

# One like all? Behavioral response range of native and invasive amphipods to neonicotinoid exposure

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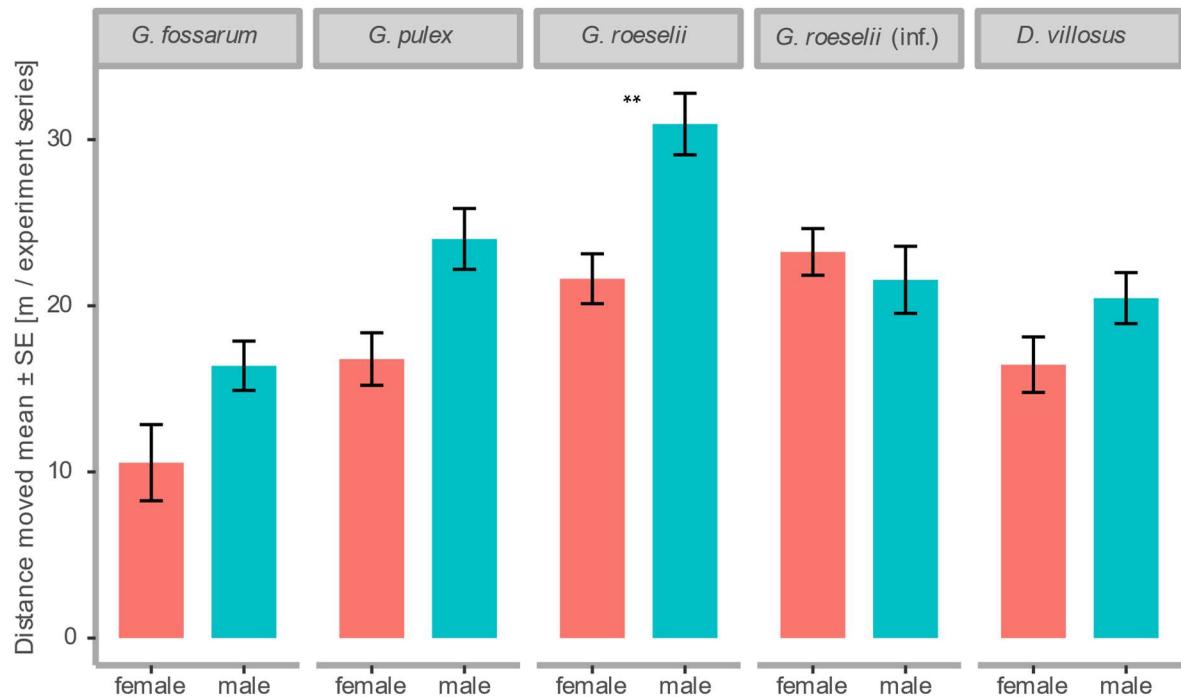
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