

Supplemental Material

GC content but not nucleosome positioning directly contributes to intron-splicing efficiency in *Paramecium*

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Table of contents

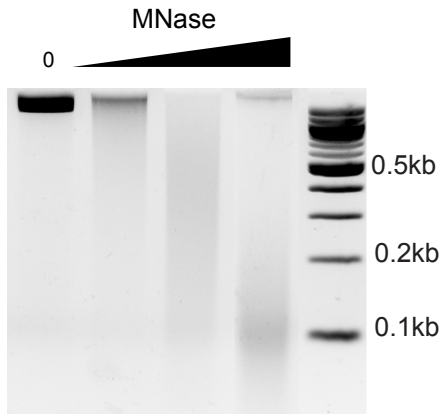
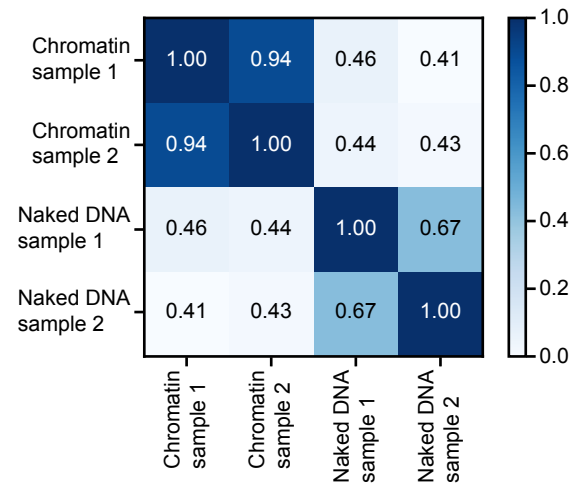
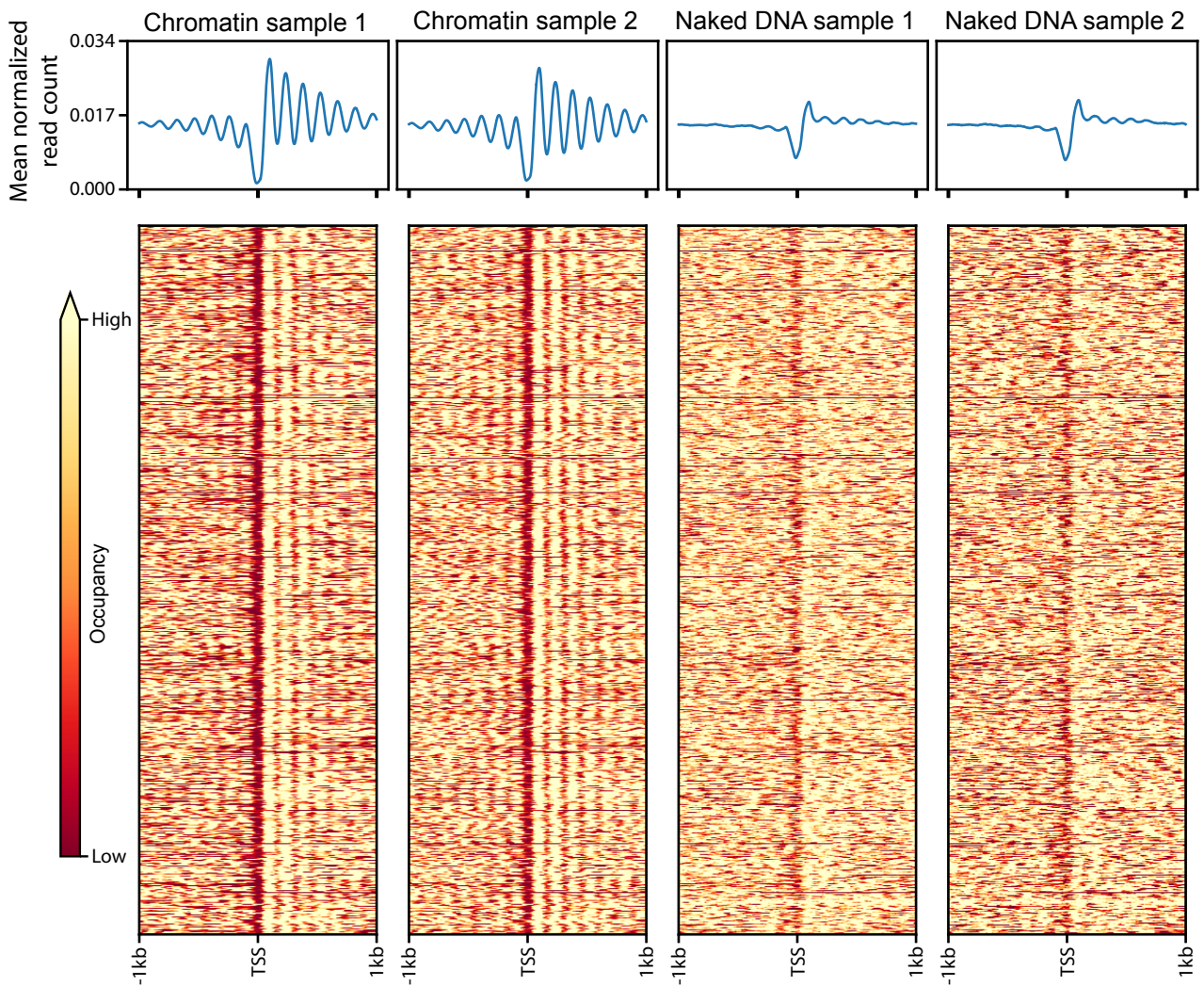
Supplemental Figure S1. Nucleosome occupancy along the MAC *Paramecium* genome.

