 1.1	0.06	<chem>C1=C2C(=CC(=C1OC)OC)C(=NC=N2)NC3=CC(=CC=C3)Cl</chem>
IPA-3	45.5 \pm 4.2	67.8 \pm 10.6	-3.92 \pm 0.3	<chem>S(SC1C(=CC=C2C=CC=CC=12)O)C3C(=CC=C4C=CC=CC=34)O</chem>
BI-D1870	49.4 \pm 1.1	80.6 \pm 6.7	0.02	<chem>C1(=C(C=C(C=C1F)NC2=NC3=C(C=N2)N(C(C(N3CCC(C)C)C)=O)C)F)O</chem>
PF-00562271	49.5 \pm 4.6	76.8 \pm 9.4	-0.09	<chem>C1(=NC(=C(C=C1)CNC2=C(C=NC(=N2)NC3=CC=C4C(=C3)CC(N4)=O)C(F)(F)F)N(C)S(=O)(C)=O).C5=CC=CC(=C5)S(O)(=O)=O</chem>
SNS-032 (BMS- 387032)	49.8 \pm 4.1	71.7 \pm 7.8	0.15	<chem>C1(=CN=C(O1)CSC2=CN=C(S2)NC(=O)C3CCNCC3)C(C)(C)C</chem>
CYC116	52.9 \pm 2.8	64.9 \pm 3.8	0.26	<chem>C1(=NC=CC(=N1)C2SC(=NC=2C)N)NC3=CC=C(C=C3)N4CCOCC4</chem>
BX-912	53.2 \pm 1.7	74.1 \pm 1.5	-0.12	<chem>N1(CCCC1)C(=O)NC2=C(C=CC=2)NC3N=C(C(=CN=3)Br)NCCC4=CN=CN4</chem>
Vandetanib (ZD6474)	56.9 \pm 4.1	69 \pm 0.5	0.2	<chem>C12=C(C(=NC=N1)NC3=C(C=C(C=C3)Br)F)C=C(C(=C2)OCC4CCN(CC4)C)OC</chem>
ENMD-2076	58.5 \pm 0.4	90.8 \pm 3	-0.15	<chem>C1(N=C(C=C(N=1)N2CCN(CC2)NC3=NNC(=C3)C)/C=C/C4C=CC=CC=4</chem>
PRT062607 (P505-15, BIIB057) HCl	60.1 \pm 16.9	88.2 \pm 10.3	0.51	<chem>N1(=C(N=C(C=C1)C(N)=O)NC2=CC(=CC=C2)N3N=CC=N3)N[C@@H]4CCCC[C@H]4N).Cl</chem>
AZD5438	61.7 \pm 2.2	93.3 \pm 4	-0.17	<chem>C1=C(C=CC(=C1)NC2=NC=CC(=N2)C3=CN=C(N3C(C)C)S(=O)(C)=O</chem>

Abbreviations: Norm.BRET: normalised BRET; ΔT_m : melting temperature shift; n/a: not available.