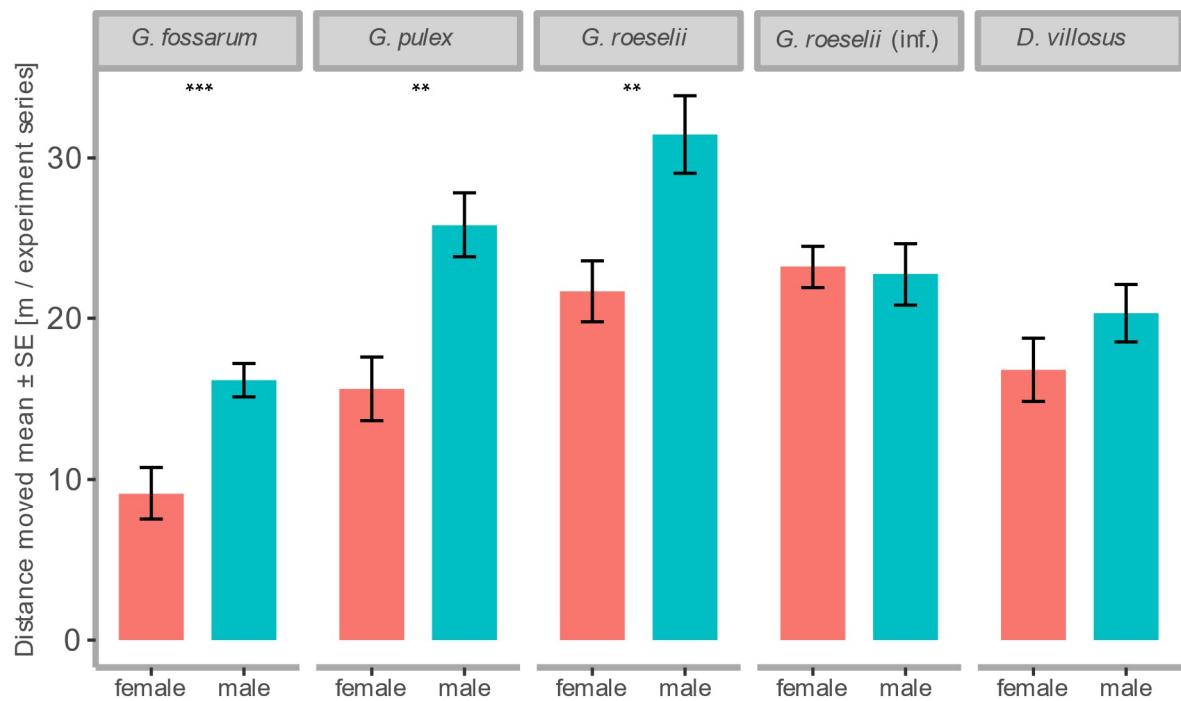
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**Keywords:** Acanthocephalan parasites; *Dikerogammarus villosus*; Freshwater invertebrates; Locomotion; Organic micropollutants; Sublethal effects