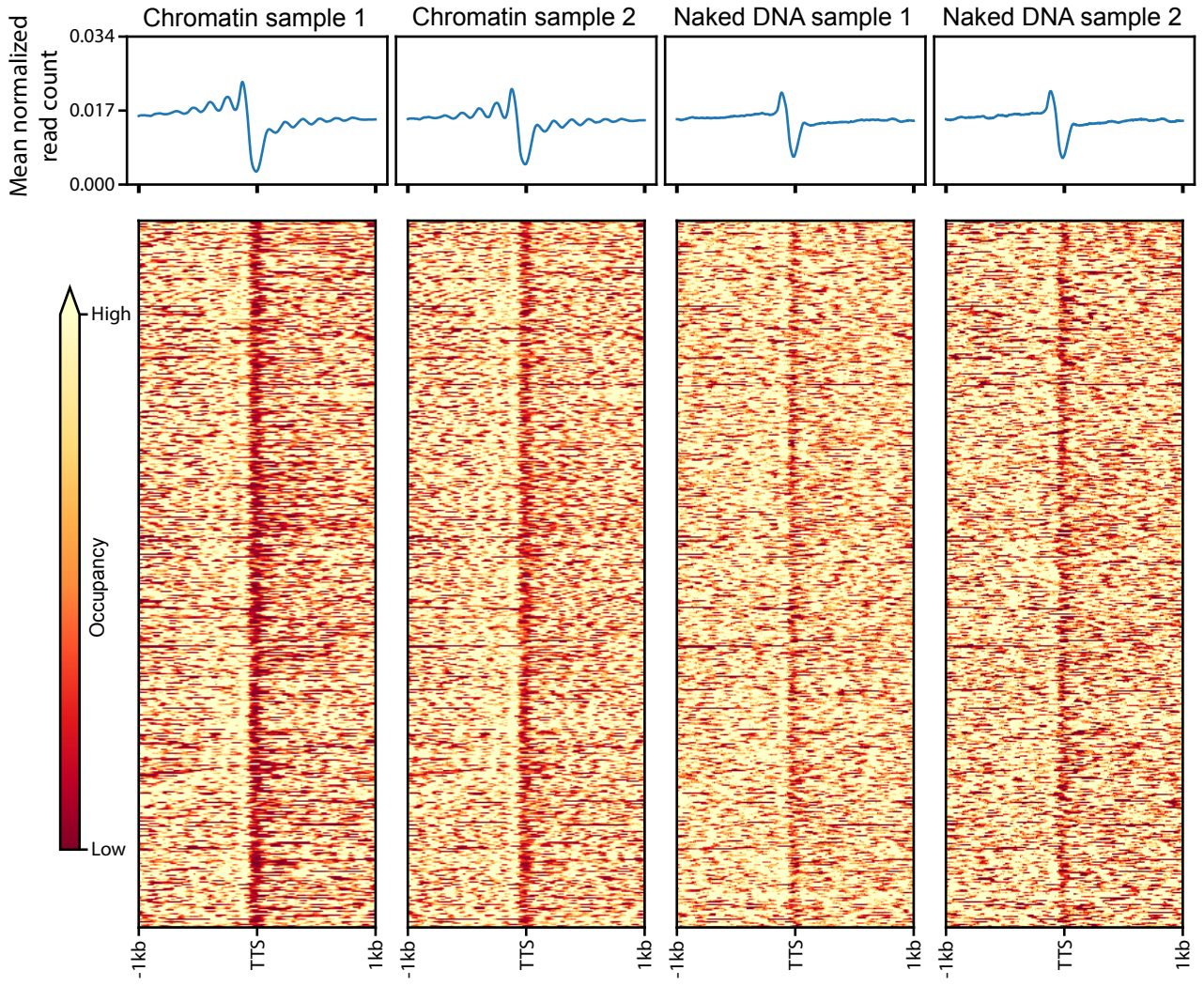
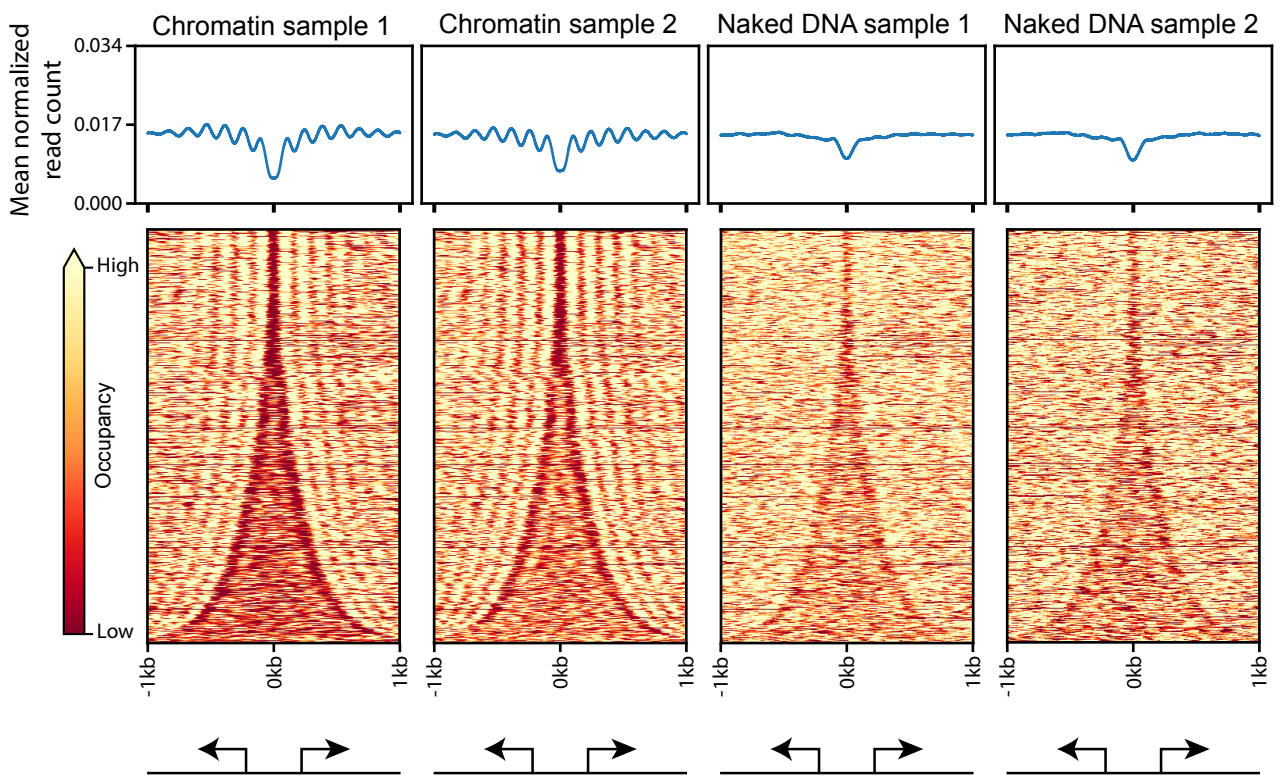
Supplemental Figure S2. Inter-nucleosomal DNA is frequently associated with intron position.

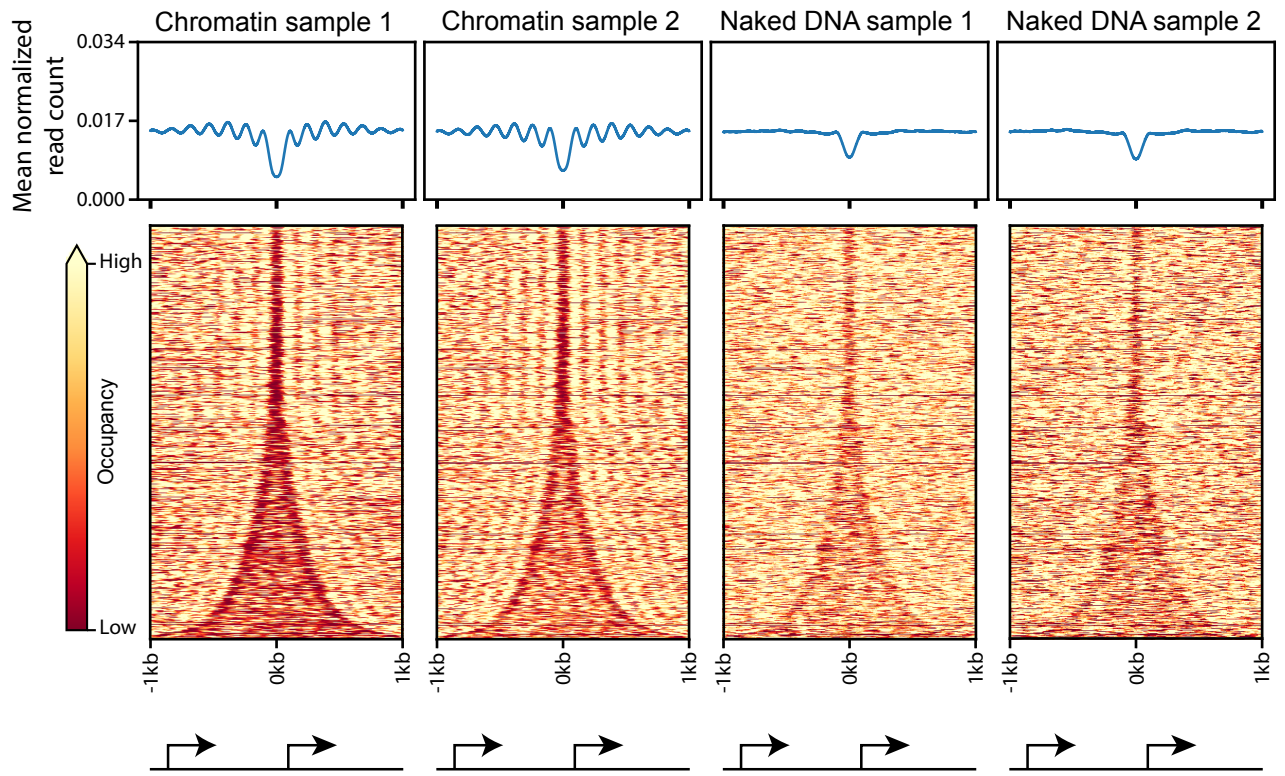
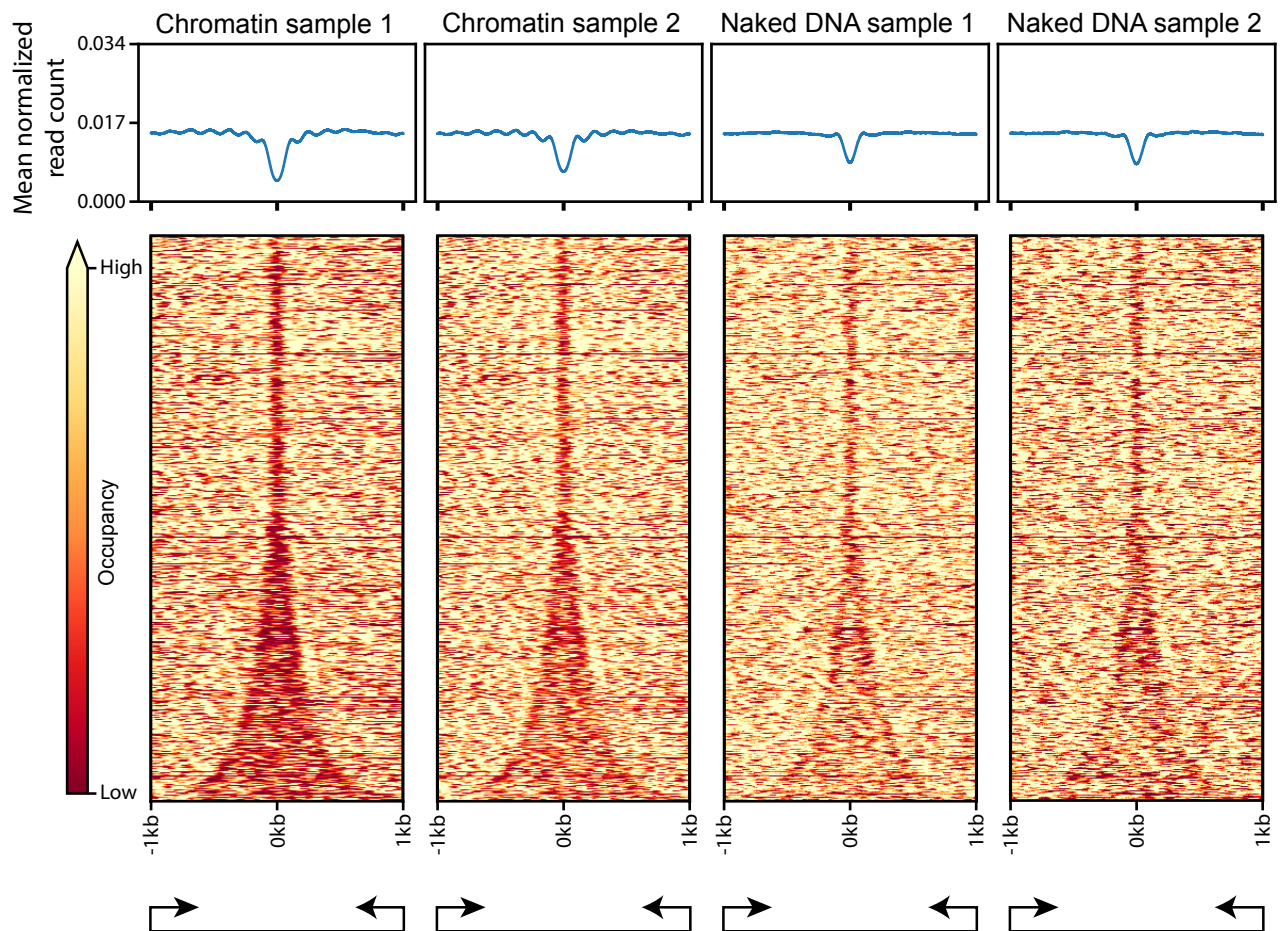
Supplemental Figure S3. Nucleosome positioning is associated with intron-splicing efficiency.