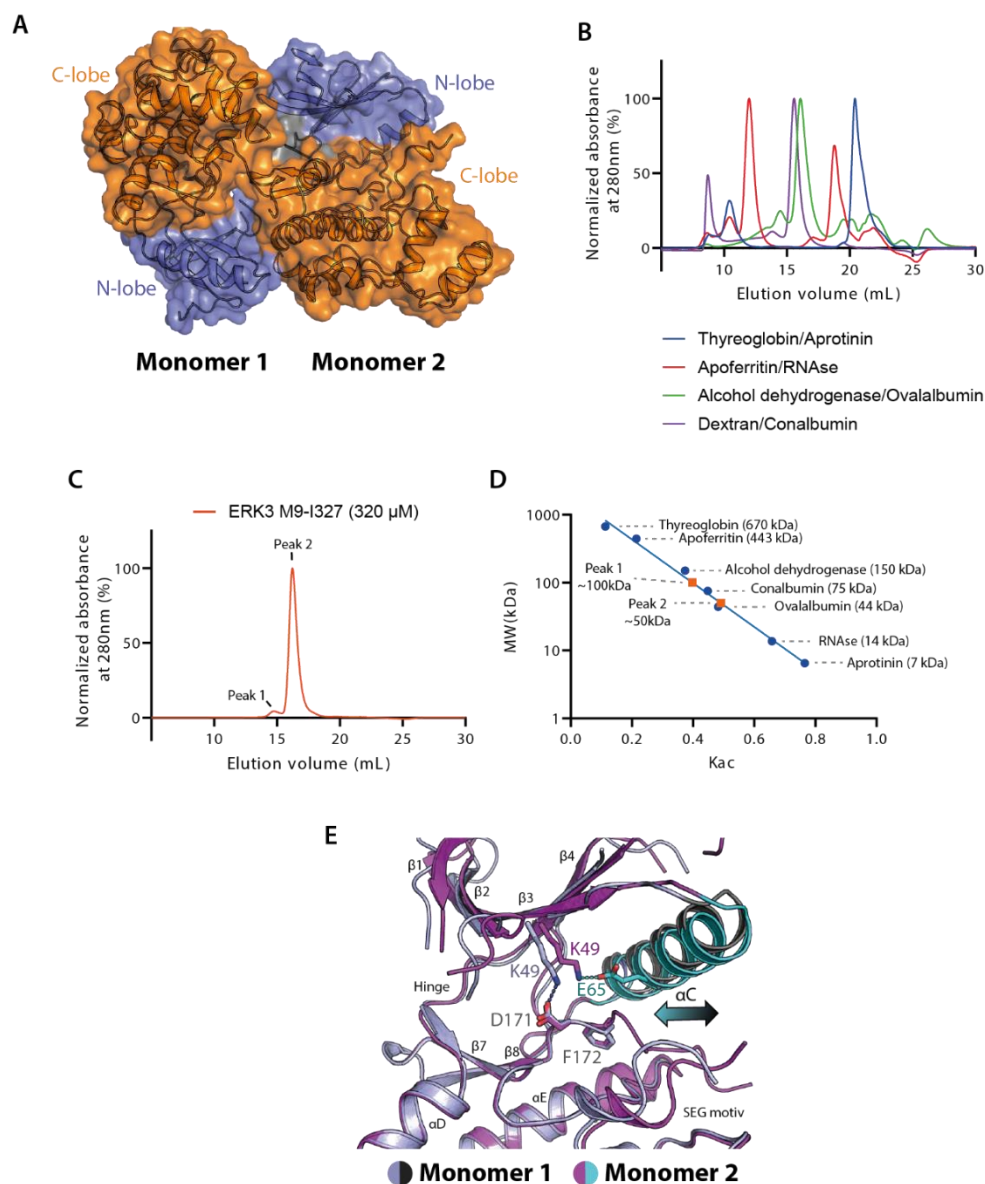


Figure S1. Dimeric ERK3 in the crystal structure. (A) Face-to-face, head-to-tail dimer of ERK3 in the asymmetric unit. The kinase N-terminal and C-terminal lobes are in blue and orange, respectively. (B) Size exclusion chromatograms of protein markers. (C) Size exclusion chromatogram of ERK3 at 320 μM. (D) The standard curve calculated from the elution profiles of protein markers. Kav values of each marker were calculated from the equation: $K_{av} = (V_e - V_o) / (V_t - V_o)$, where V_e = elution volume, V_o = void volume and V_t = column volume, and plots against protein molecular weight in kDa in logarithmic scale. Orange dots in the plots are the Kav values for ERK3 from both elution points with their estimated molecular weight calculated based on the standard curve. (E) Superimposition reveals highly conserved structure of both ERK3 monomers present in the asymmetric unit.