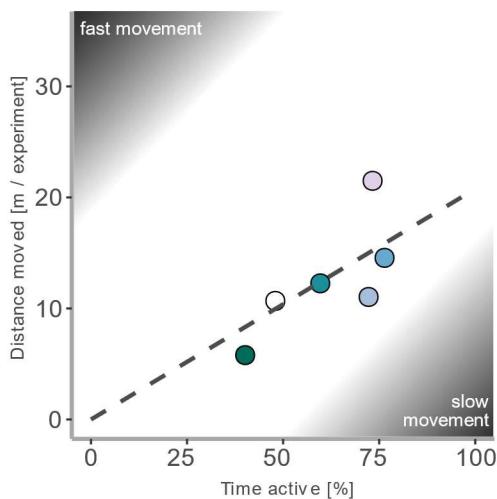
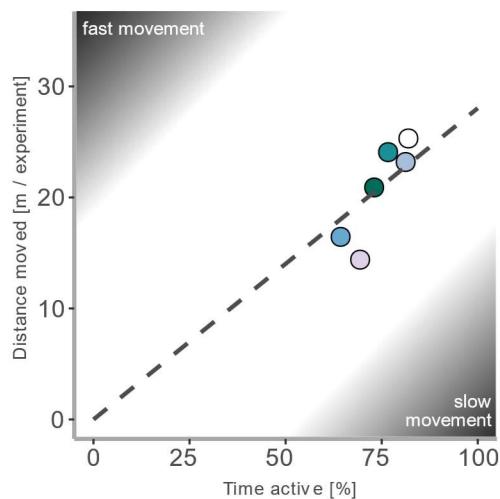
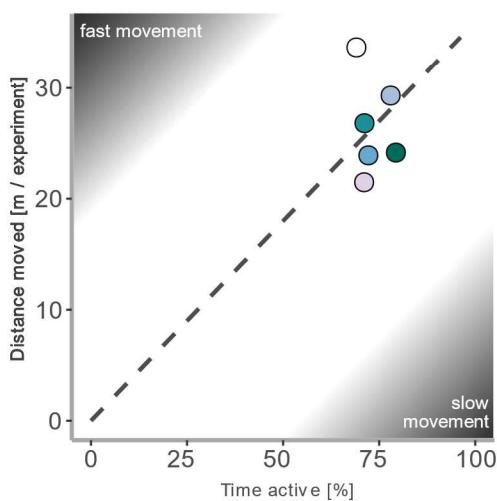
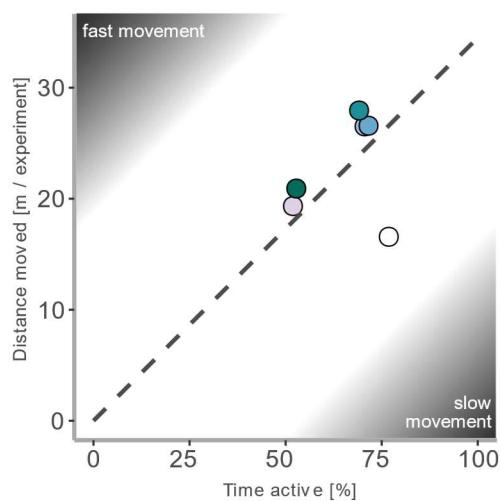
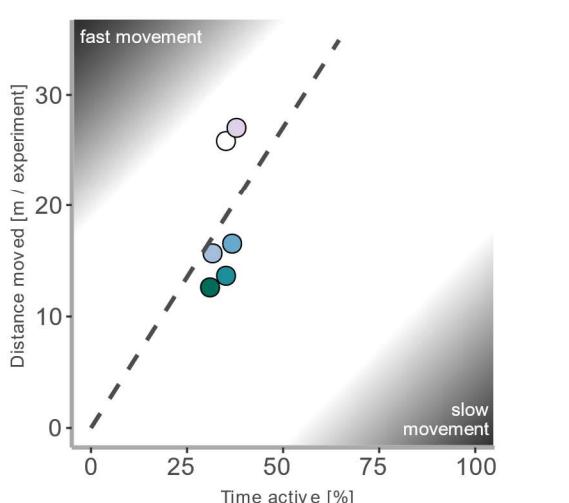
## Supplementary Material