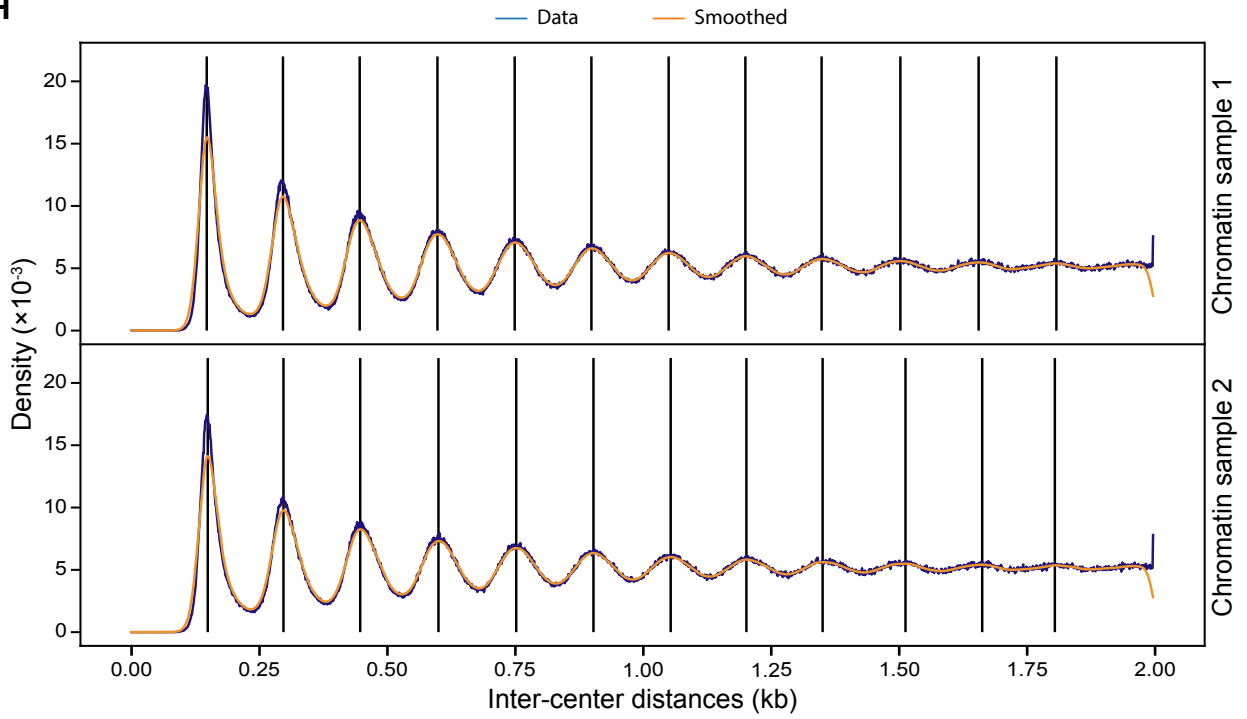
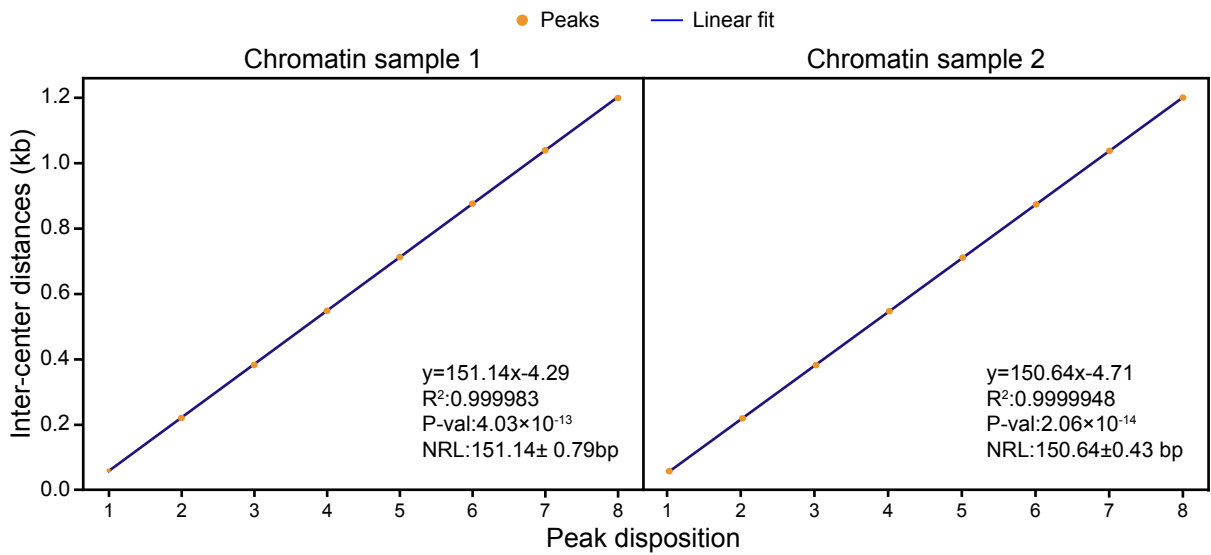
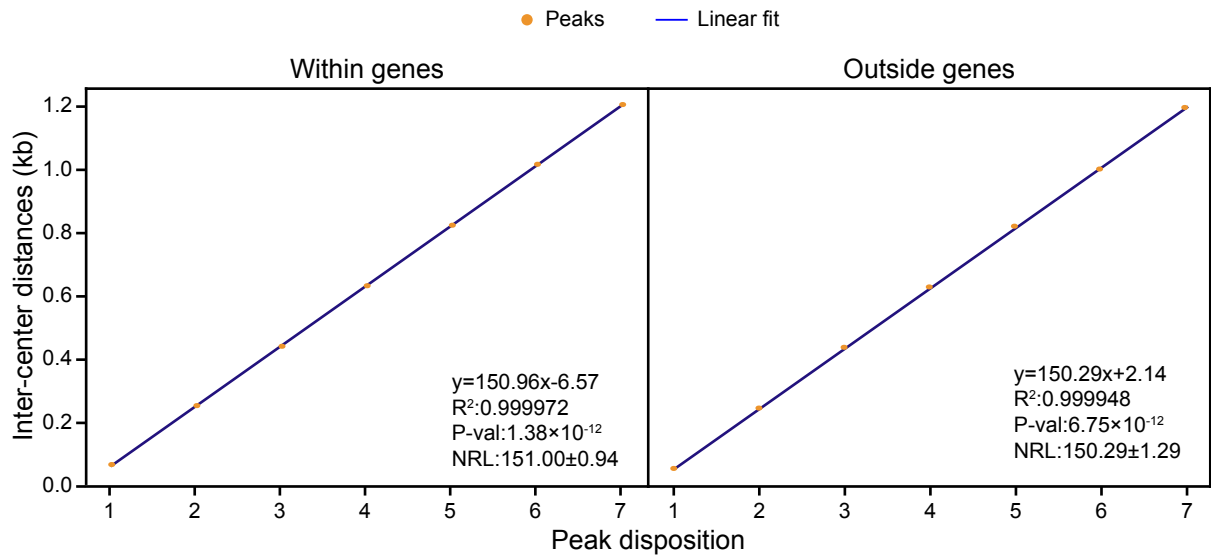
Supplemental Figure S4. GC content related to nucleosome positioning contributes to intron-splicing efficiency.