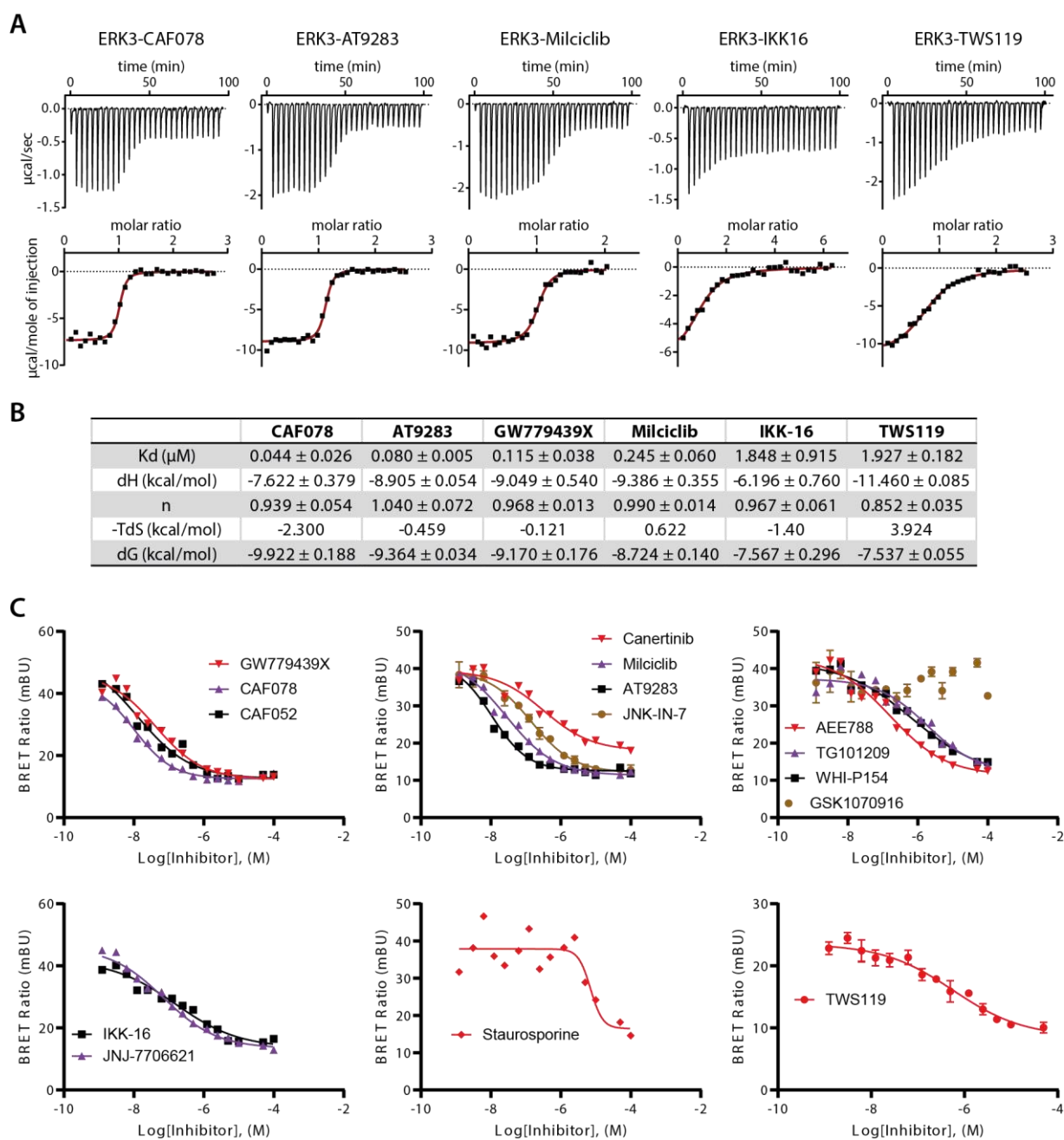


Figure S2. (A) ITC results of ERK3 titrated with various compounds. Shown are the raw heat rate measured over time and the integrated heat used for the binding model fit of one representative titration. (B) Binding affinity and thermodynamics of ERK3-inhibitors complexes. Values displayed are the average and standard deviation of at least two independent experiments (C) NanoBRET dose response curves of inhibitors measured in HEK293T cells. Shown is one representative titration for all analysed inhibitors.