*Figure S 1* Sex specific behavior of four amphipod species and infected *G. roeselii* given as total distance moved per experiment. Values are based on estimated marginal means. Error bars show the standard error. Asterisks indicate significant deviations between the sexes, based on Tukey's HSD post hoc test (\* $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ ).



*Figure S 2: Sex specific behavior four amphipod species and infected G. roeselii given as total distance moved per experiment. Values are based on estimated marginal means from single species GLMs. Error bars show the standard error. Asterisks indicate significant deviations between the sexes, based on Tukey's HSD post hoc test (\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ ).*

a) *G. fossarum*b) *G. pulex*c) *G. roeselii* (uninfected)d) *G. roeselii* (infected)e) *D. villosus*Concentration [ $\mu\text{g} / \text{L}$ ]

- 0
- 0.32
- 1.04
- 3.31
- 10.6
- 33.92

Figure S 3 Combining total distance moved and time spent active of the thiacloprid treatments for the different amphipod species used. The trend line, forced to intersect the y-axis at  $y = 0$ , illustrates the linear association between the two variables and indicates a shift from fast movement to slow movement.

Table S 1 Overview of individuals in each experiment series. Total numbers, numbers of females and males used, and sex-ratio within the species.

		total	female	male	Sex-ratio (male:female)
<i>G. fossarum</i>	NC	31	10	21	2.1
	0.32 µg/L	32	10	22	2.2
	1.04 µg/L	32	6	26	4.3
	3.31 µg/L	32	12	20	1.7
	10.6 µg/L	32	14	18	1.3
	33.92 µg/L	32	11	21	1.9
		<b>191</b>	<b>63</b>	<b>128</b>	<b>2.0</b>
<i>G. pulex</i>	NC	32	17	15	0.9
	0.32 µg/L	32	19	13	0.7
	1.04 µg/L	32	16	16	1.0
	3.31 µg/L	32	12	20	1.7
	10.6 µg/L	32	17	15	0.9
	33.92 µg/L	32	17	16	0.9
		<b>192</b>	<b>98</b>	<b>95</b>	<b>1.0</b>
<i>G. roeselii</i>	NC	32	24	8	0.3
	0.32 µg/L	33	17	16	0.9
	1.04 µg/L	32	17	15	0.9
	3.31 µg/L	32	17	15	0.9
	10.6 µg/L	32	19	13	0.7
	33.92 µg/L	32	20	12	0.6
		<b>193</b>	<b>114</b>	<b>79</b>	<b>0.7</b>
<i>G. roeselii</i> (infected)	NC	32	19	13	0.7
	0.32 µg/L	31	15	16	1.1
	1.04 µg/L	32	21	11	0.5
	3.31 µg/L	32	21	11	0.5
	10.6 µg/L	32	16	6	0.4
	33.92 µg/L	32	23	9	0.4
		<b>191</b>	<b>115</b>	<b>66</b>	<b>0.6</b>
		total	<i>P. laevis</i>	<i>P. minutus</i>	Mean count
		<b>596</b>	<b>554</b>	<b>42</b>	<b>3.1</b>
<i>D. villosus</i>	NC	32	15	17	1.1
	0.32 µg/L	32	14	18	1.3
	1.04 µg/L	32	15	17	1.1
	3.31 µg/L	31	17	14	0.8
	10.6 µg/L	32	10	22	2.2
	33.92 µg/L	32	16	16	1.0
		<b>191</b>	<b>87</b>	<b>104</b>	<b>1.2</b>
	total	958	477	472	1.2

Table S 2 Results of independent GLMs of the four local amphipod species and infected *G. roeselii*. Endpoints used were total distance moved [m] (TDM) and time active [%] (TA). Significant results are highlighted in bold.

		<i>G. fossarum</i>			<i>G. pulex</i>			<i>G. roeselii</i>			<i>G. roeselii</i> (infected)			<i>D. villosus</i>		
Dependent variable	Independent variables	Df	F value	P	Df	F value	P	Df	F value	P	Df	F value	P	Df	F value	P
a) Total distance moved [m]	concentration	5	<b>7.19</b>	<b>0.000</b>	5	2.30	<b>0.046</b>	5	1.60	0.162	5	2.10	0.068	5	<b>3.77</b>	<b>0.003</b>
	sex	1	<b>10.88</b>	<b>0.001</b>	1	10.06	<b>0.002</b>	1	<b>8.01</b>	<b>0.005</b>	1	0.49	0.486	1	1.64	0.202
	size	1	0.24	0.626	1	0.88	0.349	1	0.05	0.820	1	0.12	0.726	1	2.98	0.086
	concentration:sex	5	1.26	0.285	5	0.55	0.739	5	1.20	0.311	5	2.13	0.064	5	0.90	0.483
	Residuals	178			179			180			178			178		
b) Time active [%]	concentration	5	<b>8.23</b>	<b>0.000</b>	5	2.59	<b>0.027</b>	5	0.65	0.662	5	<b>3.07</b>	<b>0.011</b>	5	0.34	0.888
	sex	1	0.24	0.624	1	1.84	0.177	1	1.48	0.225	1	0.24	0.623	1	1.20	0.275
	size	1	0.13	0.720	1	2.55	0.112	1	0.21	0.647	1	2.20	0.140	1	1.08	0.301
	concentration:sex	5	1.91	0.095	5	0.91	0.475	5	1.65	0.149	5	1.63	0.154	5	0.90	0.484
	Residuals	178			179			181			177			178		

Table S 3 Results of pairwise comparison from Tukey's HSD post hoc tests test for each amphipod species and infected G. roeselii.