Supplemental Table S1. Parameters and their contribution in modelling the splicing efficiency in the multilinear regression model.

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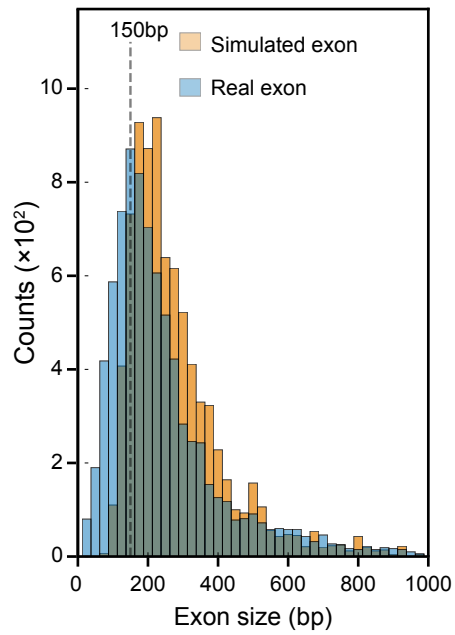
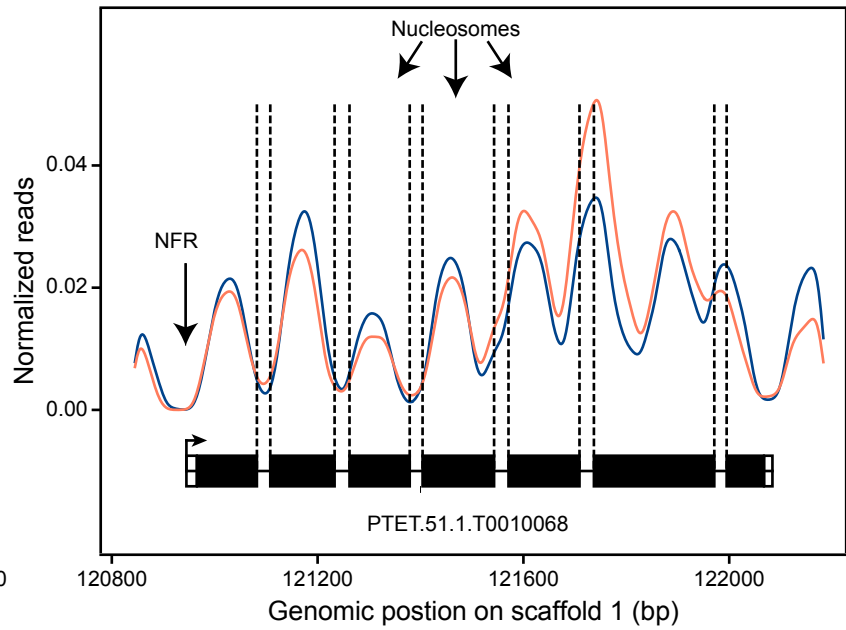
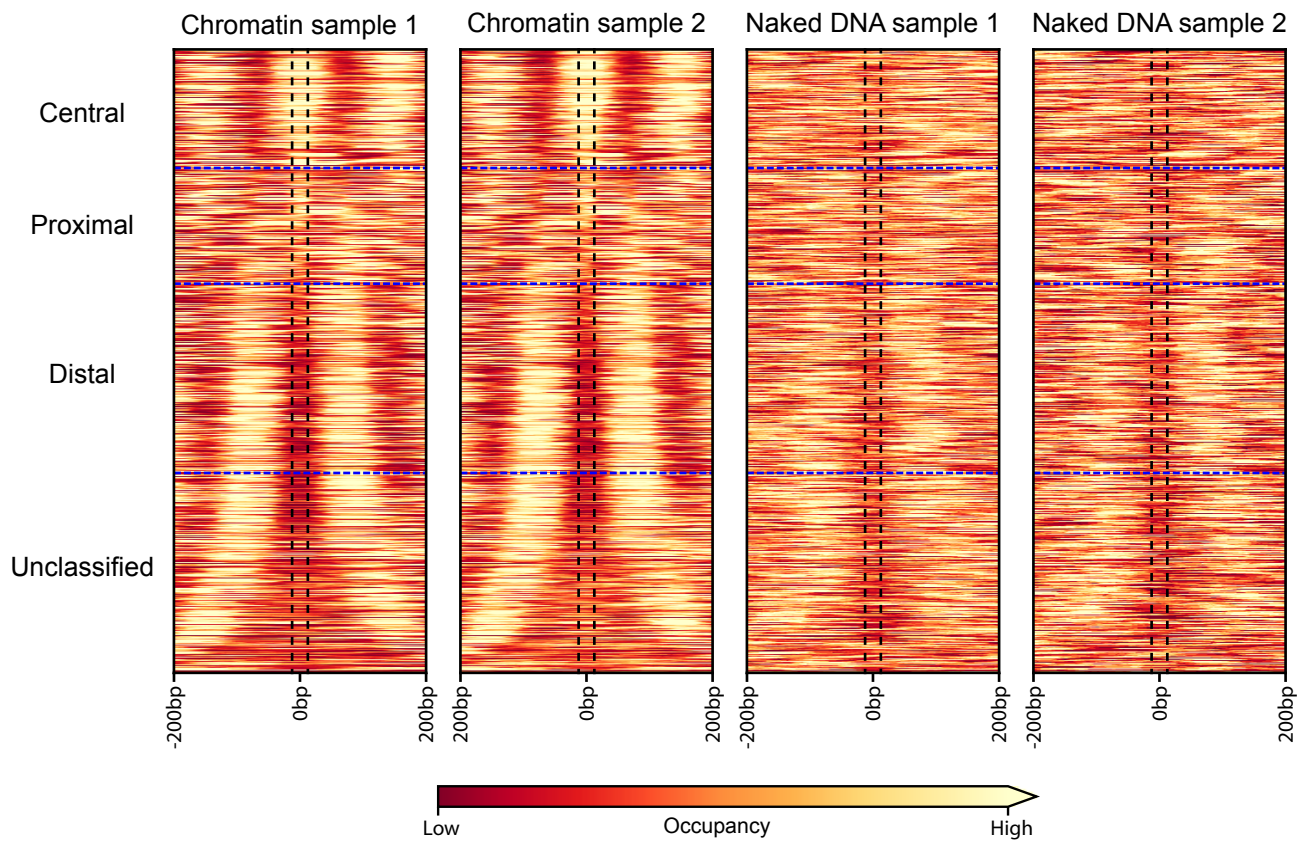
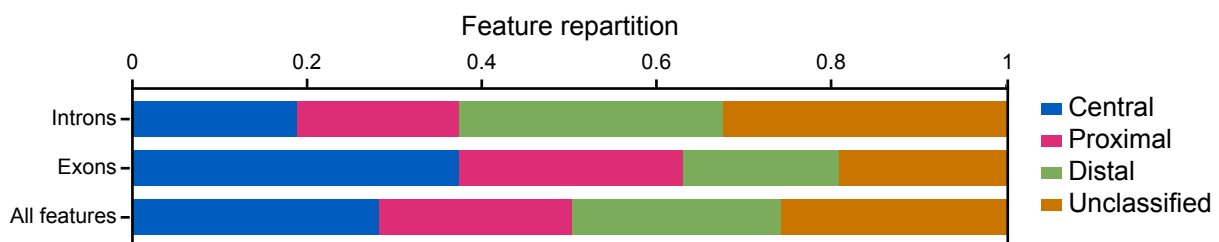
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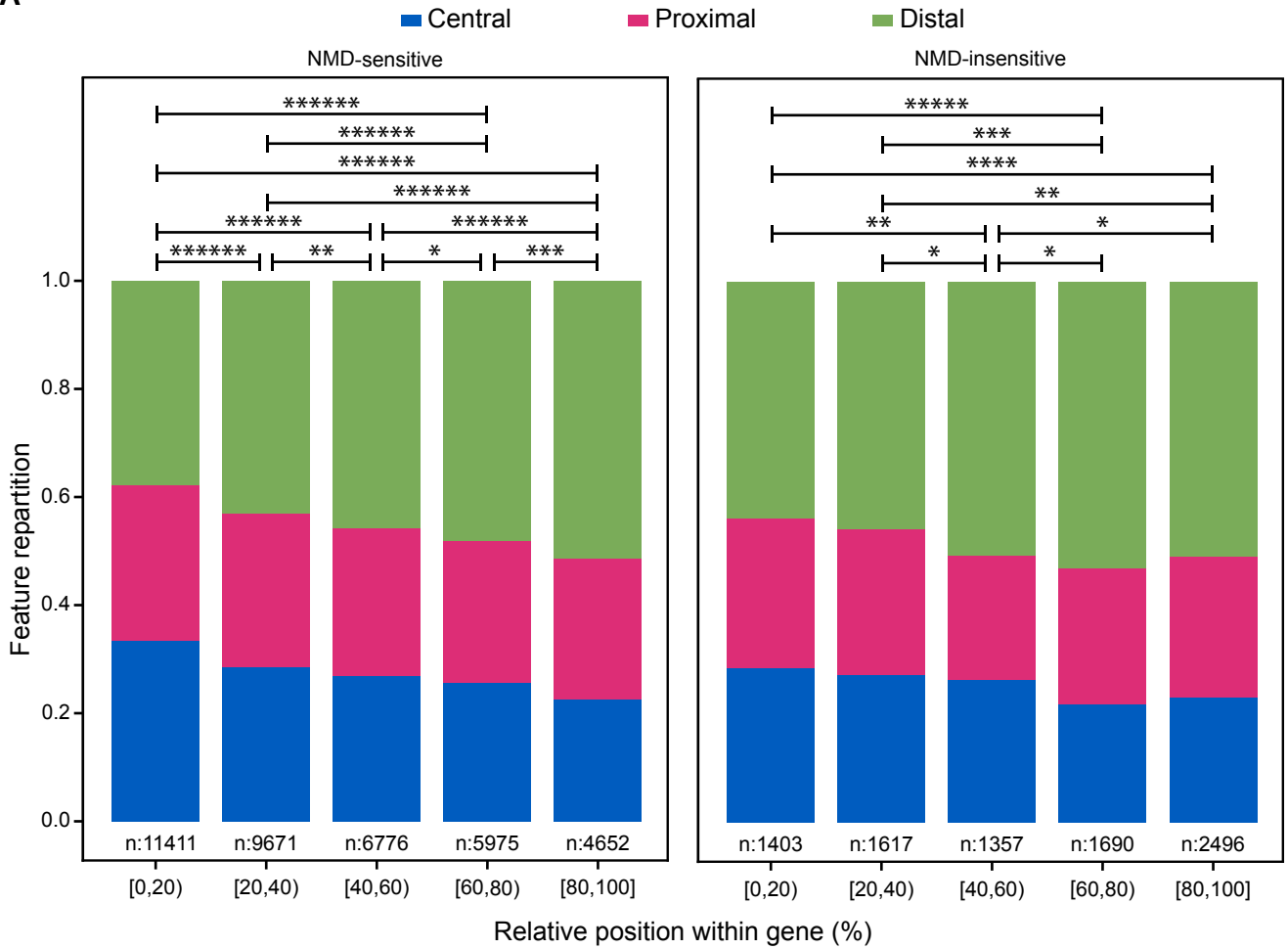