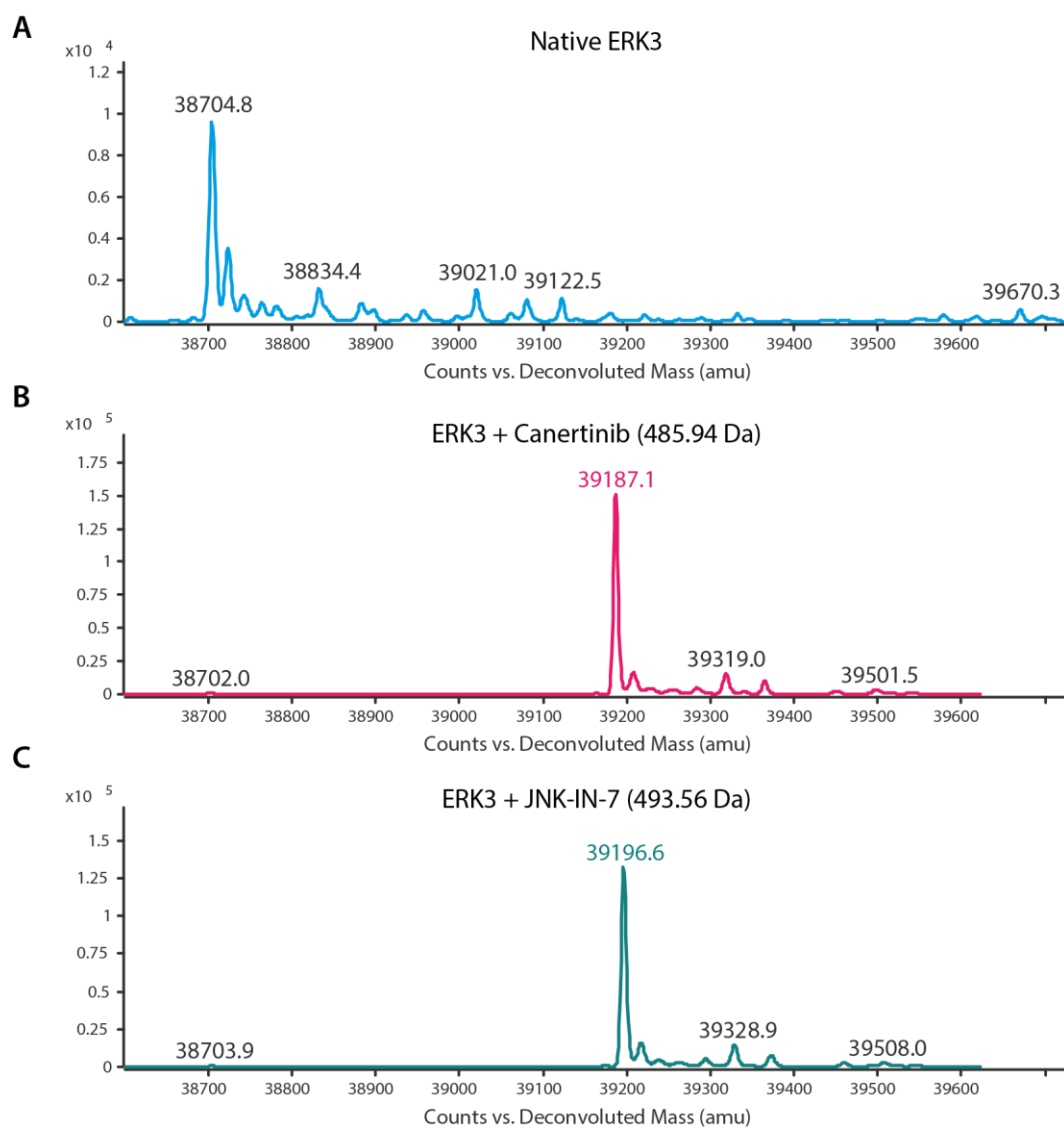


Figure S3. (A–C) Intact mass spectra to test for full covalent adduct formation. ERK3 was either incubated with DMSO (A), two-fold excess of canertinib (B) or JNK-IN-7 (C) for 4 h at room temperature.

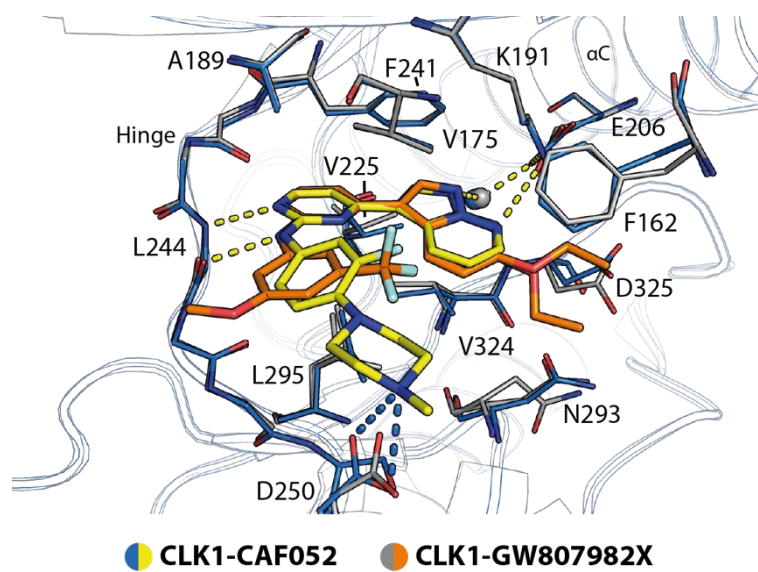


Figure S4. Structural superimposition between the CAF052- (pdb id 7ak3) and GW807982X-CLK1 complexes (pdb id 6zln) revealed highly similar binding modes of both chemically-related inhibitors in the kinase.