TDM [m]	contrast	estimate	SE	df	t.ratio	p.value	TA [%]	contrast	estimate	SE	df	t.ratio	p.value
<i>G. fossarum</i>	NC - 0.32 µg/L	<b>-10.82</b>	<b>2.92</b>	<b>178</b>	<b>-3.70</b>	<b>0.004</b>	<i>G. fossarum</i>	NC - 0.32 µg/L	<b>-25.31</b>	<b>7.37</b>	<b>178</b>	<b>-3.43</b>	<b>0.010</b>
	NC - 1.04 µg/L	-0.36	3.19	178	-0.11	1.000		NC - 1.04 µg/L	<b>-24.28</b>	<b>8.06</b>	<b>178</b>	<b>-3.01</b>	<b>0.035</b>
	NC - 3.31 µg/L	-3.88	2.86	178	-1.35	0.754		NC - 3.31 µg/L	<b>-28.43</b>	<b>7.22</b>	<b>178</b>	<b>-3.94</b>	<b>0.002</b>
	NC - 10.6 µg/L	-1.58	2.85	178	-0.56	0.994		NC - 10.6 µg/L	-11.70	7.18	178	-1.63	0.581
	NC - 33.92 µg/L	4.88	2.90	178	1.68	0.545		NC - 33.92 µg/L	7.85	7.32	178	1.07	0.892
<i>G. pulex</i>	NC - 0.32 µg/L	10.92	4.04	179	2.70	0.080	<i>G. pulex</i>	NC - 0.32 µg/L	12.56	6.52	179	1.93	0.389
	NC - 1.04 µg/L	2.12	3.95	179	0.54	0.995		NC - 1.04 µg/L	0.71	6.37	179	0.11	1.000
	NC - 3.31 µg/L	8.87	4.08	179	2.17	0.257		NC - 3.31 µg/L	17.63	6.58	179	2.68	0.085
	NC - 10.6 µg/L	1.23	3.97	179	0.31	1.000		NC - 10.6 µg/L	5.28	6.40	179	0.83	0.962
	NC - 33.92 µg/L	4.41	3.98	179	1.11	0.877		NC - 33.92 µg/L	8.91	6.41	179	1.39	0.734
<i>G. roeselii</i>	NC - 0.32 µg/L	12.23	4.79	180	2.55	0.115	<i>G. roeselii</i> (infected)	NC - 0.32 µg/L	-2.01	7.32	181	-0.27	1.000
	NC - 1.04 µg/L	4.41	4.83	180	0.91	0.943		NC - 1.04 µg/L	-8.91	7.36	181	-1.21	0.832
	NC - 3.31 µg/L	9.80	4.83	180	2.03	0.330		NC - 3.31 µg/L	-3.11	7.37	181	-0.42	0.998
	NC - 10.6 µg/L	6.90	4.86	180	1.42	0.714		NC - 10.6 µg/L	-2.10	7.38	181	-0.28	1.000
	NC - 33.92 µg/L	9.55	4.91	180	1.94	0.379		NC - 33.92 µg/L	-10.28	7.50	181	-1.37	0.745
<i>G. roeselii</i> (infected)	NC - 0.32 µg/L	-2.76	3.52	178	-0.79	0.970	<i>G. roeselii</i>	<b>NC - 0.32 µg/L</b>	<b>24.94</b>	<b>7.06</b>	<b>177</b>	<b>3.53</b>	<b>0.007</b>
	NC - 1.04 µg/L	-9.94	3.58	178	-2.77	0.066		NC - 1.04 µg/L	6.30	7.19	177	0.88	0.952
	NC - 3.31 µg/L	-10.00	3.59	178	-2.78	0.065		NC - 3.31 µg/L	5.27	7.21	177	0.73	0.978
	NC - 10.6 µg/L	-11.38	4.01	178	-2.84	0.056		NC - 10.6 µg/L	7.72	8.06	177	0.96	0.930
	NC - 33.92 µg/L	-4.35	3.82	178	-1.14	0.865		<b>NC - 33.92 µg/L</b>	<b>24.07</b>	<b>7.67</b>	<b>177</b>	<b>3.14</b>	<b>0.024</b>
<i>D. villosus</i>	NC - 0.32 µg/L	-1.19	4.52	178	-0.26	1.000	<i>D. villosus</i>	NC - 0.32 µg/L	-2.75	6.74	178	-0.41	0.999
	NC - 1.04 µg/L	10.18	4.49	178	2.27	0.212		NC - 1.04 µg/L	3.48	6.69	178	0.52	0.995
	NC - 3.31 µg/L	9.32	4.53	178	2.06	0.316		NC - 3.31 µg/L	-1.62	6.76	178	-0.24	1.000
	NC - 10.6 µg/L	12.20	4.67	178	2.61	0.099		NC - 10.6 µg/L	-0.02	6.96	178	0.00	1.000
	<b>NC - 33.92 µg/L</b>	<b>13.24</b>	<b>4.49</b>	<b>178</b>	<b>2.95</b>	<b>0.042</b>		NC - 33.92 µg/L	4.14	6.70	178	0.62	0.990