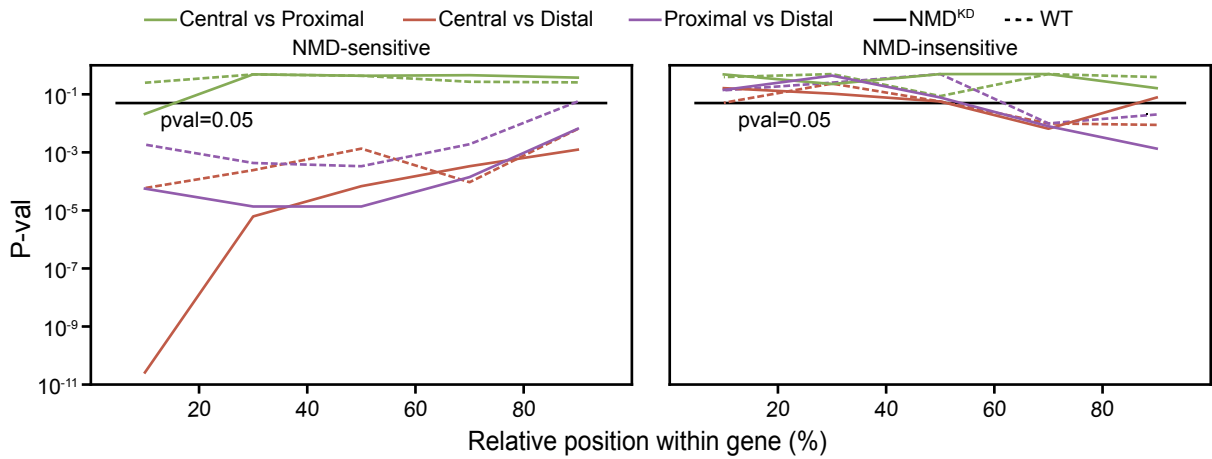
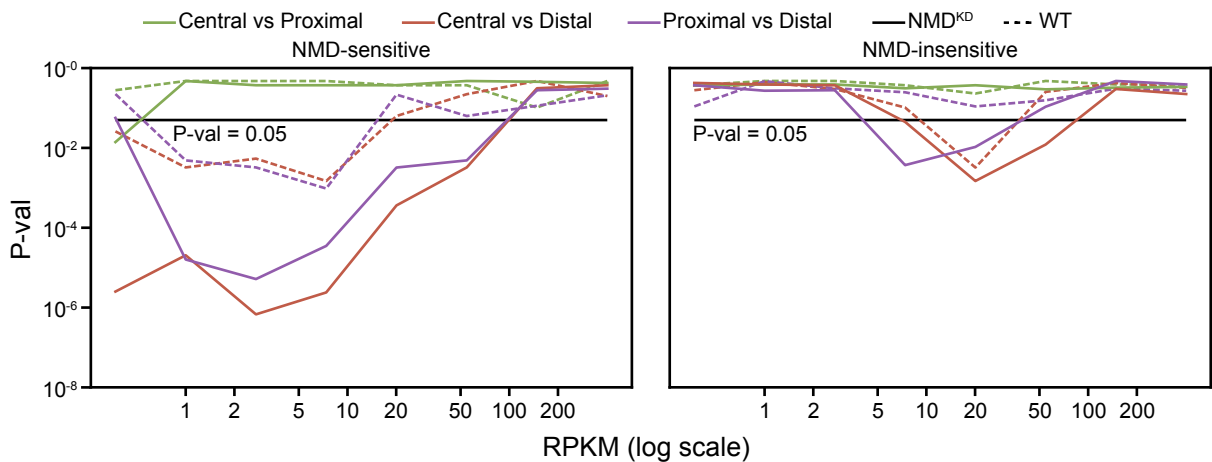
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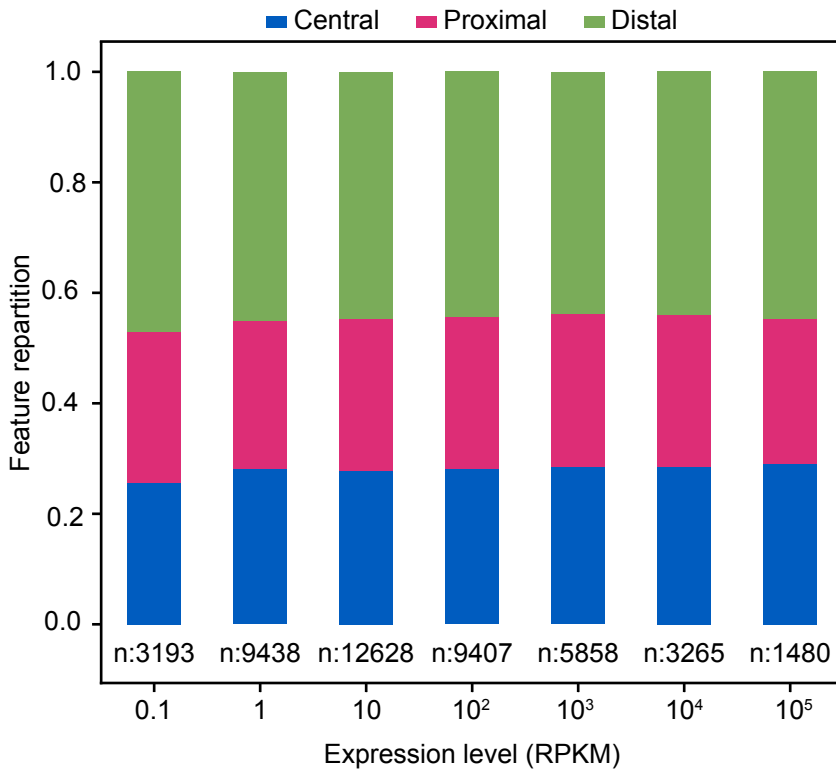
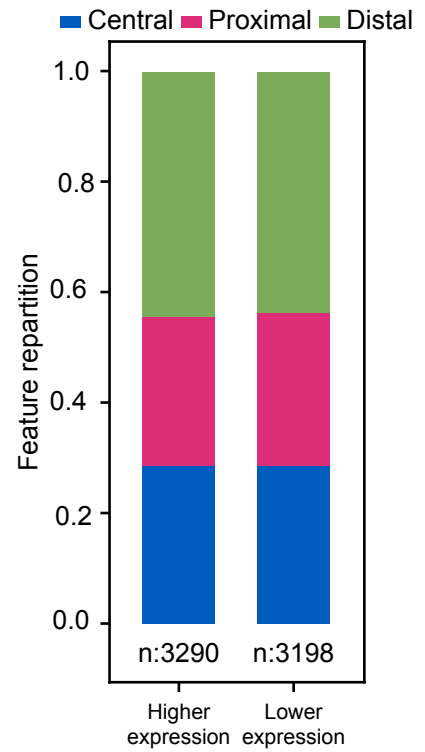
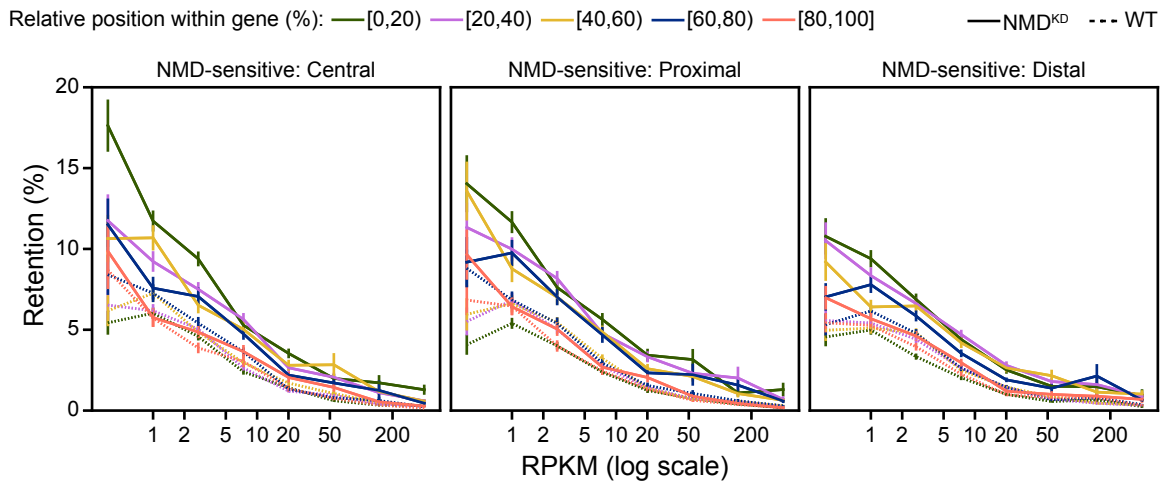
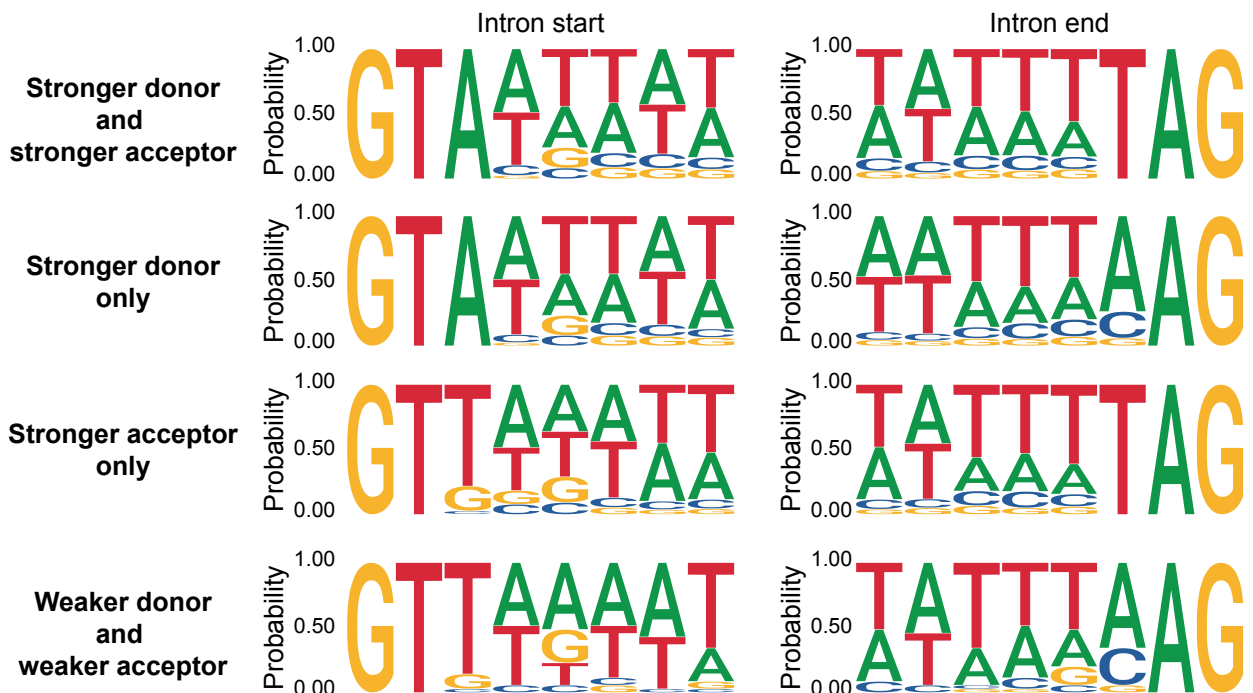
Supplemental Figure S1. Nucleosome occupancy along the MAC *Paramecium* genome.