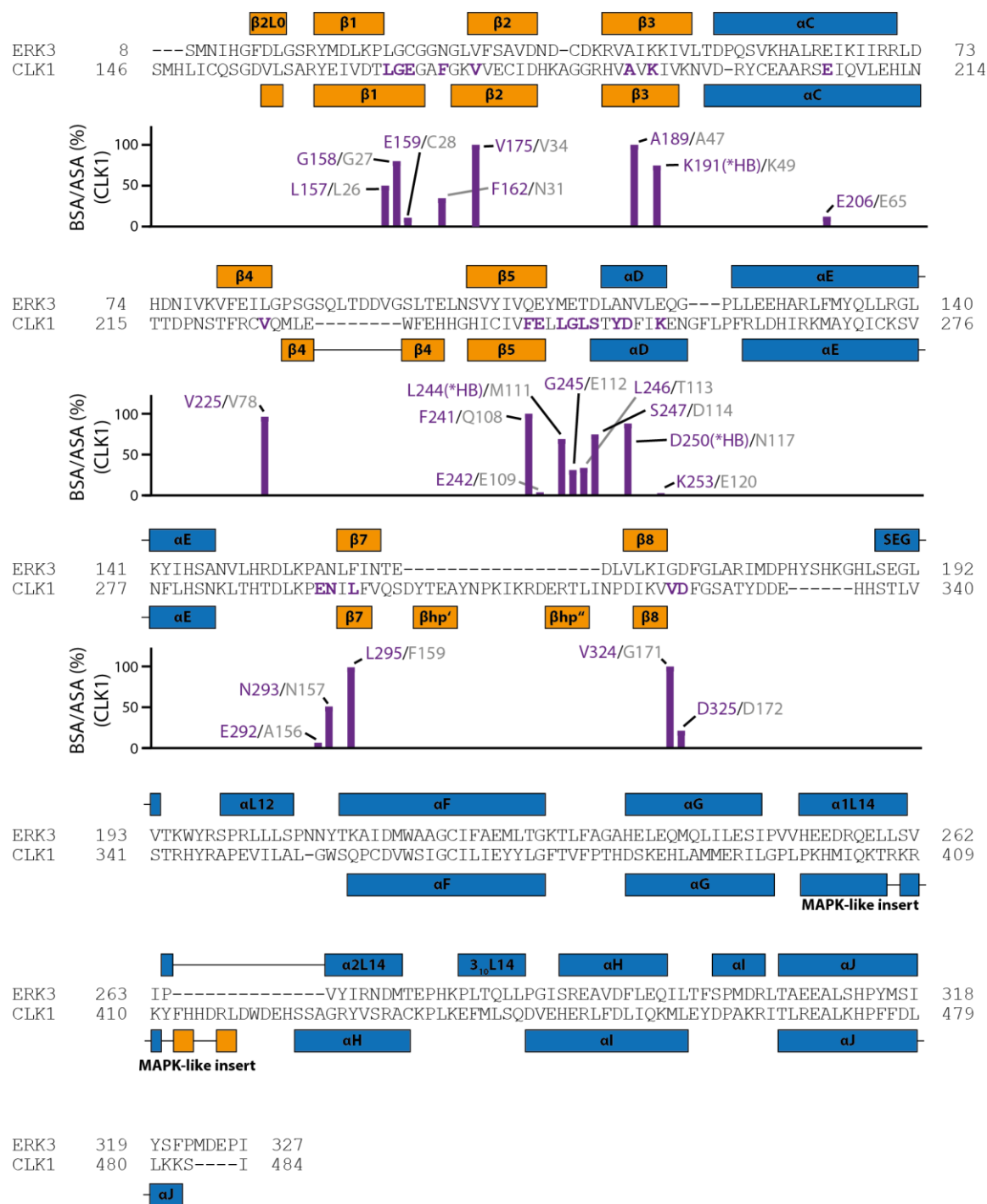


Figure S5: Structure-based sequence alignment of kinase domains of CLK1 and ERK3. Secondary structural elements (α -helices in blue and β -strands in orange) are shown for ERK3 (above) and for CLK1 (below). CLK1 residues interacting with CAF052 are highlighted in magenta. The individual contribution of each residue of CLK1 towards the complex formation with CAF052 expressed by the ratio of buried surface area (BSA)/available surface area (ASA) is shown beneath the alignment (magenta). CLK1 residues forming hydrogen-bonds with CAF052 are highlighted and denoted with “*HB”. The equivalent residues in ERK3 are displayed in grey.