*Table S 4 CRED reporting recommendations, to be used together with the accompanying guidance (Moermond et al., 2016). Overview table that gives targeted information about the method used and the results.*

Number		Reported Yes/No	Reported in section	Comments
1	General information			
a	Purpose of study	Yes	Intro	
b	Description of endpoints	Yes	Intro	
2	Test setup			
a	Performed according to standard/modified standard (e.g., OECDa, US EPAb)	No		non-standard study
b	Performed according to Good Laboratory Practices (GLP)	No		
c	Description of used control(s): negative control, solvent control, positive control	Yes	2.4 Experimental setup	
d	Control(s) mortality, growth, morbidity and other observed non-standard effects like behavior and coloring			Whole article
e	Comparison to validity criteria (e.g. control survival, growth) from appropriate standard test method	No		Need to be established
3	Test compound			
a	Identification (e.g. name, CAS-number, specify if the salt or the base is tested)	Yes	2.5 Test substances	
b	Physico-chemical characteristics that may influence the behavior of the compound during the study No data (e.g. solubility, volatility, stability (hydrolysise, photolysise, degradation), solubility, log KOW, degradability, adsorption)			Not provided due to shortening of the exposure period
c	Source	Yes	2.5 Test substances	
d	Purity in %	Yes	2.5 Test substances	
e	Composition of product formulation, and presence of impurities			
4	Test organism			
a	Scientific name	Yes	2.1 Study species	<i>Gammarus fossarum</i> , <i>Gammarus pulex</i> , <i>G. roeselii</i> (plus potential acanthocephalan parasites), <i>Dikerogammarus villosus</i>
b	Body weight, length	Yes	length	Supplemental Dataset

c	Age/life-stage	Yes	2.1 Study species	only mature individuals with fully developed sexual characteristics were used
d	Growth/reproductive condition	No		
e	Sex	Yes	2.4 Experimental setup; Table S 1	including Sex-ratio m:f
f	Strain, clone	Yes	2.2 Molecular identification	species field individuals
g	Source, including possible pre-exposure for field-collected species	Yes	2.1 Study species;	4. WWTP, agricultural fields Discussion
h	Culture handling	Yes	2.3 Sampling sites and culture handling	
5	Exposure conditions			
a	Exposure schedule (static, semi static, flow through system, other) and flow-rate (flow-through systems) or renewal time (semi-static systems)	Yes	2.4 Experimental setup; previous study: Soose et al. (2023)	
b	Open or closed system	Yes	2.4 Experimental setup; previous study: Soose et al. (2023)	
c	Test medium composition and source of test water (e.g., well water, deionised water, tap water)	Yes	2.3 Sampling sites and culture handling; 2.4 Experimental setup	
d	Temperature, and time-points for measuring	Yes	2.4 Experimental setup	
e	pH, and time-points for measuring	No	see Table S 5	
f	Hardness of water, and time-points for measuring	No	see Table S 5	
g	Conductivity, and time-points for measuring	No	see Table S 5	
h	Dissolved oxygen content, and time-points for measuring	No	see Table S 5	
i	Light intensity and quality (source and homogeneity), light/dark conditions	Yes	2.4 Experimental setup	