(A) MNase digestion of naked DNA with increasing MNase enzyme concentration. **(B)** Heatmap showing pairwise Pearson's correlation between chromatin and naked DNA samples (two biological replicates for each). It should be noted that the naked DNA samples show a lower correlation than the chromatin samples. **(C)** Average profiles and heatmaps showing nucleosome occupancy around Transcription Start Sites (TSSs) in individual samples. **(D)** Average profiles and heatmaps showing nucleosome occupancy around Transcription Termination Sites (TTSs) in individual samples. **(E-G)** Average profiles and heatmaps showing nucleosome occupancy ± 1 kb around the center of intergenic regions. Pairs of genes have been divided into three groups based on their relative orientation: divergent (E), tandem (F) and convergent (G). Genes are sorted based on their intergenic distance. **(H)** Inter-center distance between nucleosomes on the same scaffold for the two chromatin samples. In blue, distance distributions from actual data (from 1 bp to 2 kb, binning=1 bp); and in orange, the gaussian smoothed signal. Black dashed lines identify the local maxima (peak centers) of the smoothed data (Methods). **(I)** In orange, the local maxima from Supplemental Fig S1H ordered by increasing distance; in blue, the linear fitted model. On the bottom right of each panel, the information about the linear fitting and the estimated NRL for both chromatin replicates. P-values are calculated using a two-sided Z-test. **(J)** The same analysis as in Fig 1G by separating into the nucleosome centers overlapping with gene bodies (left panel) or outside of them (right panel). As in Fig 1G, local maxima are ordered by increasing distance. The relative linear fitting model is reported in blue. The bottom right gives the information about the linear fitting and the estimated NRL (Mean \pm SD). P-value is calculated using a two-sided Z-test.