j	Feeding protocols, food composition	Yes	2.3 Sampling sites and culture handling
k	Material and volume of aquarium/container and other equipment in contact with test organisms and test substance	Yes	2.4 experimental setup
l	Use of sand or sediment, and its characteristics (total organic carbon (TOC), particle size, etc.)	No	
m	Preparation of stock solutions, including solvent concentrations in test water and controls	Yes	2.5 Test substance
n	Nominal concentrations of test substance	Yes	2.5 Test substance
o	Measured concentrations of test substance, and time-points for measuring	Yes	2.5 Test substance
p	Analytical method: description of method, including limit of detection (LOD), limit of quantification (LOQ)	Yes	2.5 Test substance
q	Exposure duration and total test duration	Yes	2.4 Experimental setup
r	Time-points of observations for endpoints	Yes	2.4 Experimental setup
s	Results based on nominal or measured concentrations	Yes	3. Results nominal
t	Biomass loading (biomass per liter)	No	
6	Statistical Design and Biological Response		
a	Number of replicates for control(s) and test concentrations; setup of replicates (avoid pseudo-replication)	Yes	2.4 Experimental setup
b	Number of organisms, or algal cell concentration, per replicate	Yes	2.4 Experimental setup; Table S 1
c	Treatment design (e.g., block, randomized)	Yes	2.4 Experimental setup randomly chosen
d	Statistical method used	Yes	2.7 Statistical analyses
e	Biological response for each concentration	Yes	
f	Dose-response observed	Yes	
g	Statistically significant responses noted (e.g. ECx)	Yes	3 Results - Behavioral response to chemical exposure; Figure 2
h	Significance level for NOEC and LOEC data (0.05 or less)	Not applicable	
i	Estimation of variability for LCx and ECx data	Not applicable	
j	Availability of raw data: through supplementary information, a website, or upon request.	Yes	

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a OECD = Organisation for Economic Co-operation and Development; b US EPA = The United States Environmental Protection Agency

## References

- Moermond CT, Kase R, Korkaric M, Ågerstrand M. CRED: Criteria for reporting and evaluating ecotoxicity data. *Environ. Toxicol. Chem.* 2016; 35: 1297-1309.
- Soose, L.J., Hügl, K.S., Oehlmann, J., Schiwy, A., Hollert, H., Jourdan, J., 2023. A novel approach for the assessment of invertebrate behavior and its use in behavioral ecotoxicology. *Science of The Total Environment* 897, 165418. <https://doi.org/10.1016/j.scitotenv.2023.165418>

**Table S 5 Water parameters of medium**

*Table S 5 Measured parameters of test medium in climate controlled room at animal holding facility*

<b>Medium <i>G. fossarum</i></b>		<b>Medium <i>G. pulex</i>, <i>G. roeselii</i>, <i>D. villosus</i></b>
pH	6.6	8.2
Conductivity	254 µS	836 µS
Total hardness	4.2°dH; 80 mg/L CaCO <sub>3</sub>	11° dH; 200 mg/L CaCO <sub>3</sub>
Carbonate hardness	0.5°dH; 0.2mmol/L SBV; 12.2 mg/L HCO <sub>3</sub>	8° dH; 3.3 mmol/L SBV; 201.36 mg/L HCO <sub>3</sub>
O <sub>2</sub>	9.5 mg/L; 93.8 % at 13°C	9.89 mg/L; 96.2% at 13°C