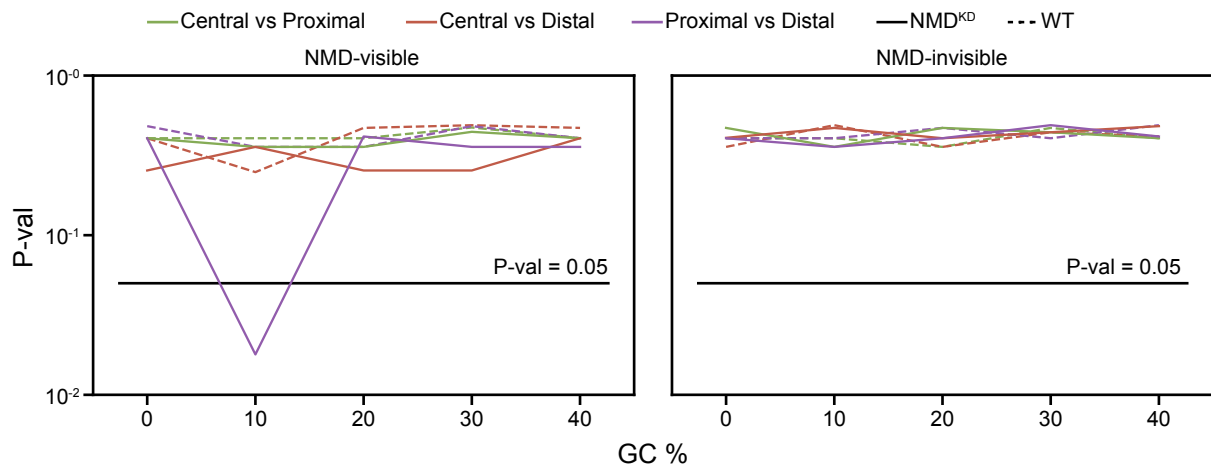
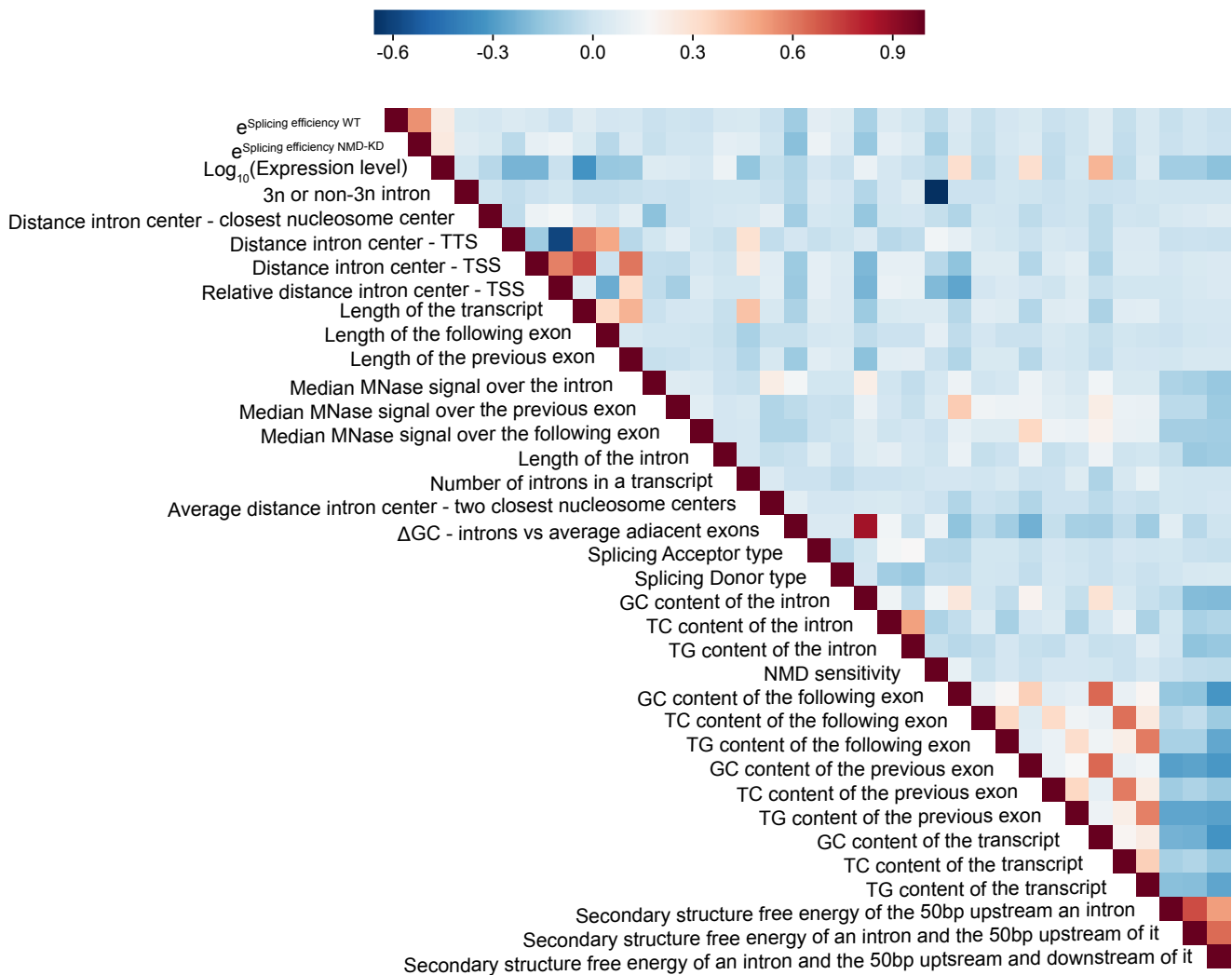
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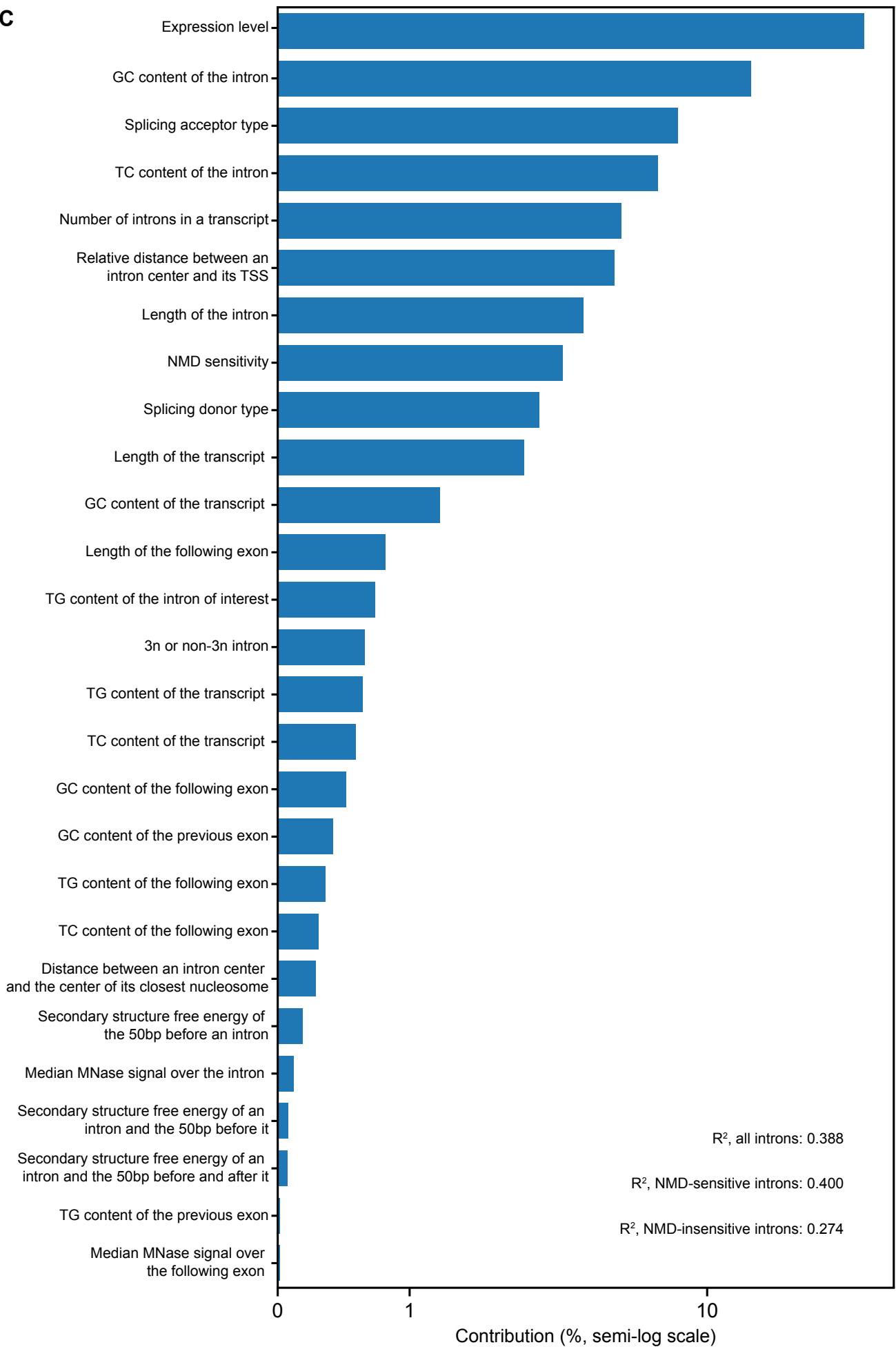
Supplemental Figure S2. Inter-nucleosomal DNA is frequently associated with intron position. **(A)** Histogram showing exon size distribution (bin size = 25 bp) of transcription units whose extremities have been identified by 5' CAP-seq and poly(A) detection: in blue, real exons; and in orange, simulated exons created assuming uniform exon sizes within each transcript. **(B)** The same genomic region as shown in Fig 2B. Tracks of two independent chromatin treated samples reporting nucleosome occupancy over a representative gene. Intron locations are indicated by dashed vertical lines. Nucleosome free regions (NFRs) are found around the gene promoters and introns are frequently associated with inter-nucleosomal DNA. **(C)** Heatmap showing nucleosome occupancy ± 200 bp around intron centers in individual samples. Introns are ordered as in Fig 2D. Horizontal blue dashed lines define the boundaries that identify central, proximal, distal and unclassified introns, while vertical black dashed lines delineate the average size of an intron (25 bp). **(D)** Same figure as Fig 2G including features with distance >75 bp. "Unclassified" corresponds to introns and/or exons that do not belong to the three defined categories (central, proximal, distal).

A**B****C**

D**E****F****G**

Supplemental Figure S3. Nucleosome positioning is associated with intron-splicing efficiency. **(A)** Barplot representation of Fig 3B. P-values are calculated using the χ^2 test and adjusted using false discovery rate (5%). Only the significant ones are indicated. **(B)** P-values relative to Fig 3C were calculated using the Mann–Whitney U test and adjusted using false discovery rate (5%). Comparisons of intron retention rates were evaluated in WT (dashed lines) and NMD-depleted cells (NMD^{KD}, solid line), respectively, for NMD-sensitive introns (left) and NMD-insensitive introns (right). The black line indicates P-value = 0.05. **(C)** P-values relative to Fig 3D were calculated using the Mann–Whitney U test and adjusted using false discovery rate (5%). Color and line style codes are the same as in panel B. **(D)** Introns analyzed in Fig 2G were further divided accordingly to their expression levels. Expression ranges were selected to have maintain a minimal statistical power. **(E)** Intron classification for genes duplicates from the last whole genome duplication. Only duplicates with the same number of identified introns were used. Each pair of genes was assigned to the “higher” or “lower” expression groups based on their expression levels. (D-E) P-values were calculated using a χ^2 test and were adjusted using the false discovery rate (5%) when needed. No significant difference was identified. **(F)** Retention level of NMD-sensitive introns as a function of expression levels in NMD-depleted (NMD^{KD}) and WT cells. Introns are grouped according to their relative position within a gene and to their relative position to the closest nucleosome: central (left), proximal (center) and distal (right). **(G)** Sequence logo plots showing the nucleotide composition of the splicing donors and acceptors sites relative to Fig 3E.

A**B**

C

Supplemental Figure S4. GC content related to nucleosome positioning contributes to intron-splicing efficiency. (A) P-values relative to Fig 4B were calculated using the Mann–Whitney *U* test and adjusted using false discovery rate (5%). Comparisons of intron retention rates were evaluated in WT (dashed lines) and NMD-depleted (NMD^{KD}, solid line) cells, respectively. Left: NMD-sensitive introns. Right: NMD-insensitive introns. The black line indicates the position of P-value = 0.05. **(B)** Pairwise Pearson’s correlation heatmap of all parameters used to model Splicing Efficiency (SE) in NMD-depleted cells. **(C)** Histogram reporting the contribution of the final parameters used in our model. P-values are displayed in Supplemental Table S1. Pearson’s correlation between model and real data for all introns, NMD-sensitive and NMD-insensitive introns are displayed at the bottom right corner.

Supplemental Table S1. Parameters and their contribution in modelling the splicing efficiency in the multilinear regression model.

Group	Parameter	Abbreviation	Contribution	p-value	Group contribution
Expression level	Expression level	EL	46.09%	$< 10^{-16}$	46.09%
Base composition of an Intron	GC content of the intron of interest	GC _I	15.30%	$< 10^{-16}$	22.23%
	TC content of the intron of interest	TC _I	6.20%	$< 10^{-16}$	
	TG content of the intron of interest	TG _I	0.74%	4.95×10^{-7}	
Splicing signals	Splicing acceptor type*	SA	7.56%	$< 10^{-16}$	9.55%
	Splicing donor type*	SD	1.99%	$< 10^{-16}$	
Transcript size and base composition	Number of introns in a transcript	N	4.37%	$< 10^{-16}$	8.71%
	Length of the transcript	L _T	1.87%	$< 10^{-16}$	
	GC content of the transcript	GC _T	1.23%	9.73×10^{-6}	
	TG content of the transcript	TG _T	0.64%	9.45×10^{-8}	
	TC content of the transcript	TC _T	0.59%	9.71×10^{-10}	
Intron size and position	Relative distance between an intron center and its TSS	D _{TSS}	4.06%	$< 10^{-16}$	7.08%
	Length of the intron	L _I	3.02%	$< 10^{-16}$	
NMD sensitivity	NMD sensitivity*	NMD	2.46%	9.60×10^{-13}	2.46%
Base composition and size of the flanking exons	Length of the following exon	L _{FE}	0.82%	2.53×10^{-7}	2.45%
	GC content of the following exon	GC _{FE}	0.52%	4.00×10^{-6}	
	GC content of the previous exon	GC _{PE}	0.42%	1.20×10^{-2}	
	TG content of the following exon	TG _{FE}	0.36%	9.73×10^{-6}	
	TC content of the following exon	TC _{FE}	0.31%	1.66×10^{-4}	
	TG content of the previous exon	TG _{PE}	0.02%	1.45×10^{-6}	
3n or non-3n introns	3n or non-3n introns*	3N	0.66%	8.84×10^{-4}	0.66%

Nucleosome position	Distance between an intron center and the center of its closest nucleosome	D_N	0.29%	2.69×10^{-3}	0.43%
	Median MNase signal over the intron	M_I	0.12%	1.18×10^{-2}	
	Median MNase signal over the following exon	M_{FE}	0.02%	2.72×10^{-2}	
Secondary structure	Secondary structure free energy of the 50 nt upstream of an intron	ΔG_{p50}	0.19%	1.66×10^{-4}	0.35%
	Secondary structure free energy of an intron and the 50 nt upstream	ΔG_{p50-I}	0.08%	4.22×10^{-3}	
	Secondary structure free energy of an intron and the 50 nt upstream and downstream	ΔG_{fp50-I}	0.08%	5.57×10^{-5}	
	Average distance between intron center and the center of the two closest nucleosomes [§]		NA	0.21	
	DeltaGC (Intron-GC content - average flanking-exons-GC contents) ^{&}		NA	NA	
	Distance between an intron center and its TSS ^{&}		NA	NA	
	Distance between an intron center and its TTS ^{&}		NA	NA	
	Length of the previous exon [§]		NA	0.29	
	Median MNase signal over the previous exon [§]		NA	0.30	
	TC content of the previous exon [§]		NA	0.45	

* Binary variables

& Parameter that was excluded after trying to lower the variance inflation factor below 5

§ Not statistically significant parameter